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Baojin Ding

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Temporal Regulation of Nuclear Factor One Occupancy by Calcineurin/NFAT Governs a Voltage-Sensitive Developmental Switch in Late Maturing Neurons

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Introduction

The timing of dendrite formation and synaptogenesis is intimately involved in establishing unique patterns of neuronal connectivity (Deguchi et al., 2011; Tripodi and Arber, 2012). Accordingly, gene expression must be precisely coordinated in time to ensure that synaptic circuits properly form (Hippenmeyer et al., 2005; Di Cristo et al., 2007; Petrovic and Hummel, 2008). Temporal dysregulation leading to altered synapse formation has been implicated in several neurodevelopmental disorders (Geschwind and Levitt, 2007; Leonardo and Hen, 2008; Meredith et al., 2012).

Cerebellar granule neurons (CGNs) function in cerebellar information processing via synaptic connections with mossy fiber inputs and Purkinje cell outputs (D’Angelo et al., 2011). CGNs have been studied extensively to elucidate mechanisms governing neuronal development, including dendritogenesis and synapse formation (Goldowitz and Hamre, 1998; Hall et al., 2000; Ito and Takeichi, 2009). CGN progenitors (CGNPs) proliferate in the external germinal layer (EGL) and generate immature neurons within the premigratory zone (PMZ). CGN cell bodies later migrate inwardly until reaching the internal granule cell layer (IGL), where postmigratory CGNs form mature dendrites and synaptic connections with input neurons. Numerous genes are sequentially expressed during these different developmental stages (Goldowitz and Hamre, 1998; Furuchi et al., 2011). Much of this developmental program is recapitated in CGN cultures (Ito...
and Takeichi, 2009; de la Torre-Ubieta et al., 2010) wherein CGNs and immature CGNs isolated from the EGL/PMZ (Raetzman and Siegel, 1999) differentiate into IGL-like cells upon plating (Manzini et al., 2006; Wang et al., 2011).

Nuclear factor I (NFI) transcription factors are important regulators of CGN maturation (Kilpatrick et al., 2012). They are expressed throughout CGN postmitotic development and regulate parallel fiber extension, migration, dendritogenesis, and synapticogenesis (Wang et al., 2007, 2010; Piper et al., 2011). Recently, NFI proteins were shown to control the dendritogenesis-linked expression of the $\alpha_6$ GABA$\alpha$ receptor subunit (Gabra6) gene within the developing IGL (Wang et al., 2011). CGNs thus provide a robust model for exploring the programming of dendrite and synapse formation and the role of NFI proteins in these events (Kilpatrick et al., 2012).

CGNs undergo a shift in membrane potential as they differentiate, becoming increasingly hyperpolarized with maturation (Rossi et al., 1998; Cathala et al., 2003; Okazawa et al., 2009). This change in membrane potential has been implicated in the control of mouse CGN differentiation in culture and within the developing cerebellum (Nakanishi and Okazawa, 2006). In particular, depolarization of immature CGNs inhibits developmental changes in gene expression as well as dendritic and synaptic maturation via Ca$^{2+}$-dependent activation of calcineurin (CaN) (Mellor et al., 1998; Ives et al., 2002; Sato et al., 2005; Iijima et al., 2009; Okazawa et al., 2009).

Here we describe an NFI-regulated temporal switch program that regulates late differentiation of postmitogenic CGNs, including dendrite formation. A central feature of this program is delayed NFI occupancy of numerous genes that are temporally upregulated as CGNs mature. Further, we show that this NFI switch is directly linked to membrane potential-dependent mechanisms via its regulation by CaN activation of nuclear factor of activated T-cells, cytoplasmic (NFATc) transcription factors.

**Materials and Methods**

**Animals and primary cultures.** Mouse CGNs were prepared from 6- to 7-day-old CD1 mouse pups of either sex as previously described (Wang et al., 2004). Individual culture experiments were performed using cells prepared from the same litter. Cells were plated at a density of 5 × $10^4$ cells/cm$^2$ onto chamber slides or cell culture dishes coated with poly-D-lysine/laminin (Invitrogen) in Neurobasal medium (Invitrogen) containing B-27 serum-free supplement (50 $\mu$g/ml; Invitrogen) (Wang et al., 2004). The following reagents were added to culture medium 1 h after cell plating and were refreshed on 3 d in vitro (DIV): 5 $\mu$M nimodipine (Tocris Bioscience), 100 $\mu$M kainic acid (KA; Sigma), vehicle control (dimethylsulfoxide). Cerebellar tissues were obtained from postnatal Nfatc4 $^{–/–}$ mice on a C57BL/6 background as well as from wild-type (WT) C57BL/6 mice (Charles River). Nfatc4 $^{–/–}$ mice and WT littermates were on a C57BL/6 background.

**Plasmids and cell lines.** Self-inactivating lentiviruses expressing hemagglutinin (HA)-tagged NFI dominant repressor (NFI/EnR) or Drosophila engraded repressor domain (EnR) were described previously (Wang et al., 2004). FLAG-tagged Nfatc4 proteins ([constitutively active (Nfatc4-Ala) and dominant-negative NFAT (dnNFAT)] were released from their expression vectors (Chow et al., 1999; Yang et al., 2002) and inserted into BamHI and XbaI sites of the lentiviral expression vector pH$^{-}$2-CpPT-CMV-W-Sin18 (Wang et al., 2004). Human embryonic kidney 293T cells were grown in DMEM (Invitrogen) containing 10% heat-inactivated fetal bovine serum (Invitrogen).

**Chromatin immunoprecipitation assays.** Chromatin immunoprecipitation (ChIP) assays were performed as previously described (Wang et al., 2011). For tissues, nuclei were first purified by Percoll gradient centrifugation (Wang et al., 2011) or by differential centrifugation (1400 × g for 3 min, 4°C) in 0.25 M sucrose. Samples were assayed by PCR using polycrylamide gels or real-time PCR. PCR data were expressed as a percentage of input using the 2$^{–\Delta\DeltaCT}$ method. ChIP antibodies were to Xenopus NFIB (xNFIB) (Puziowska-Kuznicka and Shi, 1996), NFIA (Active Motif; 39036), NFATc4 (Santa Cruz Biotechnology; sc-1153), and FLAG epitope (Sigma; F1804). Pre-immune serum or normal rabbit IgG (Millipore; PP64) were used as negative controls. PCR primer sequences for ChIP assays are available upon request.

**Immunofluorescence and dendritogenesis assay.** Immunofluorescence was performed as in previous studies (Wang et al., 2007). Briefly, primary CGNs were cultured in coated chamber slides and then were fixed on 1.5 DIV or 2 DIV with 4% paraformaldehyde/PBS. Cells were permeabilized with 1% Triton X solution, incubated with 5% normal goat serum followed by primary antibodies at 4°C overnight and then Cy3-conjugated goat anti-rabbit IgG (NG180777; Millipore). Dendrite assays used anti-Map2 antibody (Millipore; AB5622) and 1 $\mu$g/ml Bizenfluoride (Sigma) to stain nuclei. Dendrite length was measured with Image Pro Plus 6.0 software. Dendritic tibrations used antibodies for HA (NFI/EnR and EnR; C29F4; Cell Signaling Technology) or FLAG (Nfatc4 proteins; M2 antibody, Sigma).

**RNA isolation and quantitative real-time PCR.** RNA was extracted from tissues or cultured cells using Tri reagent (Sigma), processed with DNA-free kit (Ambion), and complementary DNAs were prepared as described previously (Wang et al., 2007). Real-time PCRs were performed in triplicate using a StepOnePlus Real-Time PCR System (Applied Biosystems). Target transcripts were normalized to 18S rRNA by the 2$^{–\Delta\DeltaCT}$ method, and differences relative to control samples were calculated using 2$^{–\Delta\DeltaCT}$ as previously described (Wang et al., 2011). Numerical values for 2$^{–\Delta\DeltaCT} < 1.0$ were recalculated as control relative to experimental data and expressed as negative numbers. Real-time PCR primer sequences are available upon request.

**Microarray and bioinformatic analyses.** Total RNA was purified from CGN cultures with an RNeasy Plus Micro Kit (Qiagen). RNA quality analysis and hybridizations to GeneChip Mouse Genome 430A,B or 430 2.0 arrays (Affymetrix) were performed by the Genomics Core Facility of the University of Massachusetts Medical School. Differentially expressed genes for temporal and NFI arrays were analyzed separately. For NFI arrays (four NFI/EnR expressed vs four EnR control biological replicates), gene expression values in the Affymetrix cel files were first background-corrected and normalized together using RMA algorithm (Bioconductor R package). To identify genes with robust expression, we removed probes with low expression (i.e., expression value below 40th percentile in at least seven of eight arrays). For multiple probes mapping to the same gene, only the probe showing the highest expression change for each gene was retained. Finally, moderated t statistics and FDR correction (limma R package) were used to assess the significance of differential expression. NFI differentially regulated genes were defined as probes with FDR p value < 0.15 and fold change > 1.5. Identical steps for signal normalization, background noise correction, and statistical testing were applied to temporal arrays (1.5 DIV vs 6 DIV, three biological replicates each). Differentially expressed genes were defined as having probes with FDR p value < 0.1 and fold change > 2. We used more stringent criteria to define temporally regulated genes since the variability between replicates appeared smaller. Greater variability among the NFI/EnR array data likely reflected differing infectivities of distinct lentiviral preparations used for independent experiments. Finally, NFI-regulated and temporally regulated overlap genes showed significant differential expression for the same probes in both NFI and temporal datasets. Chi-square tests were used to assess the significance of the overlap dataset as well as its partitioning into four NFI/temporal categories based on the directionality of expression changes.

**Citations related to synaptic function, signaling, and excitability were identified in the PubMed database. Mouse gene promoter sequences were extracted from GenBank and were scanned for NFI consensus sites using the TRANSFAC database.**

**Chromatin immunoprecipitation assays.** Chromatin immunoprecipitation (ChIP) assays were performed as previously described (Wang et al., 2011). For tissues, nuclei were first purified by Percoll gradient centrifugation (Wang et al., 2011) or by differential centrifugation (1400 × g for 3 min, 4°C) in 0.25 M sucrose. Samples were assayed by PCR using polycrylamide gels or real-time PCR. PCR data were expressed as a percentage of input using the 2$^{–\Delta\DeltaCT}$ method. ChIP antibodies were to Xenopus NFIB (xNFIB) (Puziowska-Kuznicka and Shi, 1996), NFIA (Active Motif; 39036), NFATc4 (Santa Cruz Biotechnology; sc-1153), and FLAG epitope (Sigma; F1804). Pre-immune serum or normal rabbit IgG (Millipore; PP64) were used as negative controls. PCR primer sequences for ChIP assays are available upon request.

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Western blot. Total nuclear proteins were prepared from cultured CGNs as previously described (Wang et al., 2004). Purified NFIA, NFIB, and NFIX proteins were purchased from Abnova. Protein samples were separated on 8% SDS-polyacrylamide gels and transferred onto Pure Nitrocellulose membranes (GE Water & Process Technologies). Following blocking for 1 h, blots were incubated with primary antibodies at 4°C overnight and then with horseradish peroxidase-conjugated secondary antibody at real time for 1 h. Bound antibodies were detected with a chemiluminescent substrate (Thermo Scientific). Primary antibodies used were as follows: xNFIB antibody (1:200), pan-NFI antibody (1:1000; Santa Cruz Biotechnology, sc-5567), and anti-Histone H3 (1:1000; Abnova, PAB0653).

NFAT transactivation and CaN-dependent dephosphorylation assays. To determine NFAT transcriptional activity, cultured CGNs were transduced with an NFAT dual luciferase reporter lentiviral vector (Na et al., 2010) (kindly provided by the van den Brink lab, Memorial Sloan-Kettering Cancer Center, NY). NFAT trans-activation was quantified by measuring NFAT-inducible click-beetle red luciferase (CBRLuc) and normalizing activity to a constitutively expressed membrane-anchored Gaussia luciferase (extGLuc) using the Dual-Glo Luciferase Assay System (Promega). CaN-dependent dephosphorylation activity was measured using a cellular assay kit (Enzo Life Sciences) according to the manufacturer’s instructions. Data were normalized to protein concentration.

Statistics. In all quantitative studies other than microarrays, data were statistically analyzed using the two-tailed t test. Results were expressed as the mean ± SD, and p values < 0.05 were considered significant.

Results

NFI controls a developmental switch in maturing CGNs

NFI proteins regulate several developmentally expressed genes in maturing CGNs (Wang et al., 2007, 2010). Here we determined whether these proteins controlled a larger temporal program using a 2D microarray analysis of temporally regulated and NFI-regulated genes. For temporal genes, we compared immature CGNs (1.5 DIV) with more differentiated (6 DIV) cultures. CGN cultures (1.5 DIV) are largely postmitotic (Wang et al., 2005) and exhibit a morphology and pattern of gene expression consistent with immature CGNs in vivo, while 6 DIV cultures resemble IGL-like CGNs (Raetzman and Siegel, 1999; Manzini et al., 2006; Wang et al., 2010, 2011). This analysis identified 844 Temporal-Up or Temporal-Down genes (Fig. 1A). NFI-regulated genes were assayed in more mature 6 DIV CGN cultures that were transduced at the time of plating (0 DIV) with NFI dominant repressor (HA-NFI/EnR) or control (HA-EnR) proteins using lentiviruses (Wang et al., 2004). This identified 686 NFI-regulated genes that were either suppressed or elevated by NFI/EnR (Fig. 1A). Based on its dominant repressor function, NFI/EnR should inhibit expression of genes directly activated by NFI proteins. Consistent with this, HA ChIP detected NFI/EnR bound to the Gabra6 and Wnt7a NFI sites (Fig. 1B). Genes upregulated by the dominant repressor may have been indirectly derepressed via suppression of intermediary regulators required for their downregulation. Other mechanisms also are possible; e.g., certain genes may have been directly repressed by endogenous NFI proteins, but less efficiently by the dominant repressor resulting in their upregulation. The latter question was not further examined here.

Overlap analysis identified 212 genes that were both NFI and temporally regulated (Fig. 1A). This overlap was highly significant (p = 4.29e-222). Temporal genes were enriched ~8-fold within the NFI-regulated set relative to non-NFI regulated genes, indicating the importance of NFI proteins in developmental gene regulation. Overlap genes segregated into four gene subsets based on their temporal patterns and regulation by endogenous NFI proteins (Fig. 1C): Early/NFI-Down, Early/NFI-Up, Late/NFI-Down, Late/NFI-Up. Early and Late genes were temporally downregulated or upregulated on 6 DIV; NFI-Up genes were...
upregulated by NFI (repressed by NFI/EnR) while NFI-Down genes were inhibited by NFI (elevated by NFI/EnR) in 6 DIV cultures.

Nearly all (92%) NFI temporal genes clustered into two groups: Early/NFI-Down (62 genes; 29% of overlap genes) and a predominant set of Late/NFI-Up genes (134; 63% of total) (Fig. 1). This skewed distribution was also highly statistically significant (Fig. 1C), and it suggested a “developmental switch” mechanism in which NFI proteins downregulate genes expressed in immature CGNs and upregulate late-expressed genes (Fig. 1D). Based on this model, dominant repression of NFI function “trapped” CGNs in an immature state, stimulating or preventing the temporal downregulation of Early/NFI-Down gene expression, and directly or indirectly suppressing expression of Late/NFI-Up genes.

Regulation by NFI/EnR was confirmed in CGN cultures for a total of 34 genes using qRT-PCR (Table 1; Fig. 2), including several transcripts that did not reach significance in the overlap set but were of regulatory or functional interest. For 25 genes we also determined NFI regulation in vivo using P15 Nfia-null mouse cerebellum, which showed identical trends with NFI/EnR data with the exception of Egr1 (Table 1). This may reflect a compensatory or required role for other NFI family members in

Table 1. Validation of NFI switch gene regulation in culture and in vivo using qRT-PCR

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>Gene ID</th>
<th>6 DIV versus 0 DIV</th>
<th>P22 versus P7</th>
<th>NFI-EnR versus EnR</th>
<th>Nfia−/− versus WT</th>
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<tbody>
<tr>
<td>Late/NFI-Up</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td>Wnt7a</td>
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<td>9.9</td>
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<td>−2.9</td>
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<tr>
<td>Npx1</td>
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<td>44</td>
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<td>8.2</td>
<td>−10.8</td>
<td>−3.1</td>
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<tr>
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<td>43</td>
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<td>−6.8</td>
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<tr>
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<td>8.1</td>
<td>−2.3</td>
<td>−3.8</td>
</tr>
<tr>
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<td>23</td>
<td>17</td>
<td>−22.7</td>
<td>−5.5</td>
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<tr>
<td>Egr1</td>
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<td>−13</td>
<td>4.6</td>
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<td>—</td>
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<td>Early/NFI-Down</td>
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<td>−142</td>
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<td>2.6 **</td>
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<td>Neurod6</td>
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<td>Tpf3a</td>
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<td>1.4 **</td>
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<td>Late/NFI-Down</td>
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<td>−2.7 **</td>
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<td>Early/NFI-Up</td>
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<td>−2.1 *</td>
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<td>Id2</td>
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<td>Tiam1</td>
<td>21844</td>
<td>—</td>
<td>−20.8</td>
<td>−3.7</td>
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</tr>
</tbody>
</table>

Data are expressed as the relative difference in expression; negative values indicate reduced expression relative to controls. Dashes, not determined; p < 0.001 except * p < 0.05, ** p < 0.01; ns, no significant difference. Data are representative of three independent experiments.

Egr1 gene regulation. Temporal expression was also determined in culture and in vivo for 16 genes (Table 1), and 8 others (Tab1m1, Ets2, Kcnk3, Slc12a5, and GABA<sub>a</sub> receptor genes (Fig. 2) were previously reported (Laurie et al., 1992; Sato et al., 2005). This revealed that the NFI switch was activated between P7 and P21, when postmigratory CGNs within the IGL form dendrites and synapses. We also identified several temporally regulated non-overlap genes that were not dependent on NFI (Tle1, Scn2a1, Kcn1, Kcnk9, Prickle1, Rbfox3) (data not shown).

NFI controls GABA<sub>a</sub> receptor maturation and synapse-related gene expression

GABA<sub>a</sub> receptors undergo a developmental switch in subunit composition as postmigratory CGNs mature within the IGL, with α1-, α6-, β2-, β3-, γ2-, and δ-subunits being upregulated (Laurie et al., 1992; Zheng et al., 1993). This receptor switch is critical for controlling GABA excitability in vivo via GABA-induced tonic inhibition (Rossi and Hamann, 1998). Gabra6 (a6), Gabra1 (a1), Gabrb2 (β2), and Gabrg2 (γ2) were identified as Late/NFI-Up transcripts by microarrays, suggesting that NFI proteins regulated the GABA<sub>a</sub> receptor switch in CGNs. qRT-PCR confirmed that expression of these four genes was reduced by NFI dominant repression in CGN cultures and by Nfia deficiency in vivo (Fig. 2). Further, expression of the GABA<sub>a</sub> switch genes Gabrb3 (β3) and Gabrd (βδ) were also dependent on NFI. Interestingly, the Gabra2 (a2) and Gabra4 (a4) genes, which are transiently upregulated in maturing CGNs (Laurie et al., 1992), also were regulated by NFI proteins (Fig. 2), although Gabra4 was only modestly regulated. In contrast, the developmentally unregulated Gabra5 (a5) and downregulated Gabrg3 (γ3) receptor genes (Laurie et al., 1992) were insensitive to NFI functional disruption (Fig. 2).

Initial analysis of overlap genes showed enrichment of genes related to cell division in the Early/NFI-Down set (e.g., Ccn2a, Cks2) and neuronal differentiation in Late/NFI-Up genes (e.g., Ntrk3, Slit3, Wnt7a), among others (data not shown). Since the NFI switch is activated as CGNs form synapses within the IGL, we performed a literature analysis of known associations of NFI-regulated temporal genes with dendritic and synaptic structure/function, excitability, and signaling. Remarkably, 63 Late/NFI-Up genes (47%) had known links to these events (Table 2), consistent with their temporal upregulation by NFI. In contrast,
only seven Early/Down genes (11%) had known synapse-related activities or localization, indicating continuing functions of this small gene cohort in more mature CGNs following their developmental downregulation.

NFI occupancy of numerous Late/NFI-Up genes is temporally regulated

The timing of Gabra6 gene expression during CGN development is linked to delayed NFI binding to its promoter (Wang et al., 2011). Further, NFI proteins and their nuclear DNA binding activity are constitutively expressed during CGN maturation in culture and in vivo (Wang et al., 2011), indicating that NFI temporal occupancy of the Gabra6 gene is regulated at the level of chromatin interactions. We therefore examined whether temporally regulated NFI chromatin occupancy was a more global mechanism for six other Late/NFI-Up genes (Wnt7a, Nptx1, Nab2, Gabra1, Tesc, Rps6ka1) using ChIP. The 5′-flanking regions for each gene contained one or more consensus NFI sites, at least one of which showed detectable NFI occupancy (Fig. 3A). In each case, the timing of NFI binding mirrored temporal expression: binding was undetectable or low in 0 DIV cells consisting of premigratory CGNPs/CGNs derived from the EGL/PMZ (Raetzman and

Table 2. NFI/Temporal overlap genes with citations related to regulation of synaptic/dendritic development, synaptic function, and signaling

<table>
<thead>
<tr>
<th>Late/NFI-Up</th>
<th>Early/NFI-Down</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sor1</td>
<td>Chn6</td>
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<tr>
<td>Nrl9</td>
<td>Zdbhcl18</td>
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<td>Gas6</td>
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Hyperlinks to publications relevant to synapse and dendrite function, localization, regulation, signaling, or excitability are provided in the online version of the article. *See text.

Figure 3. NFI occupancy of Late/NFI-Up genes is temporally regulated. A, Left, Schematic of predicted NFI binding sites in the 5′-flanking regions of the indicated gene promoters. Open ovals, weak NFI binding sites; filled ovals, strong NFI binding sites. Arrows, position of primers used for ChIP qPCR. Right, Temporal NFI ChIP results. Pre, pre-immune serum; α-NFI, anti-xNFIB1 serum. Similar ChIP results were obtained in two or three experiments for each gene. B, Left, Characterization of the specificity of xNFIB1 antibody for expressed NFI family proteins on Western blots. Pan-NFI antibody staining of the same lanes is shown below. Right, NFI protein expression in nuclear extracts of immature and more differentiated CGN cultures detected with xNFIB1 antibody. Histone H3 (H3) served as a loading control. Data are representative of two independent experiments. C, ChIP assays showed that NFIA binds in a temporally delayed manner to Late/NFI-Up genes. Foxd1 genomic sequences served as a negative control. Identical patterns were observed in two independent experiments. D, ChIP/qPCR assays of P15 Nfia (+/−) (WT) and (−/−) (KO) mouse cerebella confirmed that occupancy of the Gabra6 NFI site as detected by xNFIB1 (α-NFI) and NFIA antibodies was dependent on NFIA. ***p < 0.001. Negative control antibody values were subtracted to yield specific antibody ChIP/qPCR signals. Data are representative of two independent experiments.
Figure 4. Regulation of the NFI switch program by membrane depolarization and CaN. qRT-PCR assays in A and B were performed for Late/NFI-Up genes (left) and Early/NFI-Down genes (right). A, Depolarization by 25 mM KCl culture medium (25) inhibits the NFI gene switch in 6 DIV CGNs, which is reversed by inhibitors of L-type VGCCs (nimodipine, Ni) and of CaN (FK506, FK). CGNs cultured in medium containing 5 mM KCl (5) or 25 mM KCl containing medium (5), which is reversed by FK506. Large fold-change values for Nptx1 and Egr1 are shown numerically. **p<0.001 except *p<0.05, ***p<0.01; ns, no significant difference. B, KA inhibits NFI switch gene expression in CGNs cultured to 6 DIV in 5 mM KCl-containing medium (5), which is reversed by FK506. Large fold-change values for Nptx1 and Egr1 are shown numerically. **p<0.001 except *p<0.05, ***p<0.01; ns, no significant difference. C, CaN activity is stimulated in depolarized CGN cultures. CGNs were cultured in medium containing either 5 mM KCl (5) or 25 mM KCl (25) ± FK506 to 6 DIV and CaN activity was determined in soluble cell extracts. Stimulation of phosphatase activity in depolarizing medium was prevented by the CaN inhibitor FK506. ***p<0.001. Data in A–C are representative of three independent experiments.

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Siegel, 1999) and in the P7 cerebellum composed mainly of immature postmigratory CGNs from the IGL (Wang et al., 2011), but was markedly elevated in more mature (6 DIV) CGN cultures and P21 mouse cerebellum. In contrast, the Efib1 gene, which is a direct NFI target (Wang et al., 2007) expressed at similar levels in immature and more differentiated CGNs (data not shown), exhibited constitutive NFI binding to its cognate promoter site (Fig. 3A). Delayed NFI occupancy was thus a common and specific temporal mechanism for many Late/NFI-Up genes in CGNs.

Temporal ChIP assays were performed with an anti-Xenopus NFIB1 (-xNFIB1) antibody, which recognized mammalian NFIB and NFIA, but not NFIX (Fig. 3B). Further, the abundance and mobility of NFI proteins detected by this antibody did not vary significantly with CGN maturation (Fig. 3B), as shown previously using pan-NFI antibodies (Wang et al., 2011). Since knock-out studies implicated NFIA in regulation of the switch program, we also confirmed its temporal occupancy for several Late/Up genes in developing CGNs and cerebellum (Fig. 3C). Further, we confirmed that ChIP signals for both anti-xNFIB1 and NFIA antibodies were markedly reduced in chromatin from P15 Nfia-null mouse cerebella (Fig. 3D). Thus, NFIA appears to be a direct regulator of the NFI temporal program in CGNs via its delayed occupancy of Late/NFI-Up genes.

Depolarization blocks NFI switch expression via activation of L-type Ca$^{2+}$ channels and CaN

A membrane potential-regulated developmental program was previously identified in which depolarization induced by elevated extracellular [K$^+$] inhibited the upregulation of numerous genes expressed in more mature CGNs and stimulated or maintained several genes expressed in immature CGNs (Sato et al., 2005). This voltage-sensitive pattern was similar to that of the NFI switch program, and comparison of the NFI- and KCl-temporal gene cohorts identified 19 commonly regulated temporal genes, including Gabra6, Gabra1, Gabrd, Camkk2, Wnt7a, Nptx1, Tiam1, Slek2a5, Kcnk3, Dcx, Neurod6, Tgfb2, and Cdh10. This suggested that at least a portion of the NFI switch program was sensitive to membrane potential in maturing CGNs. qRT-PCR analysis confirmed that numerous Late/NFI-Up genes were inhibited by 25 mM KCl in 6 DIV CGN cultures, while several Early/NFI-Down genes were elevated (Fig. 4A). This included several KCl-regulated genes that were not previously identified by Sato et al. (Nab2, Camkk4, Egr1, and Ect2). Several KCl-sensitive genes detected by Sato et al. were not regulated by NFI (e.g., Kcnal1, Tle1, and Prickle1) (data not shown). Not all temporal mRNAs examined segregated into these two KCl-regulatory/NFI-regulatory patterns; for example, transcripts for the genes in developing CGNs and cerebellum (Fig. 3C). Further, we confirmed that ChIP signals for both anti-xNFIB1 and NFIA antibodies were markedly reduced in chromatin from P15 Nfia-null mouse cerebella (Fig. 3D). Thus, NFIA appears to be a direct regulator of the NFI temporal program in CGNs via its delayed occupancy of Late/NFI-Up genes.

A similar switch pattern was observed in CGNs treated with KA (Fig. 4B), which also depolarizes CGNs, raises intracellular [Ca$^{2+}$] (Courtney et al., 1990), and inhibits maturation-dependent gene expression (Iijima et al., 2009) via activation of AMPA/kainate receptors. Thus, the NFI switch program is more generally sensitive to membrane depolarization-dependent mechanisms.

Elevated KCl increases steady-state Ca$^{2+}$ levels in mouse CGNs via L-type voltage-gated calcium channels (VGCCs) (Sato et al., 2005). Nimodipine, a specific inhibitor of L-type VGCCs, derepressed Late-Up gene expression in CGNs cultured in 25 mM KCl (Fig. 4A). In contrast, only one Early/NFI-Down gene, Dcx, showed some reversal of the stimulatory effects of depolarization in 6 DIV CGNs, indicating the involvement of additional mechanisms.
depolarization-sensitive mechanisms in the regulation of this gene subset. This was not further investigated here.

CaN activation has been implicated in depolarization-dependent regulation of numerous genes in CGNs (Guerrini et al., 2000; Ives et al., 2002; Kramer et al., 2003; Sato et al., 2005; Iijima et al., 2009). To address its role in the NFI switch, we first confirmed that CaN-dependent dephosphorylation activity was stimulated (~3-fold) in 6 DIV CGNs cultured in 25 mM KCl and that this activation was blocked by the CaN inhibitor FK506 (Fig. 4C). FK506 reversed inhibition by 25 mM KCl or KA for all Late-Up genes examined (Fig. 4A, B). Similar results were obtained with the CaN antagonist cyclosporine A (data not shown). In contrast to nimodipine, FK506 significantly suppressed depolarization-induced elevation of the Early-Up genes Dcx, Neurod6, and Tgfl2, consistent with earlier findings (Sato et al., 2005).

Membrane depolarization and CaN activation prevent NFI temporal binding

Since delayed occupancy is a central feature of the NFI switch program, we asked whether NFI binding to Late/NFI-Up genes was sensitive to K⁺ depolarization. NFI association with chromatin was repressed in 6 DIV CGNs cultured in 25 mM KCl for all six genes tested (Gabra6, Wnt7a, Nptx1, Tesc, Nab2, Gabra1) (Fig. 5). Further, these inhibitory effects were reversed by the CaN inhibitor FK506 (Fig. 5). Thus, temporal NFI occupancy was a primary target of depolarization-induced suppression of Late-Up genes mediated by CaN, including the GABA_A switch genes Gabra6 and Gabra1.

NFAT activation represses the NFI temporal switch

Previous studies found no evidence for differential phosphorylation of NFI proteins in maturing CGNs (Wang et al., 2011). Consistent with this, 25 mM KCl (+FK506) did not alter the amounts of nuclear NFI proteins or their electrophoretic mobilities in maturing CGN cultures (Fig. 6A). This result indicated that CaN activation likely modified other proteins that regulate the NFI switch. NFATc proteins are transcription factors that are regulated via CaN-dependent nuclear translocation (Nguyen and Di Giovanni, 2008), and NFATc4 nuclear localization is stimulated in depolarized neurons, including CGNs (Graef et al., 1999; Benedito et al., 2005). Using a lentiviral NFAT reporter, we confirmed that depolarization of CGNs stimulated NFAT transcriptional activity (Fig. 6B). We then examined the effects of blocking Ca²⁺/CaN-dependent NFATc nuclear translocation by expressing a dominant-negative form of NFATc4 (dnNFAT) (Chow et al., 1999). DnNFAT significantly reversed depolarization-dependent inhibition of nearly all Late-Up genes tested in CGNs cultured in 25 mM KCl (Fig. 7). Similarly, dnNFAT prevented upregulation of Early-Down genes by 25 mM KCl with the exception of the noncanonical Ect2 gene, which was unaffected (Fig. 7). Thus, suppression of the NFI switch program by depolarization was largely dependent on activation of endogenous NFAT proteins.

Since nuclear translocation of NFATc4 is stimulated in depolarized neurons, we tested the effects of FLAG-tagged NFATc4 (NFATc4-Ala) that constitutively translocates to the nucleus and regulates transcription (Yang et al., 2002). In CGNs maturing in physiological medium (5 mM KCl), NFATc4-Ala strongly repressed all Late-Up genes examined (Fig. 7). Further, the Early-Down gene Dcx was upregulated by NFATc4-Ala while Neurod6 and Tgfl2 were inhibited (Fig. 7), suggesting direct or indirect repressive effects on the latter genes due to constitutive NFATc activity. Thus, enhanced NFATc activity in nondepolarized CGNs reproduced many of the effects of 25 mM KCl on NFI switch program expression.

**Figure 5.** NFI temporal occupancy is suppressed by depolarization in a CaN-dependent manner. CGNs were cultured in 5 mM or 25 mM KCl ± FK506 until 6 DIV and then assayed for NFI occupancy of the indicated Late/NFI-Up genes using ChIP qPCR. Negative control antibody signals were subtracted from ChIP qPCR values in each case. KCl (25) blocked the temporal increase in NFI binding, and CaN antagonism using FK506 reversed this inhibitory effect; 5 mM KCl; 25, 25 mM KCl; FK, FK506; 25 versus 5: ***p < 0.001; **p < 0.01; (25 + FK) versus 25: #p < 0.05; ##p < 0.01; ####p < 0.001. Data are representative of three independent experiments.

**Figure 6.** Depolarizing conditions have no effect on NFI proteins but stimulate NFAT transcriptional activity via CaN. A, Western blot of NFI proteins in 6 DIV CGNs cultured in 5 mM KCl (5) or in 25 mM KCl (25 ± the CaN inhibitor FK506. Histone H3 (H3) served as a loading control. B, CGNs were transduced with NFAT dual-reporter lentivirus on 0 DIV and cultured as in A to 6 DIV. NFAT transcriptional activity was then determined in cell extracts by assaying luciferase activity. ***p < 0.001. Data are representative of either two (A) or three (B) independent experiments.

**Figure 7.** NFAT proteins regulate NFI switch gene expression. CGNs were transduced on 0 DIV with GFP or NFATc4-Ala lentiviruses and cultured in 5 mM KCl medium (5), or were transduced with GFP- or dnNFAT-expressing lentiviruses and cultured in depolarizing (25 mM KCl-containing) medium (25). Cells were harvested on 6 DIV and assayed by qRT-PCR. Late/NFI-Up genes are shown on the left, Early/NFI-Down genes on the right. p < 0.001 except *p < 0.05, **p < 0.01; ns, no significant difference. Data are representative of three independent experiments.
NFAT proteins control NFI occupancy in depolarized CGNs

Since depolarization inhibits NFI temporal interactions with chromatin, we examined whether NFAT proteins specifically regulated NFI binding of Late/NFI-Up genes. We focused on four genes showing the most robust derepression by dnNFAT (Gabra6, Wnt7a, Nptx1, Tesc). dnNFAT expression reversed the inhibitory effects of depolarizing medium on NFI occupancy in each case (Fig. 8A). Conversely, constitutively active NFATc4-Ala inhibited NFI binding to these genes in CGNs cultured in nondepolarizing (5 mM KCl) medium (Fig. 8A).

The actions of elevated NFAT activity on NFI occupancy may have been direct or indirect. Sequence analysis identified multiple consensus NFAT binding sites near the regions of NFI occupancy within the four Late-Up gene promoters, suggesting possible direct NFAT interactions (Fig. 8B). To test this, CGN cultures were transduced with FLAG-tagged NFATc4-Ala and assayed by ChIP on 6 DIV. Elevated FLAG signal was detected within specific NFAT-site regions for all four promoters (Fig. 8C). Similarly, endogenous NFATc4 occupancy of Late/NFI-Up genes was markedly increased in nontransduced 6 DIV CGNs cultured in 25 mM KCl (Fig. 8D). Thus, NFATc4 occupancy was elevated under conditions of reduced NFI occupancy of Late/NFI-Up genes, consistent with a direct inhibitory effect of this factor on the NFI switch program in depolarized CGNs.

Endogenous CaN and NFAT proteins regulate dendrite formation in nondepolarized CGNs

Maturation-dependent reduction of CaN activity has been proposed to promote the differentiation of postmigratory CGNs within the developing postnatal IGL (Nakanishi and Okazawa, 2006). However, a direct test of CaN function in CGNs differentiating under normal, nondepolarizing circumstances was not previously addressed, to our knowledge. Consistent with this hypothesis, CaN activity declined as CGNs matured in physiological culture medium (Fig. 9B). Further, FK506 treatment increased dendrite extension by immature CGNs in 5 mM KCl cultures (Fig. 9C). Similarly, dn-NFAT enhanced dendritic length in CGNs maturing in physiological medium (Fig. 9C). Thus, endogenous CaN and NFAT proteins repress dendrite emergence during onset of differentiation by immature CGNs.

CaN and NFATc4 regulate switch expression and NFI occupancy during normal CGN differentiation

Based on the above findings, we examined whether CaN-dependent mechanisms also regulated the NFI switch program in
CGNs maturing under physiological conditions. Treatment of CGN cultures with FK506 in medium containing 5 mM KCl enhanced Late/NFI-Up gene expression and decreased transcripts for the Early/NFI-Down genes Dcx and Tgfb2 (Fig. 10A). Switch-enhancing effects were observed in both less mature (3 DIV) and more mature (6 DIV) cultures, although they were greater at 3 DIV for several genes, consistent with elevated CaN activity in immature CGNs. These results suggested an inhibitory function for CaN on the NFI switch program during normal differentiation of CGNs.

To examine the role of endogenous NFAT proteins in regulating the NFI switch, we first determined whether endogenous NFATc4 occupied Late/NFI-Up gene promoter sites in CGNs maturing under physiological conditions. Robust NFATc4 binding was detected for the Gabra6, Wnt7a, Nptx1, and Tesc promoters in 0 DIV CGNs derived from the EGL/PMZ, as well as in the P7 cerebellum composed mainly of immature CGNs in the IGL (Wang et al., 2011) (Figs. 8D, 10B). Further, NFATc4 occupancy of each site markedly declined in more differentiated, 6 DIV CGN cultures and in the P15 cerebellum (Figs. 8D, 10B), consistent with an inhibitory effect on Late/NFI-Up temporal expression in immature CGNs.

To address NFATc4 function more directly, we examined the impact of NFATc4 deficiency on expression of the NFI temporal program. Cerebella from Nfatc4-null mice were examined at P7–P9, when expression of the NFI switch, as well as at P15 when NFATc4 occupancy is low and the switch program is highly expressed. Expression of numerous Late/NFI-Up genes was significantly upregulated in Nfatc4-null cerebella at each age (Fig. 10C). Further, downregulation of the Early/NFI-Down gene Dcx was enhanced during this period. In contrast, Rbfox3/NeuN, which is upregulated as CGNs mature (Weyer and Schilling, 2003) and is not regulated by NFI (data not shown), was not altered by NFATc4 deficiency (Fig. 10C). Thus, enhanced expression of the switch program in the developing Nfatc4-null cerebellum was not due to global changes in maturation-associated gene expression. Western analysis also confirmed that NFI protein levels and mobility were unchanged in postnatal Nfatc4 (−/−) cerebellum (data not shown).

The pattern of NFATc4 temporal occupancy of Late/NFI-Up genes inversely mirrored that for CaN binding (Figs. 3A, 10B). ChIP analysis revealed that NFAT binding to the four Late/NFI-Up promoters increased severalfold in P9 Nfatc4 (−/−) cerebellum relative to WT, and remained significantly elevated at P15 (Fig. 10D). Thus, endogenous NFATc4 inhibits NFI occupancy of Late/NFI-Up genes in the developing postnatal mouse cerebellum.

**Discussion**

The GABA<sub>A</sub> receptor switch and formation of inhibitory synapses between CGNs and GABAergic Golgi type II neurons are critical for cerebellar information flow (Hamann et al., 2002), and their completion coincides with eye opening and enhanced motor activity in rodents (Tia et al., 1996). Thus, the timing of these events is important for developmental onset of sensorimotor function. The present studies reveal a central role for NFI in regulating a late developmental switch program in CGNs that becomes active during a period (P7–P15) dominated by dendritic and synaptic maturation within the IGL. In this process, genes required for mature GABA<sub>A</sub> receptor function and dendrite and synapse formation are upregulated and “immature” genes are downregulated. Delayed NFI occupancy of late-expressed genes is a key feature of this program, coinciding with upregulation of Late/NFI-Up gene expression.

Membrane depolarization is frequently linked with the promotion of neuronal maturation. For example, depolarizing actions of GABA on immature neurons are thought to play a key role in the regulation of neuronal maturation. In this study, we found that CaN activity was increased in immature CGNs, and that this activity was decreased in more mature CGNs, consistent with elevated CaN activity in immature CGNs. However, the CaN activity was not regulated by NFI in immature CGNs, as evidenced by the increased CaN activity in P7–P9 and P15 cerebella, and the lack of regulation by NFI in immature CGNs. These results suggest that CaN activity is not regulated by NFI in immature CGNs, and that this activity is not required for the regulation of gene expression in immature CGNs.

However, in more mature CGNs, CaN activity was decreased and regulated by NFI. This suggests that CaN activity is regulated by NFI in more mature CGNs, and that this activity is required for the regulation of gene expression in more mature CGNs. These results support the hypothesis that CaN activity is regulated by NFI in more mature CGNs, and that this activity is required for the regulation of gene expression in more mature CGNs. This regulation of CaN activity by NFI may be important for the regulation of gene expression in more mature CGNs, and may be important for the regulation of neuronal maturation.
role in corticogenesis and neuronal differentiation (Wang and Kriegstein, 2009; Kilb et al., 2011). However, depolarizing conditions can also block differentiation of neurons, particularly late events linked to dendritic morphology and synaptic development (Sohya et al., 2007; Okazawa et al., 2009). Inhibition of mouse CGN maturation by depolarization-induced CaN activation has been well documented (Guerini et al., 2000; Ives et al., 2002; Kramer et al., 2003; Sato et al., 2005; Iijima et al., 2009; Okazawa et al., 2009). Our studies reveal that the NFI switch program and NFI temporal occupancy are critical targets linking depolarization and CaN activation via L-type VGCCs to inhibition of dendrite formation in CGNs, with NFATc4 being an important intermediary regulator (Fig. 11). Further, these voltage-sensitive mechanisms regulate the NFI switch program and dendrite formation in CGNs differentiating under normal, nondepolarizing conditions.

Our present findings suggest a model in which CaN/NFATc4 inhibit the NFI switch program during development in vivo, delaying and modulating the magnitude of a significant portion of its expression in immature postmigratory CGNs in the IGL (Fig. 11). Declining CaN activity and NFATc4 occupancy as CGNs differentiate leads to derepression and is permissive for activation of numerous late-maturation genes associated with dendrite formation, as well as suppression of Early/NFI-Down genes. Downregulation of Early/NFI-Down genes is apparently indirect via undefined repressive factors or signaling pathways that are stimulated by NFI trans-activation. Developmental changes in resting membrane potential, which becomes more hyperpolarized as CGNs mature (Rossi et al., 1998; Cathala et al., 2003; Okazawa et al., 2009), appear to regulate late CGN maturation and dendrite formation via Ca\(^{2+}\)-dependent regulation of CaN (Nakanishi and Okazawa, 2006; Okazawa et al., 2009). Extrinsinc and intrinsic

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**Figure 10.** Regulation of the NFI switch by CaN and NFATc4 in CGNs differentiating in culture and in vivo. **A**, Differential expression of Late/NFI-Up genes in standard (5 mM KCl) culture medium with and without FK506. CGN cultures were assayed at 3 (open bars) and 6 (filled bars) DIV. Positive values indicate upregulation and negative values indicate downregulation. **B**, NFATc4 temporal occupancy of Late/NFI-Up genes declines in the developing cerebellum. ChIP qPCR analysis of NFAT binding sites (Fig. 8B) was performed for P7 and P15 mouse cerebellum. **C**, Differential expression of NFI switch genes determined by qRT-PCR in Nfatc4-null (C4KO) and WT mouse cerebellum. Rbfox3 (NeuN) served as a non-NFI regulated negative control. **D**, NFI occupancy was determined by ChIP qPCR in Nfatc4-deficient (C4KO) and WT cerebellum at P9 and P15. Negative control antibody values were subtracted to show specific binding. No significant differences except \(* p < 0.05; ** p < 0.01; *** p < 0.001\). Data are representative of three (A, C) or two (B, D) independent experiments.
mechanisms that promote hyperpolarization of maturing CGNs may therefore be important upstream drivers of NFI temporal occupancy and the NFI switch within the IGL via CaN/NFATc4 (Fig. 11). An interesting feature of the NFI temporal program is the regulation of several ion channel, transporter and receptor genes that control membrane potential and excitability, including Slc12a5/Kcc2, Kcnk3/Task1, Kcnab1/Kb1.3, and GABA<sub>A</sub> receptor subunits. This raises the possibility that NFI temporal programming and CGN excitability may reinforce each other during development, forming a self-amplifying positive feedback loop. Similar mechanisms also may operate in other neurons that undergo developmental membrane hyperpolarization (Ramoa and McCormick, 1994; Zhou and Hablitz, 1996; Tyzio et al., 2003).

The timing of neurogenesis has been linked to the formation of discrete patterns of synaptic circuitry for different neuronal subtypes (Trippodi and Arber, 2012). Thus, molecular mechanisms initiated during neuronal birth appear to contribute significantly to the timing of synaptogenesis and circuit formation. In the context of CGN development, postnatal CGNP transplantation experiments showed that maturation-dependent expression of the NFI switch gene <i>Gabra6</i> was independent of in vivo developmental and regional cues (Bahn et al., 1999). Similarly, <i>Gabra6</i> gene expression is maintained in CGNs of HNF3β transgenic mice that fail to properly migrate to the IGL (Zhou et al., 2001). These and other findings suggest that intrinsic timing mechanisms initiated during CGNP cell cycle exit trigger dendritogenesis-associated gene expression in maturing CGNs (Mellor et al., 1998; Bahn et al., 1999). NFI temporal occupancy and its regulation by developmental changes in CaN activity and NFATc4 binding may therefore be part of an internal timing mechanism contributing to dendritic development and its associated gene expression in CGNs. Modulation of these timing mechanisms by environmental factors also may be important for appropriate magnitude and temporality of NFI switch gene expression.

NFI proteins function at multiple stages of CGN development (Wang et al., 2007), and a key question is how they regulate such a diversity of events occurring over varying time frames. The present studies provide insight into their regulation of gene expression specifically required for postmitotic CGN maturation via delayed promoter occupancy. Since NFI proteins and their nuclear DNA binding activity are not temporally upregulated in maturing CGNs (Wang et al., 2011), additional transcriptional and chromatin-related factors presumably contribute to the specificity and timing of their binding and regulation of Late/NFI-Up genes. NFATc4 appears to be an important direct regulator of these events: it binds to numerous Late/NFI-Up gene promoters in immature CGNs, and it is required to prevent premature and excessive NFI occupancy in the developing postnatal cerebellum. The trans-repressor REST was recently shown to occupy the <i>Gabra6</i> promoter and to inhibit its binding by NFI in CGPNs and premigratory CGNs derived from the EGL/PMZ (Wang et al., 2011). REST occupancy is depleted in the P7 cerebellum when NFI binding remains low, indicating additional mechanisms function to prevent premature NFI binding in CGNs within the immature IGL. Our findings implicate NFATc4 in the continued repression of NFI occupancy of the <i>Gabra6</i> gene in immature postmitotic CGNs in the early postnatal (e.g., P7) IGL following REST dismissal. NFATc4 also limits the magnitude of NFI occupancy of multiple Late/NFI-Up promoters as well as switch program expression in more mature CGNs at P15 (Fig. 10C,D). NFATc4 also may repress Late/NFI-Up genes (and possibly enhance Early/NFI-Down gene expression) via additional, NFI-independent mechanisms.

NFATc4 was previously implicated in neuronal developmental and function, including axonogenesis and survival, neurotrophin signaling, and memory formation in mice (Graef et al., 2003; Nguyen et al., 2009; Vashishta et al., 2009; Quadrato et al., 2012). Our studies define a novel role for NFATc4 in late CNS development via its temporally downregulated chromatin occupancy. They also identify numerous NFATc4 neuronal targets involved in dendrite/synapse formation, implicating this factor as an important direct regulator of late-maturation gene expression in CGNs.

<i>Nfia</i> knock-out mice exhibit severe neurological disruptions, including tremor and diminished cerebellar foliation consistent with altered cerebellar function (das Neves et al., 1999; Wang et al., 2007). Dendrite and synapse formation within the IGL are also altered in <i>Nfia</i> (<i>/−</i>) mice (Wang et al., 2007, 2010), and as
shown here NFIA occupancy of Late/NFI-Up genes is temporally regulated. Further, development of both CGNs and cortical pyramidal neurons is delayed in Nfia-null mice (Wang et al., 2007; Piper et al., 2010), consistent with a more general temporal function for this protein in maturing neurons. It is noteworthy that patients with NFIA haploinsufficiency exhibit developmental delays in motor skills and language (Lu et al., 2007; Shaikh et al., 2011), both of which involve cerebellar function (Strick et al., 2009). NFIA and other NFI family members may therefore control important timing mechanisms in CGNs that are involved in developmental delay in both humans and mice.

Formation of distinct patterns of neuronal connectivity has been linked to specific temporal schedules of synaptogenesis (Deguchi et al., 2011; Tripodi and Arber, 2012). Thus, the timing of presynaptic and postsynaptic maturation is a key developmental step in establishing the complexity of neural circuitry that characterizes CNS structure and function. This requires appropriate temporal regulation of gene expression involved in dendritic and synaptic maturation, and disrupted temporal expression can dramatically alter synaptic circuitry (Hippenmeyer et al., 2005; Petrovic and Hummel, 2008). Altered expression of temporally regulated genes during critical time windows affecting synaptic maturation has been proposed to underlie several neurodevelopmental disorders (Meredith et al., 2012). The molecular mechanisms that control these temporal synaptic events are largely unknown. The present studies provide insight into the importance of NFI proteins in the onset of dendrite-related gene expression via their temporally regulated occupancy of synapse-related target genes. An interesting possibility is that altered NFI switch programming and NF temporal occupancy contribute to disrupted timing of dendrite and synapse formation associated with one or more neurodevelopmental disorders.

References


