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Autosomal Dominant Hypoparathyroidism Associated with Short Stature and Premature Osteoarthritis*

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ABSTRACT
Familial hypoparathyroidism is an unusual and genetically heterogeneous group of disorders of various inheritance patterns that may be associated with other abnormalities such as autoimmune polyglandular disease (1) and congenital syndromes such as DiGeorge, Kenney-Caffey, or Barakat (2). Hypoparathyroidism may also develop as an isolated entity and most often occurs sporadically, but may also occur in a familial pattern. In different families, isolated hypoparathyroidism shows different modes of inheritance [autosomal dominant (3), autosomal recessive (4), or X-linked recessive (5)], suggesting that different genetic defects can produce the same phenotype.

Isolated familial hypoparathyroidism has been described as a result of a mutation of the signal peptide-encoding region of the preproparathyroid hormone gene on chromosome 11p (6). It may also be caused by an activating mutation of the calcium receptor (CaR) gene, further studies were performed. Sequencing of PCR-amplified genomic DNA revealed a leucine to valine substitution at position 616 in the first transmembrane domain of the CaR, which cosegregated with the disorder. However, this amino acid sequence change did not affect the total accumulation of inositol phosphates as a function of extracellular calcium concentrations in transfected HEK-293 cells.

In conclusion, a sequence alteration in the coding region of the CaR gene was identified, but is not conclusively involved in the etiology of this novel syndrome. The cosegregation of hypoparathyroidism, short stature, and osteoarthritis in this kindred does suggest a genetic abnormality involving a common molecular mechanism in parathyroid, bone, and cartilage. (J Clin Endocrinol Metab 84: 3036–3040, 1999)


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Familial hypoparathyroidism is an unusual and genetically heterogeneous group of disorders that may be isolated or may be associated with congenital or acquired abnormalities in other organs or glands. We have evaluated a family with a novel syndrome of autosomal dominant hypoparathyroidism, short stature, and premature osteoarthritis. A 74-yr-old female (generation I) presented with hypoparathyroidism, a movement disorder secondary to ectopic calcification of the cerebellum and basal ganglia, and a history of knee and hip replacements for osteoarthritis. Two members of generation II and one member of generation III were also documented with hypoparathyroidism, short stature, and premature osteoarthritis evident as early as 11 yr.

Because of the known association between autosomal dominant hypoparathyroidism and activating mutations of the calcium-sensing receptor superfamily, CaR transcripts and protein have been localized to tissues involved in calcium homeostasis including parathyroid, C cells of the thyroid, kidney, bone cells (14), and cartilage (15). Activation of the CaR leads to decreased secretion of PTH and inhibition of renal calcium reabsorption.

We describe a family with a novel syndrome of autosomal dominant hypoparathyroidism associated with short stature and premature osteoarthritis and have investigated the possibility of a mutation in the CaR gene as the cause of this syndrome.

Subjects and Methods

Case report (Fig. 1 and Table 1)

Family members in three generations were studied after informed consent was obtained. The study was approved by the NICHD institutional review board. The index case (patient I-1) presented at age 74 yr with uncontrolled movement of the left leg. Her past medical history was significant for multiple lumbar compression fractures at age 68 yr and hypocalcemia. At age 70 yr the patient underwent bilateral knee and right hip replacements for degenerative arthritis. The physical examination was significant for short stature (16) and frequent, involuntary choreiform movements of the left leg. Additional laboratory studies revealed a serum albumin concentration of 4.0 g/dL, magnesium of 1.7 mg/dL, and 25-hydroxyvitamin D of 33 ng/mL (normal, 10–55). A computed tomography scan of the head revealed calcification of the cerebellum and basal ganglia bilaterally. Over the following 6 months
**DNA amplification and sequence analysis**

Genomic DNA was isolated from white blood cells and exons 1–6 of the CaR were PCR amplified with previously reported primers (17). Screening for mutations was performed by heteroduplex analysis using mutation detection enhancement gels (18). PCR products that showed heteroduplex bands from members III-2 and III-3 were sequenced using mutation detection enhancement gels (18). PCR-amplified genomic DNA was digested with restriction enzyme MaeIII, subjected to electrophoresis through a 6% polyacrylamide gel, and stained with ethidium bromide.

**Restriction analysis**

PCR-amplified genomic DNA was digested with restriction enzyme MaeIII, subjected to electrophoresis through a 6% polyacrylamide gel, and stained with ethidium bromide.

**Site-directed mutagenesis, cell culture, transfection, and preparation of cell membranes**

The human CaR complementary DNA (cDNA) inserted into the mutagenesis vector pAlter I (Promega Corp., Madison, WI) was obtained from NPS Pharmaceuticals, Inc. (Salt Lake City, UT). The detected nucleotide change (C to G) at nucleotide 1846 of exon 6 or other nucleotide changes were introduced into this construct by site-directed mutagenesis using the Altered Sites II system (Promega Corp.). The mutated cDNA were then isolated with restriction enzymes XbaI and HindIII, inserted into the expression vector pCDNAI/Amp (Invitrogen, San Diego, CA), and confirmed by DNA sequencing.

**Cell culture and transfection**

HEK-293 cells were cultured in DMEM. The cells were plated in 24-well plates (10⁵ cells/well) and transiently transfected with constructs encoding the wild-type and the receptors with nucleotide changes, using 5 μL Lipofectamine (Life Technologies, Inc., Gaithersburg, MD) and 0.5 μg DNA.

**Assessment of cell surface receptor expression by enzyme-linked immunosorbent assay (ELISA)**

Transfected cells were suspended in 1% BSA-DMEM for 30 min at 4°C and then incubated with monoclonal antibody 7F8 (20 μg/mL) for 1 h at 4°C. This antibody was made by immunization with the purified extracellular domain of the human CaR (Goldsmith, P. K., manuscript in preparation). After washing, cells were further incubated with peroxidase-conjugated goat antimouse secondary antibody (Kirkegaard & Perry Laboratories, Inc., Gaithersburg, MD; 1:1000 dilution). After washing, peroxidase substrate was added. Absorbance was measured at 405 nm. Four independent transfections were performed for ELISA.

**Measurement of phosphoinositides (IPs)**

Forty-eight hours after transfection, HEK-293 cells were labeled with 3 μCi/mL myo-[3H]inositol (New England Biolabs, Inc., Beverly, MA) in DMEM for 16–24 h. Cells were then incubated in PI buffer (99 mmol/L NaCl, 5 mmol/L KCl, 5.6 mmol/L glucose, 0.4 mmol/L MgCl₂, and 0.5 mmol/L CaCl₂) containing 20 mmol/L LiCl for 1 h. Cells were stimulated with the indicated concentrations of Ca²⁺ (in PI buffer) for 30 min at 37°C. The reactions were terminated with acid-methanol (167 μL HCl in 120 mL methanol). Total inositol phosphates were extracted, separated on Dowex AG1-X8 columns as previously described (Berridge, 1983), and counted by liquid scintillation. Nine independent transfections were performed at each Ca²⁺ concentration for IP measurement.

**Statistical analysis**

Results were expressed as the mean ± SEM. Significance was assessed using an unpaired t test. p < 0.05 was considered statistically significant.

**Results**

**DNA sequence analysis**

To screen for sequence alterations in the CaR, individual exons from genomic DNA were PCR amplified and subject to heteroduplex analysis. Possibly affected subject III-2 and definitely affected subject III-3 showed heteroduplex bands in exon 6 (data not shown). Direct sequencing of PCR-amplified genomic DNA from possibly affected subject III-2 and definitely affected subject III-3 revealed a heterozygous C to G basepair substitution at position 1846 in exon 6. This change produces a leucine to valine substitution at position 616 in the first transmembrane domain of the receptor (L616V).

Genomic DNA from affected subjects I-1, II-1, II-4, and III-3 and from possibly affected subjects III-1 and III-2 were screened for this possible sequence alteration by restriction
response to Ca\textsuperscript{2+} was no significant difference in the maximal or minimal concentrations, and IP accumulation was measured. There was no significant difference in the maximal or minimal concentration to Ca\textsuperscript{2+}. Although there may have been a slight leftward shift in the concentration-response curve for the L616V receptor, the decrease in EC\textsubscript{50} compared to that for the wild-type receptor did not quite reach statistical significance ($P = 0.054$).

**Discussion**

We have described a family with mild hypoparathyroidism associated with short stature and premature osteoarthritis, inherited in an autosomal dominant pattern. To our knowledge, this is the first report of this association. We considered a mutation in the CaR gene in members of this family in view of recent reports demonstrating gain of function mutations in the extracellular domain of the CaR in families with autosomal dominant hypoparathyroidism (9). Given the report of CaR expression in human bone cells (14) and in animal articular and growth plate cartilage (15), we also hypothesized that an abnormality in the CaR in bone or cartilage might also contribute to the familial short stature and premature osteoarthritis in this family.

In our study we found a heterozygous C to G basepair substitution at position 1846 in exon 6 that cosegregated with the disorder. This basepair change produced a change in the amino acid sequence of leucine to valine at amino acid position 616. This amino acid sequence change did not affect total accumulation of inositol phosphates as a function of extracellular calcium concentrations in transfected HEK-293 cells.

In the same experiment, we measured the function of the activating mutation F612S, identified in a family with autosomal dominant hypoparathyroidism (9). The results have been reported previously (12) and serve as a positive control compared to the wild-type results in Table 2. This mutation produced a leftward shift in the concentration-response curve, with a decrease in the EC\textsubscript{50} to 2.3 \pm 0.06 mmol/L ($P < 0.001$) and an increase in the maximum IP accumulation to $42 \pm 3 \times 10^3$ cpm ($P < 0.02$) compared to wild-type values.

However, the cell surface receptor expression in our in vitro system was decreased for L616V compared to wild-type receptors. In the same experiment, the cell surface expression of mutation F612S was also lower than that in wild-type receptors (1.3 \pm 0.6 optical density units), but did not reach

### TABLE 1. Clinical and biochemical features of members of a kindred with autosomal dominant hypoparathyroidism, short stature, and premature osteoarthritis

<table>
<thead>
<tr>
<th>Subject no.</th>
<th>Age (yr)</th>
<th>Serum calcium (mg/dL)</th>
<th>Serum phosphorus (mg/dL)</th>
<th>Serum PTH (pg/mL)</th>
<th>Urinary calcium (mg/24 h)</th>
<th>Ht (%)</th>
<th>Osteoarthritis</th>
</tr>
</thead>
<tbody>
<tr>
<td>I-1</td>
<td>74</td>
<td>7.8\textsuperscript{*}</td>
<td>5.6</td>
<td>21 pg/mL\textsuperscript{k}</td>
<td>133\textsuperscript{*}</td>
<td>&lt;5</td>
<td>+</td>
</tr>
<tr>
<td>II-1</td>
<td>45</td>
<td>6.0\textsuperscript{*}</td>
<td>3.8</td>
<td>27 ng/mL\textsuperscript{k}</td>
<td>146,208\textsuperscript{*}</td>
<td>&lt;5</td>
<td>+</td>
</tr>
<tr>
<td>II-4</td>
<td>31</td>
<td>7.8\textsuperscript{*}</td>
<td>4.0</td>
<td>21 pg/mL\textsuperscript{k}</td>
<td>54\textsuperscript{*}</td>
<td>&lt;5</td>
<td>+</td>
</tr>
<tr>
<td>III-1</td>
<td>24</td>
<td>7.9\textsuperscript{*}</td>
<td>4.0</td>
<td>31 pg/mL\textsuperscript{k}</td>
<td>&lt;10</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td>III-2</td>
<td>13</td>
<td>8.6\textsuperscript{*}</td>
<td>6.4</td>
<td>140 pg/mL\textsuperscript{k}</td>
<td>68</td>
<td>&lt;5</td>
<td>−</td>
</tr>
<tr>
<td>III-3</td>
<td>11</td>
<td>8.1\textsuperscript{*}</td>
<td>5.6</td>
<td>110 pg/mL\textsuperscript{k}</td>
<td>24</td>
<td>5</td>
<td>+</td>
</tr>
</tbody>
</table>

\textsuperscript{*} Normal, 2.5–4.5 mg/dL.

\textsuperscript{a} Percent 18 yr-old height for adults and percent age-matched height for subjects III-2 and III-3 (16).

\textsuperscript{b} Normal, 50–340 pg/mL, intact.

\textsuperscript{c} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{d} Normal, 9–0–10.6 mg/dL.

\textsuperscript{e} Normal, 8.4–10.2 mg/dL.

\textsuperscript{f} Normal, 5.6–110 pg/mL, midregion.

\textsuperscript{g} Normal, 8.5–10.5 mg/dL.

\textsuperscript{h} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{i} Normal, 50–340 pg/mL, midregion.

\textsuperscript{j} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{k} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{l} Normal, 8.4–10.2 mg/dL.

\textsuperscript{m} While taking calcium and ergocalciferol.

\textsuperscript{n} While taking calcium and ergocalciferol, and hydrochlorothiazide/triamterine.

\textsuperscript{o} While taking calcium and ergocalciferol, and hydrochlorothiazide/triamterine.

\textsuperscript{p} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{q} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{r} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{s} Normal, 8.5–10.2 mg/dL.

\textsuperscript{t} Normal, 50–340 pg/mL, midregion.

\textsuperscript{u} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{v} Normal, 8.5–10.2 mg/dL.

\textsuperscript{w} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{x} Normal, 8.5–10.2 mg/dL.

\textsuperscript{y} Normal, 10–65 pg/mL, C-terminal.

\textsuperscript{z} Normal, 8.5–10.2 mg/dL.

\textsuperscript{AA} Normal, 2.5–4.5 mg/dL.

\textsuperscript{BB} Normal, 8.4–10.5 mg/dL.

\textsuperscript{CC} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{DD} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{EE} Normal, 50–340 pg/mL, midregion.

\textsuperscript{FF} Normal, 8.4–10.2 mg/dL.

\textsuperscript{GG} Normal, 8.5–10.5 mg/dL.

\textsuperscript{HH} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{II} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{JJ} Normal, 50–340 pg/mL, midregion.

\textsuperscript{KK} Normal, 8.4–10.2 mg/dL.

\textsuperscript{LL} Normal, 8.5–10.5 mg/dL.

\textsuperscript{MM} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{NN} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{OO} Normal, 50–340 pg/mL, midregion.

\textsuperscript{PP} Normal, 8.4–10.2 mg/dL.

\textsuperscript{QQ} Normal, 8.5–10.5 mg/dL.

\textsuperscript{RR} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{SS} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{TT} Normal, 50–340 pg/mL, midregion.

\textsuperscript{UU} Normal, 8.4–10.2 mg/dL.

\textsuperscript{VV} Normal, 8.5–10.5 mg/dL.

\textsuperscript{WW} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{XX} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{YY} Normal, 50–340 pg/mL, midregion.

\textsuperscript{ZZ} Normal, 8.4–10.2 mg/dL.

\textsuperscript{AAA} Normal, 8.5–10.5 mg/dL.

\textsuperscript{BBB} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{CCC} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{DDD} Normal, 50–340 pg/mL, midregion.

\textsuperscript{EEE} Normal, 8.4–10.2 mg/dL.

\textsuperscript{FFF} Normal, 8.5–10.5 mg/dL.

\textsuperscript{GGG} Normal, 0–180 pg/mL, C-terminal.

\textsuperscript{HHH} Normal, 18–120 pmol/L, C-terminal.

\textsuperscript{III} Normal, 50–340 pg/mL, midregion.
statistical significance \( (P = 0.06) \) \( (12) \). The signal transduction activity per receptor may be higher for \( L616V \) than for wild-type receptors. If the cell surface expression of the \( L616V \) receptor \( \textit{in vivo} \) was similar to that of the wild-type receptor, then this possibly increased activity per receptor might account for the phenotype. Alternatively, a mutation in the promoter or in other regulatory genes for the expression of the CaR cannot be ruled out. It is also possible that this sequence alteration may affect another signal transduction pathway or that the causative mutation lies in a nearby gene.

There are several families with autosomal dominant hypoparathyroidism in whom CaR mutations have not been documented despite extensive analysis \( (8, 9) \).

A clinical issue to be considered in caring for members of this family is surveillance for ectopic calcifications, which have had significant effects on family member I-1 despite the mild nature of her disease. Basal ganglia calcifications and subsequent movement disorders have been described in mild chronic hypoparathyroidism \( (20) \). Periodic brain imaging studies might be performed in these patients. Whether treatment of affected members to raise their serum calcium concentrations to low normal levels will be of benefit in preventing or delaying ectopic calcifications remains to be elucidated. Hypercalcuiuria, usually a concern in the treatment of hypoparathyroidism, was not demonstrated in this family. Hypercalcuiuria has been found in families with documented mutations of the CaR, probably related to a decrease in the tubular reabsorption of calcium \( (10) \).

The description of this family extends previous observations that familial hypoparathyroidism is a heterogeneous disorder. A sequence alteration in the coding region of the CaR gene was identified and cosegregates with the phenotype, suggesting a causal relationship. However, the expression data and functional analysis do not conclusively demonstrate that this sequence alteration activates the receptor. The association of hypoparathyroidism, short stature, and osteoarthritis in this kindred does suggest a genetic abnormality involving a common molecular mechanism in parathyroid, bone, and cartilage.

**Acknowledgments**

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