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Direct interactions promote eviction of the Sir3 heterochromatin protein by the SWI/SNF chromatin remodeling enzyme

Benjamin J. Manning and Craig L. Peterson

Heterochromatin is a specialized chromatin structure that is central to eukaryotic transcriptional regulation and genome stability. Despite its globally repressive role, heterochromatin must also be dynamic, allowing for its repair and replication. In budding yeast, heterochromatin formation requires silent information regulators (Sirs) Sir2p, Sir3p, and Sir4p, and these Sir proteins create specialized chromatin structures at telomeres and silent mating-type loci. Previously, we found that the SWI/SNF chromatin remodeling enzyme can catalyze the ATP-dependent eviction of Sir3p from reconstituted nucleosomal arrays, and this activity enhances early steps of recombinational repair in vitro. Here, we show that the ATPase subunit of SWI/SNF, Swi2p/Snf2p, interacts with the heterochromatin structural protein Sir3p. Two interaction surfaces are defined, including an interaction between the ATPase domain of Swi2p and the nucleosome binding, Bromo-Adjacent-Homology domain of Sir3p. A SWI/SNF complex harboring a Swi2p subunit that lacks this Sir3p interaction surface is unable to evict Sir3p from nucleosomes, even though its ATPase and remodeling activities are intact. In addition, we find that the interaction between Swi2p and Sir3p is key for SWI/SNF to promote resistance to replication stress in vivo and for establishment of heterochromatin at telomeres.

BAH | SWI/SNF | chromatin remodeling | Sir3 | heterochromatin

All eukaryotic genomes are stored within the nucleoprotein structure of chromatin, the core subunit of which, the nucleosome, consists of 147 base pairs (bp) of DNA wrapped ∼1.7 times around an octamer of histone proteins (1). Over millions of years, eukaryotes have incorporated chromatin structure into the regulation of many aspects of DNA metabolism, from simple nuclear packaging to transcriptional control (2). This diversity of purpose is reflected in two general types of chromatin structures within the nucleus— euchromatin, which is decondensed and transcriptionally active, and heterochromatin, which is typically localized to the nuclear periphery and repressive for DNA recombination and transcription. Heterochromatin structures are commonly associated with centromeres and telomeres, and these domains package much of a genome’s repetitive DNA (3). Consequently, the maintenance of heterochromatin is key for genomic integrity, because it prevents illicit recombination among DNA repeats and promotes chromosome segregation during mitosis (4, 5).

On a molecular level, heterochromatic loci are marked by specific chromatin posttranslational modifications, which are recognized and bound by characteristic nonhistone proteins. In many vertebrates, heterochromatin is characterized by members of the heterochromatin protein 1 (HP1) family of proteins, whereas in budding yeast, the silent information regulator (Sir) proteins, Sir2p, Sir3p, and Sir4p, create heterochromatin structures at telomeres and the silent mating-type loci (6, 7). Sir3p is believed to be the key structural component of yeast heterochromatin—Sir3p contains numerous protein–protein interaction motifs (8–10), including an N-terminal Bromo-Adjacent Homology (BAH) domain that interacts with the nucleosomal surface (11–13). BAH domains are found in many other chromatin-associated factors, including the Rsc2p subunit of the remodels structure of chromatin (RSC) remodeling enzyme and the Orc1p subunit of the Origin Recognition Complex (ORC) (14). The stability of the Sir3p BAH–nucleosome complex requires deacetylated histone H4 lysine 16 (15); consequently, amino acid substitutions at H4-K16 disrupt Sir3p–nucleosome binding and eliminate heterochromatin assembly in vivo (15–17).

Despite the repressive structure of heterochromatin, these domains must be replicated and repaired, implying that mechanisms exist to regulate heterochromatin disassembly. Previously, we described an in vitro assay to monitor early steps of recombinational repair with reconstituent nucleosomal array substrates (18). Whereas the repair machinery was not hindered by the simple presence of nucleosomes, we reported that the binding of the Sir proteins, or even Sir3p by itself, led to dramatic repression of recombinational repair events on nucleosomal arrays (18, 19). Surprisingly, we discovered that the ATP-dependent chromatin remodeling enzyme, SWI/SNF, was able to counteract these repressive effects of heterochromatin in vitro, stimulating early steps of homologous recombination. Intriguingly, these assays uncovered that SWI/SNF catalyzed the ATP-dependent eviction of Sir3p from nucleosomes, an activity not shared by several other remodeling enzymes (19). Thus, these studies suggested that the SWI/SNF enzyme may have a unique ability to disrupt heterochromatin structures.

In this work, we identify a physical interaction between SWI/SNF and the heterochromatin protein Sir3p. We identify a pair of interactions—between the Swi2p Helicase SANT Adjacent HSA domain and the Sir3p AAA+ domain and between the Swi2p ATPase domain and the Sir3p BAH domain. Surprisingly, the ATPase–BAH interaction is conserved between many Swi2p/Snf2p ATPase family members and between two classes of BAH domains, suggesting a common mode of binding between these domains. Mutations are generated that ablate the interaction between Swi2p and Sir3p, and we find that the Swi2p–Sir3p interaction surfaces are required for SWI/SNF to evict Sir3p from nucleosomal arrays in vitro. Furthermore, in vivo studies indicate that SWI/SNF–Sir3p interactions are important both for

Significance

Heterochromatin is a repressive mode of genetic storage that prevents cellular machineries from accessing DNA sequences. Here, we investigate how a protein machine, called SWI/SNF, can disrupt these heterochromatin structures and facilitate nuclear processes. We identify physical and functional interactions between SWI/SNF and a key heterochromatin protein, silent information regulator 3p (Sir3p).

Author contributions: B.J.M. and C.L.P. designed research; B.J.M. performed research; B.J.M. and C.L.P. analyzed data; and B.J.M. and C.L.P. wrote the paper.

The authors declare no conflict of interest.

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resistance to replication stress and for establishment of silenced heterochromatic domains.

**Results**

**SWI/SNF Interacts with Sir3p.** To investigate the unique ability of SWI/SNF to displace Sir3p from nucleosomes, we began by asking whether SWI/SNF and Sir3p physically interact. First, Sir3p–FLAG was affinity-purified from yeast and immobilized on anti-FLAG antibody resin. Purified SWI/SNF and RSC remodeling enzymes were incubated with Sir3p-bound beads, and bound and free fractions were analyzed by Western blotting (Fig. 1A). Strikingly, SWI/SNF, but not the highly related RSC complex, was able to interact with bead-bound Sir3p (Fig. 1A). This interaction was also apparent if SWI/SNF was immobilized on beads and incubated with purified Sir3p (Fig. S1A). To confirm the interaction and to gain insight into which SWI/SNF subunit might be involved, we used far Western analysis. Purified SWI/SNF, RSC, and Sir2p/Sir4p complexes were separated on an SDS/PAGE gel and transferred to a membrane. The membrane was incubated in buffer to stimulate protein renaturation and then incubated with purified Sir3p (Fig. 1B). Proteins bound to Sir3p were then detected by Western blotting, using antisera to Sir3p.

As expected, Sir3p interacted strongly with Sir4p in this assay, but little interaction was detected with subunits of RSC (Fig. 1B). However, two polypeptides were then detected by Western blotting, using antisera to Sir3p. The largest species comigrated with the Swi2p ATPase domain bound the Swi2p ATPase domain identified a 49-amino-acid fragment within the first RecA-like fold that retained Sir3p binding activity (Fig. 1C). Interestingly, these domains were found to interact in a pairwise manner—the Swi2p ATPase domain bound the Sir3p BAH domain, and the Swi2p HSA domain bound the Sir3p AAA⁺ domain (Fig. 2C). Progressive N- and C-terminal truncations of the GST–HSA fusion protein (Fig. S1D) defined a region of 10 amino acids in the Swi2p HSA domain that is required for interaction with Sir3p (Fig. 2D). Furthermore, deletion of the Swi2p ATPase domain identified a 49-amino-acid fragment within the first RecA-like fold that retained Sir3p binding activity (Fig. 2C). Interestingly, these analogous residues from the ATPase domain of the Sir3p catalytic subunit, Sth1p, were unable to bind to Sir3p (Fig. 2D).

**SWI/SNF and RSC Interact with Core BAH Domains.** To test whether a BAH core domain might generally be sufficient for interaction with SWI/SNF-like enzymes, the core BAH domain of Rsc2p was assayed for interactions. Indeed, both SWI/SNF and RSC interacted well with the Rsc2p BAH core domain; however, inclusion of the conserved C-terminal (CT-1) domain eliminated interactions with both SWI/SNF and RSC (21). Furthermore, SWI/SNF also bound to the BAH domain from Orc1p, a subunit of the ORC (Fig. 3B). The RSC remodeling enzyme was also able to bind to the Orc1p BAH, despite being unable to interact with Sir3p BAH. Both SWI/SNF and RSC were also competent to bind to the human ORC1 BAH (Fig. S1D). In contrast, the Swi2p remodeling enzyme did not interact at detectable levels with either the Sir3p or Orc1p BAH domain, suggesting that BAH interactions may be a general feature of only the SWI/SNF subfamily of chromatin-remodeling enzymes (Fig. 3B). These data also suggest that sequences C-terminal to BAH core domains may govern the specificity of remodeling enzyme interactions.

**Swi2/Snf2–Sir3p Interactions Are Required for Sir3p Eviction in Vitro.** Having identified Sir3p-interaction domains within Swi2p, we asked whether they were required for the ATP-dependent eviction of Sir3p by SWI/SNF. To this end, a 10-amino-acid deletion within the HSA domain (Δ10) as well as a 197-amino-acid swap between the Sth1p and Swi2p ATPase domains (Sth1[R]) (termed swi2Δ–10R; Fig. 2D). This region of Sth1p encompasses the first RecA-like lobe of the ATPase domain. This region is nearly homologous to that of Swi2p, with the exception of a central, 52-amino-acid divergent region. A C-terminal, TAP-tagged version of Swi2Δ–10R was then expressed in yeast from its normal promoter on a low-copy CEN/ARS plasmid, and SWI/SNF–Δ10R complexes that harbors Swi2pΔ–10R was isolated by tandem affinity purification. The concentration of active enzyme was determined by ATPase assays, and equal ATPase units of wild-type and SWI/SNF–Δ10R complexes were analyzed by SDS/PAGE and silver staining. The subunit composition of the SWI/SNF–Δ10R complex...
was nearly identical to that of wild-type SWI/SNF, with the exception of an approximately twofold depletion of the Arp7p and Arp9p subunits (Fig. 4A). Because Arp subunits have been implicated in the regulation of ATPase kinetic parameters (22), we characterized the ATPase activity of the SWI/SNF–Δ10R complex. Importantly, the SWI/SNF–Δ10R complex exhibited kinetic parameters for DNA-stimulated ATPase activity indistinguishable from the wild-type complex (Fig. 4B).

The activity of the SWI/SNF–Δ10R complex was also monitored in several chromatin-remodeling assays. First, equal ATPase units of wild-type and SWI/SNF–Δ10R complexes were incubated with mononucleosomes positioned in the center of a radiolabeled 282-bp DNA fragment by a 601-nucleosome positioning sequence. The ATP-dependent movement of the nucleosome toward the DNA ends leads to faster mobility on native PAGE, and in this assay, the SWI/SNF–Δ10R enzyme showed activity equivalent to wild type (Fig. 4C). Chromatin remodeling was also assessed by a nucleosomal array accessibility assay (23). This quantitative assay uses a positioned array of 11 nucleosomes, where the central nucleosome of the array occludes a unique Sall restriction enzyme recognition site. As the array is remodeled by SWI/SNF, this central nucleosome is repositioned or removed, increasing the rate of Sall cleavage. Similar to the ATPase and mononucleosome remodeling assays, the SWI/SNF–Δ10R enzyme showed equivalent activity compared with the wild-type complex (Fig. 4D).

Finally, we assayed the ability of the SWI/SNF–Δ10R enzyme to catalyze the ATP-dependent eviction of Sir3p protein from nucleosomal arrays (Fig. 4D). In this assay, 12-mer nucleosomal arrays were assembled with recombinant histone octamers, and ∼15% of the octamers contained histone H2A biotinylated at an engineered cysteine within the exposed C-terminal domain (18). Purified Sir3p protein was bound to these arrays at a ratio of two Sir3p monomers per nucleosome (24) and then incubated with chromatin-remodeling enzyme in the presence of ATP. Reactions were captured on streptavidin-coated magnetic beads, and bound and unbound (U) fractions were subjected to Western blotting, probing for both histone H3 and Sir3p. In these reactions, wild-type SWI/SNF was able to evoke ∼35% of the Sir3p into the unbound fraction, whereas the SWI/SNF–Δ10R complex was defective at Sir3p eviction (Fig. 4E). Indeed, the SWI/SNF–Δ10R complex resembled the activity of RSC, in which the central nucleosome of the array occludes a unique SalI restriction enzyme recognition site. As the array is remodeled by SWI/SNF, this central nucleosome is repositioned or removed, increasing the rate of SalI cleavage. Similar to the ATPase and mononucleosome remodeling reactions, wild-type SWI/SNF was able to evict Sir3p into the unbound fraction, whereas the SWI/SNF–Δ10R complex showed a marked sensitivity to the replication stress agent hydroxyurea (HU; Fig. 5A). Previous studies have suggested that the HU⁺ phenotype of swi/snf mutants may be due to a defect in transcriptional induction of ribonucleotide reductase (RNR) genes (26); however, the swi2–Δ10R strain exhibited wild-type levels of RNR3 transcriptional induction (Fig. S3A).

Indeed, no significant changes in RNA expression were observed between wild-type and swi2–Δ10R strains when assayed by RNA sequencing (RNA-seq) (Fig. S3 B and C and Dataset S1). Consistent with previous work (27), swi2–Δ10R did not affect sir2 or sir3 expression (Fig. S3D and Dataset S1). Interestingly, the HU⁺ phenotype of the swi2–Δ10R strain was suppressed by deletion of Sir3, consistent with a functional interaction between SWI/SNF and Sir3p during replication stress.

To test whether SWI/SNF regulates the dynamics of heterochromatin assembly, wild-type and swi2–Δ10R strains were assayed in a transcriptional silencing establishment assay (28). This assay was performed in strains with a URA3 gene integrated adjacent to the telomere on right arm of chromosome V (TELVR::URA3). In this location, URA3 expression is repressed by the spreading of adjacent subtelomeric heterochromatin, creating a biphasic population of Ura⁺ and Ura⁻ cells. To monitor the establishment of the silenced state, cells were first grown in medium lacking uracil, to enrich for cells in which URA3 is in the ON state (Ura⁺). Cells were then grown in the presence of uracil for increasing time and then plated onto plates that contain 5-fluoroorotic acid (5-FOA), scoring for cells that have silenced URA3 (Ura⁻). Compared with the wild type, the swi2–Δ10R mutant had a delayed onset of silencing and achieved a lower final level of silencing (Fig. 5D).

Furthermore, the swi2–Δ10R strain formed much smaller colonies, suggesting that silencing was inherited less stably (Fig. 5D). Thus, these results suggest that interactions between SWI/SNF and Sir3p impact heterochromatin dynamics in vivo.

**SWI/SNF Is Not Required for Heterochromatic Recombinational Repair.** Yeast mating-type switching requires that a double-strand break (DSB) induced at the MAT locus is repaired by homologous recombination with sequences from a heterochromatic HM locus (29). Previously, in vivo studies suggested that SWI/SNF is essential for mating-type switching and that SWI/SNF promotes repair only when the donor sequences are heterochromatic (19, 30). As an initial test for whether the swi2–Δ10R allele impacts heterochromatic mating-type switching, a plasmid expressing a galactose-inducible homothallic (HO) endonuclease was introduced into isogenic wild-type, swi2Δ, and swi2–Δ10R strains. The strand-invasion step of mating-type switching was then assayed by a PCR-based assay following a switch to galactose medium.
nor...domain, and an N-terminal portion of the Swi2p strains were competent to switch HM strains showed swi2 locus. In these assays, they reported that Manning and Peterson strain, de- strain was created by locus. The swi2 www.pnas.org/cgi/doi/10.1073/pnas.1420096111

Discussion
Here, we have defined two distinct protein–protein interfaces between the Sir3p heterochromatin protein and the Swi2p sub-unit of the SWI/SNF chromatin remodeling enzyme. The HSA domain from Swi2p interacts with a region of Sir3p that contains its AAA domain, and an N-terminal portion of the Swi2p ATPase domain interacts with the nucleosome-binding, BAH domain of Sir3p. Intriguingly, Sth1p, the related ATPase from the RSC remodeling enzyme, can also bind to the Sir3p BAH domain, but only after elimination of flanking sequence elements. Furthermore, both Swi2p and Sth1p are able to bind to the central core of the Rsc2p and Orc1p BAH domains, suggesting that SWI/SNF-like ATPase domains may harbor a general affinity for BAH domains. Importantly, elimination of Sir3p interaction surfaces within Swi2p (Swi2p Δ10R) disrupts the ability of SWI/SNF to catalyze the ATP-dependent eviction of Sir3p from nucleosomal arrays in vitro, without impairing its ATPase or more canonical chromatin-remodeling activities. Furthermore, these alterations led to specific phenotypes in vivo, consistent with functional interactions between SWI/SNF and Sir3p-dependent heterochromatin structures.

What is the functional role for Sir3p eviction by SWI/SNF? A previous study from Laurent and colleagues (30) was consistent with this activity playing an essential role in recombinational repair events that involve heterochromatin. Specifically, they used strains harboring a galactose-inducible HO endonuclease to create a single DNA DSB at the euchromatic MAT locus. The recombinational repair of this DSB requires a successful homology search and strand invasion of a homologous, but heterochromatic, HM locus. In these assays, they reported that inactivation of the Swi5p subunit of SWI/SNF had no effect on early steps of HR, but that swi5Δ eliminated capture of the heterochromatic donor sequences, and repair was blocked (30). Subsequently, we showed that SWI/SNF is not required for

(31). Surprisingly, neither the swi2Δ10R nor swi2Δ strains showed a significant defect in strand invasion (Fig. S4A).

To confirm this observation, a swi2Δ strain was created by tetrad dissection in a strain harboring a chromosomal, galactose-inducible HO gene. Notably, this is the same background as used in previous studies (30). Multiple swi2Δ segregants from independently created diploids showed severe growth defects (Fig. S4B) and delayed galactose induction kinetics that precluded kinetic analyses of strand invasion. However, after growth for 4 h in galactose medium, swi2Δ strains were competent to switch mating types with efficiencies similar to the wild-type strain (Fig. S4C). To circumvent the galactose induction defects of a swi2Δ and to study the kinetics of strand invasion, an auxin-inducible degron system was used to conditionally deplete Swi2p (32). After a 2-h treatment with synthetic auxin [1-naphthaleneacetic acid (NAA)] to deplete Swi2p, galactose was added to cultures, and PCR was used to monitor DSB formation and strand invasion. Consistent with the results from the swi2Δ strain, de- pletion of Swi2p did not alter DSB repair kinetics (Fig. S4D). Because the Swi2p ATPase is essential for SWI/SNF function, these results indicate that SWI/SNF is dispensable for mating-type switching, even with a heterochromatic donor.

Publications and Data Availability
This study was supported by an American Cancer Society Grant to P.A.M. and S.P., and in part by grants from the Department of Energy (DE-FG02-05ER15785) and the National Institutes of Health (GM085671). This work was performed in part at the Stanford University Small Molecule Mass Spectrometry Facility, which is supported by the National Science Foundation (CHE-1265688).

Fig. S4. Swi2p–Sir3p contacts are required for eviction of Sir3p from nucleosomes. (A) SDS/PAGE analysis of SWI5SNF and SWI5SNF–Δ10R complexes, visualized by silver staining. Equal levels of ATPase activity were loaded for each enzyme. (B) DNA-stimulated ATPase kinetics of SWI5SNF and SWI5SNF–Δ10R are equivalent. ATPase reactions were performed with varying concentrations of DNA cofactor, and hydrolysis rates were fit to Michaelis–Menten kinetic parameters. (C) Mononucleosome mobilization by SWI5SNF and SWI5SNF–Δ10R enzymes is equivalent. Varying concentrations of enzymes were incubated with a mononucleosome positioned in the center of a radiolabeled, 282-bp DNA fragment harboring a 601 positioning sequence. Predicted positions of mononucleosomes are indicated to the left. (Upper) Gel. (Lower) Quantification (error bars reflect SD). (D) Schematic of the chromatin capture assay. Biotinylated nucleosomal arrays are bound to Sir3p, incubated with chromatin-remodeling enzyme and ATP and captured on streptavidin-coated magnetic beads. Chromatin-bound B and unbound U are assayed by Western blotting. (E) SWI5SNF–Δ10R is defective for Sir3p eviction from nucleosomes. Increasing amounts of chromatin-remodeling enzyme were incubated with Sir3p-bound nucleosomal array, and Sir3p eviction into the chromatin-unbound fraction U was measured by Western blotting. (Left) Representative blots. (Right) Quantification.

Fig. 3. SWI5SNF ATPases interact with BAH core domains. (A) Schematic shows C-terminal truncations within the Sir3p BAH domain. The indicated GST–BAH fusion proteins were incubated with either SWI5SNF or RSC, and bound fractions were assayed by Western. The Rsc2p BAH fusion contains only the core BAH domain; the BAH–CT-1 fusion also contains the C-terminal conserved CT-1 domain from Rsc2p. Western analyses used sera to the Arp9p subunit, common to both remodeling enzymes. (B) SWI5SNF, RSC, or swi2Δ complexes were incubated with GST–BAH fusions from yeast Orc1p or Sir3p. Bound fractions were assayed by Western to the indicated subunits. Lower shows Ponceau-stained membrane, depicting levels of GST fusions.

Fig. 4. Swi2p–Sir3p contacts are required for eviction of Sir3p from nucleosomes. (A) SDS/PAGE analysis of SWI5SNF and SWI5SNF–Δ10R complexes, visualized by silver staining. Equal levels of ATPase activity were loaded for each enzyme. (B) DNA-stimulated ATPase kinetics of SWI5SNF and SWI5SNF–Δ10R are equivalent. ATPase reactions were performed with varying concentrations of DNA cofactor, and hydrolysis rates were fit to Michaelis–Menten kinetic parameters. (C) Mononucleosome mobilization by SWI5SNF and SWI5SNF–Δ10R enzymes is equivalent. Varying concentrations of enzymes were incubated with a mononucleosome positioned in the center of a radiolabeled, 282-bp DNA fragment harboring a 601 positioning sequence. Predicted positions of mononucleosomes are indicated to the left. (Upper) Gel. (Lower) Quantification (error bars reflect SD). (D) Schematic of the chromatin capture assay. Biotinylated nucleosomal arrays are bound to Sir3p, incubated with chromatin-remodeling enzyme and ATP and captured on streptavidin-coated magnetic beads. Chromatin-bound B and unbound U are assayed by Western blotting. (E) SWI5SNF–Δ10R is defective for Sir3p eviction from nucleosomes. Increasing amounts of chromatin-remodeling enzyme were incubated with Sir3p-bound nucleosomal array, and Sir3p eviction into the chromatin-unbound fraction U was measured by Western blotting. (Left) Representative blots. (Right) Quantification.

Fig. 5.
recombinational repair of these same sequences when they are euchromatic, suggesting that this role for SWI/SNF might be specific for the heterochromatic context (19). To our surprise, however, our studies presented here do not support this key role for SWI/SNF in heterochromatic recombinational repair. We created swi2Δ strains that harbor a GAL–HO gene by tetrad dissection, and we found that these strains are competent to repair an HO-induced DSB, leading to mating-type switching with efficiencies similar to wild type. Furthermore, we used an inducible degron strategy to remove Swi2p from these GAL–HO strains, but in this case as well, the loss of Swi2p, and thus SWI/SNF, had no impact on repair of a DSB at the MAT locus. Why our results differ from those of Laurent and colleagues in not clear. Unfortunately, the original snf5Δ strain is no longer available. The most likely explanation is that the previously observed phenotype was specific to this particular snf5Δ isolate that was created by direct cell transformation, rather than tetrad dissection. Alternatively, it could represent a phenotype that is unique to a snf5Δ mutant and does not reflect a role for SWI/SNF per se.

Yeast strains that lack SWI/SNF show a variety of phenotypes, including growth defects on rich medium or medium containing alternative carbon sources (e.g., galactose or raffinose), inositol auxotrophy (25, 33) and sensitivity to DNA-damaging and replication stress agents (26, 30). Consistent with the intact chromatin-remodeling activities of the SWI/SNF–Δ10R enzyme, strains harboring the swi2–Δ10R allele showed normal growth on nearly every condition tested. The lone exception, however, was sensitivity to the replication stress agent HU. Furthermore, this phenotype was suppressed by deletion of the SIR3 gene, consistent with a role for ATP-dependent Sir3p eviction during replicative stress. This phenotype was not due to a defect in transcriptional induction of the RNR genes, and the swi2–Δ10R allele did not lead to significant transcriptional changes that could be detected by RNA-seq. Thus, this HU phenotype is likely to reflect a transcription-independent role of SWI/SNF action in antagonizing Sir3p during DNA replication. One simple model posits that SWI/SNF is required for efficient replication through Sir heterochromatin and that HU-induced fork stress heightens the need for SWI/SNF to remove Sir3p. Alternatively, Taddei and colleagues have shown that Sir proteins can be recruited to stalled replication forks (34). Perhaps SWI/SNF plays a role in removing Sir proteins from stalled forks, alleviating the negative consequences of this Sir recruitment. This model may also provide an explanation for the defect in heterochromatin establishment observed in the swi2–Δ10R strain, because an accumulation of Sir3p at stalled forks may titrate Sir proteins from heterochromatic domains, interfering with heterochromatin assembly.

The ATP-dependent eviction of Sir3p from chromatin is reminiscent of the ability of the yeast Mot1p ATPase to catalyze the eviction of the general transcription factor TATA-binding protein (TBP) from DNA. Mot1p is a member of the Swi2p/Snf2p family of DNA-stimulated ATPases and DNA topoisomerases, and the ability of Mot1p to disrupt TBP–DNA interactions appears to be key for redistributing TBP from TATA-containing binding sites to less-preferred, TATA-less promoter elements (35, 36). Similar to the SWI/SNF-dependent eviction of Sir3p from nucleosomes, Mot1p evicts TBP from a preformed TBP–DNA complex in an ATP-dependent reaction. Mot1p binds to TBP using two distinct interaction domains—a region containing multiple HEAT domains binds to the convex surface...
of the TBP–DNA complex, a distinct “latch” domain interacts with the surface of TBP that is bound to DNA (37). These structural studies have led to a model in which Mot1p binds to DNA adjacent to the TBP–DNA complex, allowing its HEAT domain to make extensive contacts with the exposed, convex surface of TBP. As Mot1p hydrolyzes ATP, DNA translocation leads to the removal of TBP from DNA, and the latch domain of Mot1p interacts with the DNA-binding surface of TBP, preventing reassociation with promoter DNA (37). By analogy, we propose that the HSA domain of Sir2p may interact with the Sir3p–nucleosome complex, facilitating Sir3p removal during the DNA translocation reaction. Likewise, sequences within the N-terminal lobe of the ATPase domain may function as latches that bind the Sir3 BAH domain, preventing reassociation with the nucleosome (Fig. 6).

Although the Sir2 ATPase domain is uniquely able to interact with the Sir3p BAH, the ATPase domains from both Sir2p and Sir1p can interact with the yeast Orc1p BAH domain. Likewise, both the SWI/SNF and RSC complexes can bind to the BAH domain of human Orc1. These latter interactions are surprising given that the primary sequence of the yeast and human Orc1 BAH domains have diverged considerably, although the overall structures are homologous (Fig. S3A). Orc1p is a highly conserved subunit of the ORC that is essential for cell viability and important for DNA replication (38–40). Orc1p and Sir3p are paralogs, and as such they display domain and primary sequence conservation, particularly in their N-terminal BAH domains (47% identical sequence). In *Kluyveromyces lactis*, Orc1p has been shown to function analogously to the role of Sir3 in heterochromatin formation, in addition to its traditional role in replication (41). We postulate that the binding interaction between SWI/SNF-family enzymes and Orc1-like BAH domains is ancestral and that specificity for Sir3p and Sir2p arose following the silencing subfunctionalization of Sir3p. Indeed, the sequences within the Sir2p ATPase domain that diverge from Sir1p and that appear to provide specificity for Sir3p are not well conserved in mammalian Sir2p/Sir3p homologs (Fig. S5B). The specificity for different BAH domains seems to be imparted by interactions within the BAH and by species differences that surround and regulate access to the core BAH fold. In line with this hypothesis, we found that the truncated, core BAH domains of Rsc2p and Sir3p were able to interact with both the SWI/SNF and RSC enzymes, but inclusion of C-terminal regions that wrap about the folds inhibited RSC and SWI/SNF binding. Given the plethora of BAH domains associated with chromatin (14), this theme of BAH accessibility and gating might help regulate ATP-dependent chromatin-remodeling enzyme activities in a context-dependent manner.

**Materials and Methods**

Detailed information on reagent preparation, biochemical assays, and yeast culture work is located in SI Materials and Methods. Oligonucleotides, plasmids, and yeast strains used in this study are listed in Tables S1, S2, and S3, respectively.

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