11-24-2010

Maturation-dependent licensing of naive T cells for rapid TNF production

Bhavana Priyadharshini
University of Massachusetts Medical School

Raymond M. Welsh
University of Massachusetts Medical School

Dale L. Greiner
University of Massachusetts Medical School

See next page for additional authors

Follow this and additional works at: http://escholarship.umassmed.edu/oapubs

Part of the Immunology and Infectious Disease Commons, and the Medicine and Health Sciences Commons

Repository Citation
Priyadharshini, Bhavana; Welsh, Raymond M.; Greiner, Dale L.; Gerstein, Rachel M.; and Brehm, Michael A., "Maturation-dependent licensing of naive T cells for rapid TNF production" (2010). Open Access Articles. 2361.
http://escholarship.umassmed.edu/oapubs/2361

This material is brought to you by eScholarship@UMMS. It has been accepted for inclusion in Open Access Articles by an authorized administrator of eScholarship@UMMS. For more information, please contact Lisa.Palmer@umassmed.edu.
Maturation-dependent licensing of naive T cells for rapid TNF production

Authors
Bhavana Priyadharshini, Raymond M. Welsh, Dale L. Greiner, Rachel M. Gerstein, and Michael A. Brehm

Rights and Permissions
Copyright: © 2010 Priyadharshini et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

This article is available at eScholarship@UMMS: http://escholarship.umassmed.edu/oapubs/2361
Maturation-Dependent Licensing of Naive T Cells for Rapid TNF Production

Bhavana Priyadharshini1,2, Raymond M. Welsh1,2, Dale L. Greiner2,3,4, Rachel M. Gerstein2,5, Michael A. Brehm2,3,4,*

1 Department of Pathology, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, 2 Program in Immunology and Virology, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, 3 Program in Molecular Medicine, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, 4 Diabetes Center of Excellence, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America, 5 Department of Molecular Genetics & Microbiology, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America

Abstract

The peripheral naïve T cell pool is comprised of a heterogeneous population of cells at various stages of development, which is a process that begins in the thymus and is completed after a post-thymic maturation phase in the periphery. One hallmark of naïve T cells in secondary lymphoid organs is their unique ability to produce TNF rapidly after activation and prior to acquiring other effector functions. To determine how maturation influences the licensing of naïve T cells to produce TNF, we compared cytokine profiles of CD4+ and CD8+ single positive (SP) thymocytes, recent thymic emigrants (RTEs) and mature-naïve (MN) T cells during TCR activation. SP thymocytes exhibited a poor ability to produce TNF when compared to T cells despite expressing similar TCR levels and possessing comparable activation kinetics (upregulation of CD25 and CD69). Provision of optimal antigen presenting cells from the spleen did not fully enable SP thymocytes to produce TNF, suggesting an intrinsic defect in their ability to produce TNF efficiently. Using a thymocyte adoptive transfer model, we demonstrate that the ability of T cells to produce TNF increases progressively with time in the periphery as a function of their maturation state. RTEs that were identified in NG-BAC transgenic mice by the expression of GFP showed a significantly enhanced ability to express TNF relative to SP thymocytes but not to the extent of fully MN T cells. Together, these findings suggest that TNF expression by naïve T cells is regulated via a gradual licensing process that requires functional maturation in peripheral lymphoid organs.


Editor: Jacques Zimmer, Centre de Recherche Public de la Santé (CRP-Santé), Luxembourg

Received September 14, 2010; Accepted October 10, 2010; Published November 24, 2010

Copyright: © 2010 Priyadharshini et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported in part by National Institutes of Health grants A1017672 and A081675 to R.M.W., AI46629 to D.L.G., AI43534 to R.M.G., A1083911 to M.A.B., and an institutional Center for AIDS Research (CFAR) grant AI042845 and grants from the Juvenile Diabetes Research Foundation. R.M.W., D.L.G. and R.M.G. are members of the UMass Diabetes Endocrinology Research Center (DERC) grant DK32520. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: michael.brehm@umassmed.edu

Introduction

T cell development begins within the thymus and is driven to completion after single positive (SP) thymocytes exit the thymus and seed secondary lymphoid organs, where they undergo progressive phenotypic and functional maturation [1]. The peripheral naïve T cell pool is therefore comprised of a heterogeneous population of cells at different stages of post-thymic development, encompassing T cell subsets from the fully mature to the most recently emigrated thymic T cells [2]. The recent thymic emigrants (RTEs), which are 0–2 weeks old in the periphery have a distinct phenotypic profile (CD24high, Qa2low, CD43RBlow) relative to their mature naïve (MN) counterparts, that are resident in the periphery for >3 weeks (CD24low, Qa2high, CD43RBhigh) [1,3]. RTEs have been shown to also differ functionally, producing less IL-2, exhibiting a decreased ability to proliferate upon 48 hours of in vitro TCR stimulation and producing less IFN-γ after 7 days of infection with ovalbumin-expressing Listeria monocytogenes (Lm-OVA) [1,3]. Resting naïve T cells in secondary lymphoid organs are quiescent in nature requiring a low level of TCR signaling from self peptide-MHC ligands to maintain immune homeostasis [4]. Upon antigen-specific activation, naïve T cells differentiate and clonally expand to become effectors that are capable of secreting cytokines (IL-2, TNF and IFN-γ) and exhibiting cytolytic function [5,6,7]. In contrast to this conventional paradigm, naïve CD4+ and CD8+ T cells (CD44lo, CD11alo) have recently been shown to rapidly produce TNF within 4 to 5 hours of TCR engagement, before ensuing cell division or producing other effector cytokines such as IL-2 or IFN-γ [8,9]. The kinetics of TNF production by naïve T cells suggest that this potent immunomodulatory cytokine is released during the initial encounter between T cells and APCs, a critical phase in the programming of antigen-specific responses [5,10,11,12,13,14,15,16]. However, when and how naïve T cells acquire this unique capability to produce TNF during development is not known.

TNF is a potent pro-inflammatory cytokine that elicits pleiotropic effects during an immune response, affecting immune cell activation, survival, death and differentiation [17,18]. The effects of TNF are mediated through two distinct receptors,
peptide (data not shown). To determine if the reduced TNF production was not overcome by increasing the concentrations of the stimulatory splenic T cells during TCR stimulation. This inability to produce TNF cannot be overcome by increasing the concentrations of the stimulatory splenic T cells during TCR stimulation. This inability to produce TNF is not known at what stage of development naïve T cells acquire the capability of producing TNF. To determine this, thymocytes and splenocytes from CD8+ T lymphocytes (CD44lo) from secondary lymphoid organs rapidly produce TNF after TCR engagement before gaining other effector functions [8]. However, it is not known at what stage of development naïve T cells acquire the ability to produce TNF. To determine this, thymocytes and splenocytes from CD8+ and CD4+ TCR-transgenic mice (P14, OT-1 & SMARTA and OT-2) were stimulated with specific antigens. These studies reveal that CD4+ CD8− and CD4− CD8+ SP thymocytes possess a poor ability to produce TNF upon stimulation when compared to their counterparts in secondary lymphoid organs. Contact with secondary lymphoid cells (spleen and lymph nodes) during TCR activation partially enables SP thymocytes to produce TNF in vitro by providing optimal antigen-presentation. However, the frequency of TNF producing cells is still significantly lower than in the periphery. RTEs in the spleen on the other hand, display an intermediate TNF response, which is higher than their SP thymic precursors but lower relative to the fully mature T cells. The differences in the TNF profile exhibited by these 3 populations of lymphocytes mirrors their distinctive maturation status. Moreover, as developing T cells mature in the periphery, they show a progressive increase in their capability to produce TNF upon TCR activation. Together, these findings suggest that naïve T cells become gradually licensed to efficiently produce TNF in a maturation-dependent manner that requires their localization to secondary lymphoid organs.

Results

SP thymocytes have an impaired ability to produce TNF after TCR activation

Naive CD4+ and CD8+ T lymphocytes (CD44lo) from secondary lymphoid organs rapidly produce TNF after TCR engagement before gaining other effector functions [8]. However, it is not known at what stage of development naïve T cells acquire the ability to produce TNF. To determine this, thymocytes and splenocytes from CD8+ and CD4+ TCR-transgenic mice (P14, OT-1 & SMARTA and OT-2) were stimulated with specific peptides and αCD28 costimulation for 4 hrs in vitro. Fig. 1A shows that a lower proportion of CD4+ CD8− and CD4− CD8+ SP thymocytes produced TNF when compared to naïve (CD44lo) splenic T cells during TCR stimulation. This inability to produce TNF was not overcome by increasing the concentrations of the peptide (data not shown). To determine if the reduced TNF response by SP thymocytes was due to a lower TCR expression on SP thymocytes relative to naïve splenic T cells [40,41,42], SP P14-CD80 thymocytes and naïve (CD44lo) splenic T cells were stained with mAbs to TCR Vα2 and TCR Vβ8.1. Fig. 1B shows that Vα2 and Vβ8.1 expression in SP P14-CD80 thymocytes and splenetic T cells were similar. Next, to determine if the reduced ability of SP thymocytes to produce TNF was due to a generalized defect in their activation, we examined the expression of activation markers CD25, CD69, CD44 and CD62L on the P14-CD80 SP thymocytes and splenic T cells. As shown in Fig. 1C, SP thymocytes and splenic T cells exhibited a comparable level of activation at 4 hours, with the expression of CD25 and CD69 being up-regulated and the expression of CD62L down-regulated, as previously shown [8,43]. These results suggest that SP thymocytes are incompetent to produce TNF when compared to splenic T cells upon TCR stimulation despite exhibiting similar TCR levels and similar phenotypic changes in the expression of activation markers.

Optimal antigen presentation is not sufficient to enable thymocytes to produce TNF efficiently

Recent studies have shown that thymic DCs and splenic DCs have unique properties and that the microenvironment contributes to their distinct functions [44]. We hypothesized that differences in TNF production between SP thymocytes and naïve splenic T cells may be attributed to the differences in antigen presentation between the two organs. To test this, enriched CD451.1 SP P14-CD80 thymocytes were stimulated in the presence of either WT CD451.2 H-2Db-positive B6 splenocytes or CD451.2 H-2Db-deficient B6 splenocytes (incapable of presenting GP33 to P14 cells). In comparison to the high frequencies of TNF-producing splenocytes depicted in Fig. 1A, there was only a partial increase in the proportion of SP P14-CD80 thymocytes that produced TNF, when stimulated in the presence of WT B6 splenocytes (Fig. 1D). Both purified splenic B cells and non-B and -T cell populations that contained CD11c+ DCs in the flow-through stimulated this small increase in TNF production (21.4% and 17.3% of SP P14 CD80 thymocytes were TNF positive when stimulated purified B cells and non-B and -T cell populations, respectively). We then determined whether SP P14-CD80 thymocytes produced TNF in a consequence of changes occurring in antigen-presenting cells during the co-culture, by using irradiated splenocytes. We found that SP P14-CD80 thymocytes stimulated with irradiated (3000cGy) WT B6 splenocytes showed a similar increase in the proportion of TNF-producing cells comparable to SP P14-CD80 thymocytes stimulated with live WT B6 splenocytes (Fig. 1D), indicating that viable splenocytes were not necessary for this effect. As expected, thymocytes stimulated in the presence of H2Db-deficient splenocytes (live or irradiated) did not produce TNF. Together, these results indicate that although optimal TCR-MHC interactions provided by spleen cells enable a small subset of SP thymocytes to produce TNF, the frequency of cells and the level of TNF produced on a per cell basis was reduced compared to peripheral naïve CD8+ splenic T cells, suggesting an intrinsic defect in SP thymocytes to produce TNF efficiently upon TCR activation.

Lower level of TNF transcription in SP thymocytes relative to naïve splenic T cells during TCR activation

We wanted to determine if the reduced ability of thymocytes to produce TNF may be related to the levels of mature TNF message expressed within SP thymocytes. Therefore, we quantified the steady-state levels of mature TNF mRNA in SP thymocytes
Maturation Determines TNF Production by T Cells

A. Transgenic CD8+ and CD4+ T Cells

Thymus

Spleen

B. TCR Vα2 and TCR Vβ8.1

Thymus

Spleen

C. CD25, CD69, CD62L, CD44, TNF

Thymus

Spleen

Unstimulated

GP33+αCD28

D. SP Thymocytes

+H-2Db positive splenocytes

Live

Irradiated

SP Thymocytes

+H-2Db KO splenocytes

TNF

CD44
CD4^+ CD8^+ and their respective naive splenic (CD44^hi) counterparts purified from (transgenic) P14 mice at resting state. SP P14-CD8^+ thymocytes were sorted to 98% purity and (CD44^hi) P14-CD8^+ splenic T cells were sorted to 90.3% purity. Fig. 2A shows the copy number of TNF transcripts detected in the indicated groups by quantitative real-time PCR. The levels of mature TNF message in transgenic SP thymocytes (CD4^+ CD8^+) and their naive splenic counterparts were similar and the differences were not significant (Fig. 2A). Next we compared the levels of TNF transcripts in purified SP P14-CD8^+ thymocytes and naive (CD44^hi) splenic T cells that were stimulated as indicated. We found that the levels of TNF mRNA were dramatically higher in (CD44^hi) P14-CD8^+ splenic T cells during GP33 and GP33+CD28 stimulation relative to SP thymocytes. The levels of TNF transcripts increased in the thymic subsets upon stimulation but not to the extent detected in the splenic subset (Fig. 2B). Together, these results indicate that despite having a basal level of transcription of the TNF gene, SP thymocytes appear to lack the ability to induce TNF transcription efficiently upon stimulation relative to naive splenic T cells.

**Differential ability of SP thymocytes and naive splenic T cells to produce TNF during TCR activation in vitro**

To first determine if naive splenic T cells produced TNF in the presence of physiologically relevant levels of antigen, we performed an in vivo cytokine assay [45]. Briefly, CD45.1^+ P14-CD8^+ and CD45.1^+ SMARTA-CD4^+ TCR-transgenic splenic T cells were treated in vitro with biA, a golgi transport inhibitor that blocks cytokine secretion. These cells were then mixed and co-transferred into recipients that were infected with either WT LCMV-Armstrong or a GP33-CTL escape variant of LCMV (GP/IV) for 2 days. The spleens of recipient mice were recovered 4 hours later and directly stained for intracellular TNF by the donor T cells. Both P14-CD8^+ and SMARTA-CD4^+ donor T cells produced TNF in mice that were infected with WT LCMV-Armstrong (Fig. 3A). Only T cells that had down-regulated CD62L, which is consistent with a TCR-mediated activation event, were able to produce TNF. In contrast, P14-CD8^+ donor T cells in mice that were infected with GP1V mutant virus were impaired in their ability to produce TNF (Fig. 3A) while SMARTA-CD4^+ donor T cells were unaffected. These findings indicate that the in vivo production of TNF by naive CD8^+ T cells during LCMV infection was specifically initiated by TCR-mediated signaling and was not due to non-specific effects on naive T cells by virus-induced inflammation.

We next examined the ability of SP P14-CD8^+ thymocytes and SMARTA-CD4^+ thymocytes to produce TNF in the same scenario. A small but reproducible proportion of both donor SP P14-CD8^+ and SMARTA-CD4^+ thymocytes produced TNF (4 to 6% TNF positive of total T cells) during LCMV infection (Fig. 3B). As expected, infection with GP1V mutant impaired the ability of the donor SP P14-CD8^+ thymocytes to produce TNF. Fig. 3C shows the average percentages of TNF producing SP P14-CD8^+ donor thymocytes and splenic T cells that had downregulated their CD62L expression under the indicated conditions. Together, these results confirm our in vitro data indicating that SP thymocytes are impaired in their ability to produce TNF efficiently when compared to naive splenic T cells during a viral infection.

**TNF producing capability of SP thymocytes correlates with their maturation state**

SP thymocytes are comprised of a heterogeneous population consisting of cells at different levels of maturity [46]. Immature SP thymocytes express high levels of CD24 (HSA), which is downregulated as cells progress into maturity [47]. This is accompanied by down-regulation of CD69 and the up-regulation of other markers such as CD62L, CD45RB and Qa2 [48,49,50]. We first compared the maturation profile of the total TNF-producing thymocytes with their TNF non-producing counterparts. We broadly classified SP P14-CD8^+ thymocytes based on their maturation status determined by CD24 and Qa2 expression. CD8^+ SP thymocytes were divided into 4 subgroups from the least mature to most mature (Fig 4A). Subgroup 1 was comprised of CD24^lo Qa2^lo cells (least mature) followed by subgroup 2 (CD24^lo Qa2^hi), subgroup 3 (CD24^hi Qa2^hi) which was the most mature [51]. The small population of SP P14-CD8^+ thymocytes that produced TNF displayed a more mature phenotypic profile with the majority of the TNF producers falling in subgroups 2 and 3 compared to the TNF non-producers that fell mostly in subgroups 1 and 2. The maturation differences between the TNF-producing SP thymocytes and the non-producers were also seen in the MFI changes in CD24, CD45RB and Qa2 (dotted line histograms and gray histograms in Fig 4C). However, the TNF producing SP P14-CD8^+ thymocytes had a less mature phenotype when compared to their splenic counterparts. As described in Fig 4A and Fig 4B, >60% of the TNF producing thymocytes constituted subgroups 2 and 3 relative to the TNF producing splenic T cells that constituted >80% in subgroups 3 and 4. The differences were also reflected in the MFI of maturation markers (dark line histograms and black histograms in Fig 4D). We next examined the TNF-producing capability of each of the 4 subgroups in the SP thymic subset individually. The subgroups showed increasing mean fluorescence intensities of CD45RB, consistent with their maturation state (Fig 4D). There was a progressive increase in TNF production on a per cell basis that correlated with maturation with Subgroup 4 having the highest percentage of TNF^+ cells. Together, these results suggest that although the small population of TNF producing SP P14-CD8^+ thymocytes is more mature than the TNF non-producing counterparts, these cells are still phenotypically less mature than P14-CD8^+ naïve T cells localized in the spleen.
Adaptively transferred transgenic SP thymocytes progressively gain the ability to produce TNF in the periphery

The differences in TNF production between SP P14-CD8α thymocytes and naive P14-CD8α splenic T cells upon TCR stimulation parallels the differences in the maturation status of T cells in these two compartments as shown in Fig 4. The functional maturation of developing T cells occurs progressively with time upon contact with secondary lymphoid organs after their exit from the thymus [1]. Given this, we hypothesized that SP thymocytes migrating into the periphery will gradually acquire the capability to produce TNF efficiently upon TCR stimulation. To recapitulate thymic emigration, $2 \times 10^6$ CD45.1+ P14 thymocytes were adoptively transferred into uninfected CD45.2+ B6 congenic mice. Spleens were harvested from recipient mice at the indicated time points (Fig 5: plots iii,iv,v,vi) and stained for donor CD45.1+ SP P14-CD8α thymocytes producing TNF upon in vitro TCR stimulation. The proportion of donor CD45.1+ SP P14-CD8α thymocytes producing TNF upon TCR stimulation increased over the time of the experiment (boxed quadrants in Fig 5: plots iii,iv,v,vi). The donor CD45.1+ SP P14-CD8α thymocytes capable of TNF production also exhibited an increasing maturation phenotype (down-regulation of CD24 and up-regulation of CD45RB and Qa2) that approached a level similar to that of splenic T cells by day 14 after transfer. While the recovery of donor cells diminished over time, as shown in Table 1, we also observed increases in the mean fluorescence intensity (MFI) of the TNF signal in naive (CD44+) donor CD45.1+ SP P14-CD8α thymocytes producing TNF from day 2 to day 14 after transfer (Table 1). This increase in expression of TNF on a per cell basis by donor SP CD45.1+ P14-CD8α thymocytes was significant ($p<0.05$) and was consistent with the increasing maturation phenotype observed at these time points (Fig 5 and Table 2). We next compared the changes in MFI of maturation markers in the TNF-producing and non-producing donor thymocytes at day 1 and 2 after transfer, as the TNF-negative populations were very small at later time points (Table 2). The TNF-producing cells were more mature, again suggesting that the changes in the maturation state of donor thymocytes correlated with increasing capability to produce TNF efficiently on a per cell basis. Pre-transfer stimulation of thymocytes ex vivo in the presence of CD45.2+ B6 splenocytes did not affect their maturation status. Together, these results suggest that the progressive maturation of transferred SP P14-CD8α thymocytes in the periphery positively influences their capability to competitively produce TNF upon TCR stimulation.

Post-thymic maturation of naturally emigrating polyclonal SP thymocytes licenses them to produce TNF efficiently in the periphery

Polyclonal naive CD4+ and CD8+ T lymphocytes (CD44+) from secondary lymphoid organs rapidly produce TNF after TCR engagement before gaining other effector functions [8]. However, it is not known if polyclonal SP thymocytes also lack the capability to produce TNF like their transgenic counterparts. To determine this, thymocytes and splenocytes from naive non-transgenic B6 mice were stimulated using both monoclonal αCD3 and αCD28 antibodies for 4 hrs in vitro, respectively. Fig. 6A shows that, similarly to the transgenic T cells, a lower proportion of polyclonal CD4+ CD8α and CD4+ CD8α SP thymocytes produced TNF when compared to naive (CD44+) splenic T cells during TCR stimulation (Fig 6A). This inability to produce TNF was not overcome upon receiving optimal signals from APCs of secondary lymphoid organs (spleen and lymph nodes) during TCR activation, suggesting that SP thymocytes...
Maturation Determines TNF Production by T Cells

A.

B.

C.
possess an intrinsic defect in their ability to produce TNF efficiently upon stimulation. SP thymocytes eventually gain full competence to produce TNF upon TCR stimulation as they undergo post-thymic maturation in the periphery and join the mature-naïve T cell pool in secondary lymphoid organs and this licensing for TNF production does not require homeostatic cell division (data not shown).

Several studies have shown that TNF is expressed in the thymus and have demonstrated that this TNF has a physiological role within the thymus. In situ hybridization studies revealed the localization of TNF mRNA to the cortical regions of the thymus during ontogeny, and studies by Giroir et al. showed that there was constitutive expression of TNF in thymic lymphocytes [52,53]. In vitro functional studies showed that TNF induced CD25 expression in developing CD117+CD25+ thymocytes in vitro [54]. TNF also induced apoptosis of CD4+CD8+ double negative thymocytes that coexpressed both TNFR1 and TNFR2 at low doses but stimulated proliferation at higher doses [55]. Interestingly, TNFR1/2 double KO mice exhibited thymic hypotrophy with an overall increase in total thymocytes but had a normal distribution of SP CD4+ and SP CD8+ T cell subsets due to absence of apoptosis in DN thymocytes [55]. Collectively, the dual role of TNF in the thymus appears paradoxical and may depend on the location and the quantity of its production at various stages of development.

Alternatively, there is also evidence showing the dangerous effects of deregulated production of TNF in the thymus. For instance, mice that over-express human TNF within the thymus exhibit thymic atrophy, which is primarily associated with premature apoptosis of double negative (DN2) developing thymocytes and diminished numbers of cortical thymic epithelial cells (cTECs) [56]. Mice infected with Trypanosoma Cruzi show severe thymocyte depletion of CD4+CD8+ DP thymocytes due to an exacerbated inflammatory reaction mediated by TNF [57]. Increased levels of TNF and IFN-γ message have been associated with increased thymocyte depletion and cortical depletion observed in the thymus of patients with Down-Syndrome (DS) [58,59]. It is proposed that this abnormality may be due to improper interactions between developing thymocytes and thymic stromal cells mediated by elevated levels of LFA-1 and ICAM-1 and an abnormal distribution of ICAM-1 in DS thymi that is then exacerbated by the expression of TNF and IFN-γ in DS thymi [60]. These reports suggest that overproduction of TNF in the thymus may be detrimental to the T cell development process. We detected that resting unstimulated SP thymocytes expressed a small level of mature TNF message similar to their splenic counterparts. However, there was no spontaneous production of TNF protein detected in these cells. This suggests that despite having a similar basal level of TNF transcription, there is a lack of translation of the TNF protein under resting conditions in both these subsets. Therefore, the reduced ability of SP thymocytes to rapidly produce TNF during TCR engagement in the thymus under normal circumstances may be beneficial for the survival of SP thymocytes during T cell selection.

A previous report revealed that TNF production by T cells does not require de novo mRNA expression from the TNF locus, as primary CD4 T cells contain a premature TNF transcript which, following TCR engagement, is spliced to form a mature TNF message, resulting in the synthesis of TNF protein in the absence of new transcription [61]. However, we have previously observed that treatment of naïve CD8+ T cells with the transcriptional inhibitor actinomycin-D completely abrogated the production of TNF, suggesting that transcription is vital for TNF protein synthesis by naïve CD8+ T cells [8]. Our results here show that there is a slight but significant upregulation of TNF message in the CD8+ SP thymocytes upon TCR stimulation, but this is insufficient for the optimal production of protein. The splenic T cells, on the other hand, show a dramatic upregulation in TNF message and protein upon activation. Work done in Jurkat T cells and macrophages has revealed that the AU rich regions of the 3’UTR of TNF mRNA is vital for TNF regulation [62]. These findings suggest that there may either be a distinct transcriptional or post-transcriptional control of TNF gene expression in SP thymocytes relative to splenic T cells.

Stimulation in the presence of splenocytes partially enabled SP thymocytes to produce TNF efficiently as there was only a partial increase in the proportion of cells producing the cytokine. This effect may be either due to better antigen presentation of exogenous peptides by splenocytes to SP thymocytes. For example, secondary lymphoid cells (spleen and lymph nodes) might offer additional accessory signals leading to enhanced clustering of adhesion molecules during immunological synapse formation and to the partial production of TNF [63,64]. The alternative possibility is that there are more antigen presenting cells in the splenic environment that increase the strength of TCR-MHC interactions making SP thymocytes more sensitive to stimulation. We are currently exploring these possibilities. Moreover, this effect occurs regardless of whether SP thymocytes are stimulated in the presence of live or killed (γ-irradiated) splenocytes, suggesting that the peptide-MHC complexes and other potential molecules present on the surface of splenocytes are sufficient for this process. Nonetheless, thymocytes are still incapable of producing producing TNF efficiently despite receiving optimal antigen presentation in the splenic environment.

Contact with secondary lymphoid organs is vital for the completion of post-thymic maturation of developing SP thymocytes [1]. RTEs that reach the periphery undergo phenotypic and functional maturation as they become resident in secondary lymphoid organs [3,65]. Our findings indicate that the full competence for TNF production by naive T cells in the peripheral T cell pool is acquired gradually in a post-thymic maturation dependent manner. One caveat for the adoptive transfer model is that the recovery of thymocytes from the spleens of the recipient mice decreased over time. Therefore, we cannot exclude the possibility that the increase in the percentage of TNF producing donor CD45.1+ SP P14-CD8+ thymocytes with time in the periphery may be due to preferential survival of SP thymocytes that are capable of TNF production. Nevertheless, the hierarchical
Figure 4. TNF producing SP thymocytes exhibit a lower maturation profile relative to their splenic counterparts. A and B, CD45.1\(^+\) P14-CD8\(^+\) thymocytes were stimulated with GP33\(^a\)CD28 for 4 hours in vitro and then stained for maturation markers and intracellular TNF, as described in Materials and Methods. The TNF producers and non producers of the thymic and the splenic subsets were each classified into 4 subgroups based on their CD24 and Qa2 expression as shown namely Subgroup 1 (CD24\(^{hi}\) Qa2\(^{lo}\)) followed by Subgroup 2 (CD24\(^{hi-int}\) Qa2\(^{lo}\)), and so on.
pattern observed in the MFI of the TNF signal of transferred thymocytes and polyclonal RTEs clearly indicates that the progressive gain in the TNF producing capability of naïve T cells occurs as they mature in the periphery.

In mice, developing thymocytes emigrate and populate the periphery at the rate of 1–2% of thymocytes per day throughout the life [66,67]. Therefore, at any given time in an adult immune system, the naïve T cell pool is comprised of cells at various stages of post-thymic development, unlike neonates whose peripheral lymphoid organs are predominantly populated with RTEs [2,65]. The post-thymic maturation status of T cells is a component that has been recently shown to influence T cell fate decisions at the time of antigen encounter [2]. This study showed that RTEs produced fewer memory-precursor effector cells (MPECs) and more short-lived effector cells (SLECS) during the immune response to LCMV. Our data show that RTEs produce less TNF relative to MN T cells. Given the immunoregulatory functions of TNF, we speculate that the differential ability of TNF production linked to the post-thymic maturation status of antigen-specific naïve T cells, may also contribute to influencing the fate of the responding T cells during the initial phase of activation.

In conclusion, our findings indicate that the licensing of naïve T cells for rapid TNF production is determined by their developmental state. It is an intrinsic property of the developing T cells that is acquired gradually, where functional maturation in secondary lymphoid organs drives developing naïve T cells to eventually attain full competence to produce TNF efficiently during TCR stimulation.

Materials and Methods

Ethics Statement

All the experiments with animals were done in compliance with the institutional guidelines as approved by the University of Massachusetts Institutional Animal Care and Use Committee (IACUC). Animals were maintained in accordance with the Guide for the Care and Use of Laboratory Animals (Institute of Laboratory Animal Resources, 1996).

Mice

Male and female CD45.2+ C57BL/6J (B6) mice were purchased from The Jackson Laboratory (Bar Harbor, ME) and used at 6–12 weeks of age. CD45.1+ P14 CD8+ TCR-transgenic mice, with T cells that recognize the D3-restricted, LCMV epitope GP33-41 [68,69], and CD45.1+ SMARTA CD4+ TCR-transgenic mice, with T cells that recognize the IAα-restricted, LCMV epitope GP69-80 [70], were bred at the University of Massachusetts Medical School (UMMS) Department of Animal Medicine. CD45.1+ OT-1 CD8 TCR-transgenic mice, with T cells that recognize the Kb-restricted, ovalbumin epitope OVA257-264 [71], and CD45.1+ OT-2 CD4 TCR-transgenic mice, with T cells that recognize the IAα-restricted, ovalbumin epitope OVA323-339 [72] were provided by Dr. Kenneth Rock (Department of Pathology, UMMS, Worcester, MA). NG-BAC transgenic mice, originally obtained from Dr. Michel Nussenzweig were back-crossed to the CD45.1+ and CD45.2+ background and were used at 6–12 weeks of age [3,73]. Homozygous C57BL/6J-Dnd1<sup>tm1</sup>N12 (H-2D<sup>+</sup>) KO mice were purchased from Taconic Farms and used at 6 weeks of age. All animals were housed and maintained within the Department of Animal Medicine at UMMS.

Viruses

Stocks of LCMV, strain Armstrong, and a LCMV variant GP1V virus that possesses an amino acid mutation at position 38 (F to L) in the GP33-41 epitope of LCMV Armstrong were used. This mutation results in the escape of the virus from recognition by LCMV specific D<sup>+</sup>-restricted CTL [74]. Both LCMV stocks were prepared in baby hamster kidney cells (BHK21), as previously described, and mice were infected with 5×10<sup>5</sup> PFU of each virus strain i.p. [8].

Flow cytometry and intracellular cytokine assays

Single cell suspensions of thymocytes and splenocytes were prepared in RPMI 1640 supplemented with 10% FBS, 100U/ml penicillin, 100µg/ml streptomycin sulfate and 2mM L-glutamine and stimulated as indicated. For intracellular cytokine assays, lymphocytes (2×10<sup>6</sup> cells) were stimulated with either 1 µM of the indicated peptide with monoclonal antibodies specific for CD3ε (0.25 µg/ml, 145-2C11, BD Pharmingen) and CD28 (2.5µg/ml, 37.51, BD Pharmingen) or with PMA (0.5 µg/ml) and ionomycin (0.5µg/ml) in the presence of GolgiPlug™ (0.1 µg/ml) for 4 hours at 37°C in 5% CO2. In some experiments, lymphocytes were cultured at 1:1 ratios with either splenocytes from the indicated mouse strains or with the indicated cell populations derived from the spleens of congenic B6 mice and stimulated simultaneously. After the incubation, cells were stained with monoclonal antibodies specific for congenic markers (CD45.1: A20) and (CD45.2: 104), CD4 (RM4-5), CD8 (53-67.6), CD25 (PC61), CD44 (IM7), CD62L (MEL-14), CD69 (H1.2F3), CD24 (M1/69), Qa2 (1-1-2), TCR Vα2 mAb (B20.1) and Vβ8.1 mAb (MR5-2) purchased from BD Pharmingen and C57BL/6J (B6) mice were purchased from The Jackson Laboratory (Bar Harbor, ME) and used at 6–12 weeks of age [3,73]. Homozygous C57BL/6Ji-Dbtm1 N12 (H-2D<sup>+</sup>) KO mice were purchased from Taconic Farms and used at 6 weeks of age. All animals were housed and maintained within the Department of Animal Medicine at UMMS.

Cell purification and enrichment

Single cell suspensions of thymocytes and splenocytes from P14 TCR transgenic mice were purified by staining with anti-CD4, anti-CD8 and anti-CD4 antibodies in 1× PBS with 2% FBS, 2mM EDTA and sorted for CD4<sup>+</sup> CD8<sup>+</sup> SP thymocytes and naïve (CD4<sup>+</sup>CD8<sup>+</sup>) splenic T cells using the MoFlo<sup>TM</sup> XDP cell sorter (Beckton Coulter). The purity of SP CD8 thymocytes was 98%.
Figure 5. Post-thymic maturation status of naïve P14 transgenic T cells determines their TNF producing capability. Female CD45.1\(^+\) P14-CD8\(^+\) thymocytes were transferred into female CD45.2\(^+\) B6 congenic mice. Host spleens were recovered after the indicated time periods and were stimulated in vitro for 4 hours with GP33\(+\,\alpha\,CD28\) and donor CD45.1\(^+\) T cells were stained for intracellular TNF as described in Materials and Methods. Dead cells were excluded using Live Dead Aqua Dead cell stain for this experiment. For analysis, cells were gated on the live donor SP P14-CD8\(^+\) T cells and the maturation profile of donor cells that are TNF\(^+\) (indicated by arrows in the boxed quadrants in plots iii, iv, v, vi) were compared at all the time points shown (corresponding histograms). Additionally, some CD45.1\(^+\) P14-CD8\(^+\) thymocytes were stimulated before transfer in the context of CD45.2\(^+\) B6 splenocytes in vitro for 4 hours with GP33\(+\,\alpha\,CD28\) and their maturation profile was compared to CD45.1\(^+\) P14-CD8\(^+\) thymocytes and splenocytes stimulated alone in vitro (plots i, ii and vii).

doi:10.1371/journal.pone.0015038.g005
and the purity of CD6+CD44low splenic T cells was 90.3%. For cell enrichment, subsets of P14-CD8+ T cells were obtained by negative magnetic selection in 1× PBS with 2% FBS, 2mM EDTA. For this, thymocytes were depleted of CD4+ cells and splenocytes were depleted of CD4+ and CD19+ cells by initially staining the cells with biotinylated anti-CD4 (RM4-5; BD Pharmingen) and anti-CD19 (ID3; BD Pharmingen) followed by staining the cells with biotinylated anti-CD4 (RM4-5; BD Pharmingen) and anti-CD19 (ID3; BD Pharmingen) followed by selection with Streptavidin (SA) microbeads (Miltenyi Biotech, Auburn, CA). The purification of CD8+ cells after negative selection was 60% from both tissues. To isolate cell subsets from the CD45.2+ splenocytes for the co-culture experiments described above, B cells were positively selected using anti-CD19 microbeads and T cells were positively selected by Thy1.2 microbeads (Miltenyi Biotech). The cells remaining in the flow-through were used as a source of splenic APCs (20% CD11c+).

Real-time PCR

T cell subsets purified either by sorting or enrichment were used as indicated. Total RNA was isolated using a RNA isolation kit (Qiagen Valencia, CA). An additional step was incorporated to remove genomic DNA using a RNase-free DNase kit (Qiagen).

The concentration of recovered RNA was determined using the NanoDrop® ND-1000 spectrophotometer (Thermo Scientific Wilmington, DE). RNA (25 or 50 ng as indicated) was reverse-transcribed into cDNA using Superscript™ III first strand synthesis system (Invitrogen Carlsbad, CA) using oligo(dT) primers. Amplification of the cDNA was then performed by Real time PCR with the SYBR® green mastermix (Applied Biosystems Foster City, CA) using MyQ™ BioRad iCycler. The following TNF primers: FW 5′-CAT CTT CTC AAA ATT CGA GTG CTA CCA TGT G-3′, RV 5′-TGG GAG TAG ACA AGG TAC AAC CC-3′ primers (annealing temp: 60°C and 175 bp product) [75]; β actin primers: FW 5′-CGA GCC CCA GAG CAA GAG AG-3′, RV 5′-GGG TTG GCC TTA GGG TGC AG-3′ and (annealing temp: 62°C and 150 bp product) were used. The following program was used for the real time PCR reaction, Cycle 1: (1× step 1: 95°C for 10:00; Cycle 2: (40× step 1: 95°C for 00:15; step 2: 60°C for 00:10. For absolute quantification of the data, standard curves were generated using serial dilution of pCR® 4–TOPO M13 plasmids containing cDNA clones of TNF and β actin.

### Table 1. SP thymocytes acquire the ability to produce TNF as a function of time in the peripherya.

<table>
<thead>
<tr>
<th>Group</th>
<th>Days post-transfer</th>
<th>Absolute number of donor P14-CD8+ T cells (x10⁶)</th>
<th>TNF MFI of TNF+CD44hi donor T cells (x10⁶)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.</td>
<td>Day 1</td>
<td>48±8</td>
<td>24±4</td>
</tr>
<tr>
<td>2.</td>
<td>Day 2</td>
<td>36±18</td>
<td>24±2</td>
</tr>
<tr>
<td>3.</td>
<td>Day 7</td>
<td>14±10b</td>
<td>47±4f</td>
</tr>
<tr>
<td>4.</td>
<td>Day 14</td>
<td>5±2*</td>
<td>64±19*a</td>
</tr>
</tbody>
</table>

aThe recovery of the donor (CD44hi) SP P14-CD8+ thymocytes from recipient spleens and the MFI of TNF expression at the indicated time points post-transfer are shown. The average recovery and the MFI of TNF expression by donor thymocytes (n = 6 per time point) were analysed using One-way ANOVA with a Tukey post-test as described in Materials and Methods. Error indicates SD. N/A, Not Applicable.

b p<0.05 vs Table II, group 1.
c p<0.05 vs Table I, group 1.
d p<0.05 vs Table II, group 1.
e p<0.05 vs Table I, group 2.
f p<0.05 vs Table I, group 3.
g p<0.05 vs Table I, group 4.
h p<0.05 vs Table II, group 1.
i p<0.05 vs Table II, group 2.
j p<0.05 vs Table II, group 3.
k p<0.05 vs Table II, group 4.
l p<0.05 vs Table III, group 1.
m p<0.05 vs Table III, group 2.

### Table 2. Maturation state of SP thymocytes reflects their TNF producing capabilitya.

<table>
<thead>
<tr>
<th>Group</th>
<th>Days post-transfer</th>
<th>CD24 MFI of donor T cells (x10²)</th>
<th>CD45RB MFI of donor T cells (x10²)</th>
<th>Qa2 MFI of donor T cells (x10²)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>TFN+</td>
<td>TFN−</td>
<td>TFN+</td>
<td>TFN−</td>
</tr>
<tr>
<td>1.</td>
<td>Day1</td>
<td>24±6</td>
<td>34±7h</td>
<td>239±15</td>
</tr>
<tr>
<td>2.</td>
<td>Day2</td>
<td>20±3</td>
<td>31±5i</td>
<td>305±33d</td>
</tr>
<tr>
<td>3.</td>
<td>Day7</td>
<td>15±3b</td>
<td>N/A</td>
<td>588±26f</td>
</tr>
<tr>
<td>4.</td>
<td>Day14</td>
<td>13±2c</td>
<td>N/A</td>
<td>629±27f</td>
</tr>
</tbody>
</table>

aThe average MFI of maturation markers CD24 (n = 6), CD45RB (n = 6) and Qa2 (n = 3 for group 1 and 2 and n = 6 for group 3 and 4) in TNF producing donor SP P14-CD8+ are shown. The averages were analysed using One-way ANOVA with a Tukey post-test as described in Materials and Methods. Error indicates SD. N/A, Not Applicable.

b p<0.05 vs Table II, group 1.
c p<0.05 vs Table I, group 1.
d p<0.05 vs Table II, group 1.
e p<0.05 vs Table I, group 2.
f p<0.05 vs Table II, group 1.
g p<0.05 vs Table I, group 3.
h p<0.05 vs Table II, group 4.
i p<0.05 vs Table III, group 1.
j p<0.05 vs Table III, group 2.
k p<0.05 vs Table III, group 3.
l p<0.05 vs Table III, group 4.

**Maturation Determines TNF Production by T Cells**

DOI: 10.1371/journal.pone.0015038.t001

DOI: 10.1371/journal.pone.0015038.t002
Figure 6. Post-thymic maturation status of naive polyclonal T cells determines their TNF producing capability. A, The percentages of non-transgenic CD8+ and CD4+ cells (both CD44lo and CD44hi) from thymi and spleens of B6 mice staining positive for TNF cytokine are shown. B, Thymocytes and splenocytes from NG-BAC transgenic mice were stimulated with αCD3+αCD28 for 4 hours and then stained for maturation markers and intracellular TNF. The GFP profile of SP thymocytes, RTEs and MN T cells in the CD8+ and CD4+ compartments is shown. B and C, The percentages
In-vivo Brefeldin A (bFA) Assay

This assay was modified from a previously published protocol [45] and used to detect TCR-transgenic T cells producing TNF in vivo. Briefly, unpurified thymocytes or splenocytes from CD45.1+ P14 and CD45.1+ SMARTA mice 10×10^6 each (mixed at a 1:1 ratio) were treated in vitro with 0.5 μg/ml GolgiPlug (BD biosciences) for 20 min at 37°C. Following the incubation, the cells were adoptively transferred into CD45.2+ B6 hosts that were infected 2 days previously with 5×10^5 PFU of LCMV Armstrong or GP1V CTL escape variant. Additionally each mouse received 250 μg of bFA (Sigma) i.v. Four hours after transfer, host spleens were harvested and donor T cells were stained directly for TNF using the intracellular cytokine staining protocol as described above. Additionally, 20×10^6 P1-4 thymocytes and splenocytes were transferred separately into uninjected CD45.2+ B6 hosts in the absence of bFA. Host spleens were harvested 1, 2, 7 and 14 days after transfer and stimulated in vitro with 1 μM GP33 peptide and 2×CD28 (2.5μg/ml) in the presence of GolgiPlug™ (0.1 μg/ml) for 4 hours at 37°C in 5% CO₂ followed by standard intracellular cytokine staining protocol for TNF by donor T cells as described above. Dead cells were excluded using Live Dead Aqua Dead cell stain (Invitrogen; Molecular probes Carlsbad, CA) for this experiment.

Statistics

Sample analyses were done using Graph Pad Prism (Graph Pad Software). A one-way ANOVA with a Tukey post-test. The data are representative of 4 individual mice. The error bars indicate SD.

Acknowledgments

We thank the University of Massachusetts Medical School Flow Cytometry Core Lab for performing cell sorting. We thank Dr. Kenneth Rock for providing us with OT-II mice. We also thank Dr. Rabinarayana Mishra and Keith Daniels for the helpful discussions and excellent technical assistance and Erin Cloherty for expert mouse management.

Author Contributions

Conceived and designed the experiments: BP RMW DLG MAB. Performed the experiments: BP MAB. Analyzed the data: BP RMW MAB. Contributed reagents/materials/analysis tools: RMG. Wrote the paper: BP RMW RMG MAB.

References


