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Lipopolysaccharide induces and activates the Nalp3 inflammasome in the liver

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Lipopolysaccharide induces and activates the Nalp3 inflammasome in the liver

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Abstract

AIM: To examine the activation of the Nalp3 inflammasome and its downstream targets following lipopolysaccharide (LPS)-induced stimulation in the liver.

METHODS: Six-to-eight-week-old C57BL/6 chow fed mice were injected intraperitoneally with 0.5 μg/g bodyweight LPS and sacrificed 2, 4, 6, 18 or 24 h later. LPS-induced liver damage was confirmed by a biochemical assay to detect alanine aminotransferase (ALT) levels. To determine if LPS stimulation in the liver led to activation of the inflammasome, real-time quantitative polymerase chain reaction was used to evaluate the mRNA expression of components of the Nalp3 inflammasome. Enzyme-linked immunosorbent assays were used to determine the protein expression levels of several downstream targets of the Nalp3 inflammasome, including caspase-1 and two cytokine targets of caspase-1, interleukin (IL)-1β and IL-18.

RESULTS: We found that LPS injection resulted in liver damage as indicated by elevated ALT levels. This was associated with a significant increase in both mRNA and protein levels of the proinflammatory cytokine tumor necrosis factor (TNF)-α in the liver, as well as increased levels of TNFs in serum. We showed that LPS stimulation led to upregulation of mRNA levels in the liver for all the receptor components of the inflammasome, including Nalp3, Nalp1, pannexin-1 and the adaptor molecule apoptosis-associated speck-like, caspase recruitment domain-domain containing protein. We also found increased levels of mRNA and protein for caspase-1, a downstream target of the inflammasome. In addition, LPS challenge led to increased levels of both mRNA and protein in the liver for two cytokine targets of caspase-1, IL-1β and IL-18. Interestingly, substantial baseline expression of pre-IL-1β and pre-IL-18 was found in the liver. Inflammasome and caspase-1 activation was indicated by the significant increase in the active forms of IL-1β and IL-18 after LPS stimulation.

CONCLUSION: Our results show that the Nalp3 inflammasome is upregulated and activated in the liver in response to LPS stimulation.

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Key words: Endotoxin; Nod-like receptor; Interleukin-1β; Interleukin-18; Caspase-1

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INTRODUCTION

The endotoxin lipopolysaccharide (LPS), a component of Gram-negative bacteria, plays an important role in...
acute liver injury as well as chronic liver diseases including fatty liver associated with either alcohol consumption or metabolic syndrome and obesity\(^1,^2,^3\). LPS has also been implicated in insulin resistance as well as in steatohepatitis in non-alcoholic fatty liver disease\(^3,^4\). Increasing evidence suggests that gut-derived LPS through the gut-liver axis affects the extent of liver damage in many different types of inflammatory liver diseases\(^5,^6\). LPS is a prototypical ligand for the pattern recognition receptor (PRR), Toll-like receptor 4 (TLR4). TLR4 induces downstream signaling via the MyD88 adapter molecule and induces production of proinflammatory cytokines through activation of the regulatory factor nuclear factor (NF)-κB\(^7,^8\). In the liver, TLR4 is expressed in both parenchymal and immune cells, thereby providing potential for LPS-induced activation\(^7\).

Studies have shown that the Nalp3 inflammasome is activated by both pattern associated molecular patterns (PAMPs), including LPS and bacterial RNA, and danger-associated molecular patterns (DAMPs)\(^9,^10\). The Nalp3 inflammasome is a caspase-1 activating multiprotein complex, which has been implicated in numerous inflammatory processes and human diseases including gout, pseudogout, contact hypersensitivity, and most recently Alzheimer’s disease\(^11,^12\). Furthermore, mutations in the Nalp3 gene that lead to gain-of-function mutations, result in several hereditary syndromes including: Muckle-Wells syndrome, familial cold autoinflammatory syndrome, and chronic infantile neurological cutaneous and articular syndrome\(^13\).

In response to stimulation by either DAMPs or PAMPs, Nalp3 interacts with pro-caspase-1 through the adaptor molecule [apoptosis-associated speck-like, caspase recruitment domain (CARD)-domain containing protein (ASC)] to form the inflammasome, which leads to activation of caspase-1. Active caspase-1, previously known as interleukin (IL)-1β converting enzyme, a heterodimer of p20 and p10 subunits, is the central effector protein of the inflammasome complex and promotes the cleavage of pro-IL-1β, pro-IL-18 and pro-IL-33 to their biologically active, mature forms\(^14\). In addition, active caspase-1 has been shown to cleave other substances, such as caspase-7 and sterol regulatory element-binding proteins, therefore playing a pivotal role in cell death and survival\(^15\).

The mRNA levels of the components the Nalp3 inflammasome are expressed in the eye, heart and lung in response to LPS stimulation\(^16\). Although mRNA levels of IL-1β have also been shown to increase in the liver in response to LPS stimulation, the role of the Nalp3 inflammasome in the liver in response to LPS stimulation has not been determined\(^17\). In the liver, the Nalp3 inflammasome is upregulated upon acetaminophen-induced toxicity\(^18\) and the contribution of Nalp3 has been described in Propionibacterium acnes and LPS-induced acute liver injury\(^19\); however, a role for Nalp3 in other liver conditions has not been identified.

Although the role of the proinflammatory cytokine, tumor necrosis factor (TNF)α, has been extensively studied in both alcoholic and non-alcoholic fatty liver disease, the role of IL-1β and the inflammasome has yet to be explored. LPS plays an important role in liver disease; elevated LPS levels are detected in the portal and systemic blood of patients with alcoholic and non-alcoholic fatty liver disease\(^19-21,^22\). Furthermore, there is increased sensitivity to LPS in both alcoholic liver disease and fatty liver disease\(^22-24\). Here, we examined if LPS induces the Nalp3 inflammasome in the liver. In this study, we showed that LPS stimulation led to induction of the components of the Nalp3 inflammasome at both the mRNA and protein level in the liver and this is associated with increased IL-1β production.

MATERIALS AND METHODS

**Mice**

Six-to-eight-week-old wild-type C57BL/6 chow fed mice received intraperitoneal injections of 0.5 μg/g LPS in PBS (Sigma, St Louis, MO, United States) (three mice per group) for 2, 4, 6, 18 and 24 h. Serum was separated from whole blood and stored at -80 °C. Liver samples were snap frozen in liquid nitrogen or stored in RNA later (Qiagen Sciences, Germantown, MD, United States) for RNA extraction. All animals received proper care in agreement with animal protocols at the University of Massachusetts Medical School Institutional Animal Use and Care Committee.

**Biochemical alanine aminotransferase assay**

Serum alanine aminotransferase (ALT) was determined using a kinetic method (DTEK LLC, Bensalem, PA, United States).

**RNA analysis**

RNA was purified from livers using the RNeasy kit (Qiagen Sciences) and on-column DNA digestion. cDNA was transcribed with the Reverse Transcription System (Promega, Madison, WI, United States). Real-time quantitative polymerase chain reaction (qPCR) was performed using iCycler (Bio-Rad Laboratories, Hercules, CA, United States) as described previously\(^22\). Primer sequences are shown in Table 1.

**Cytokine enzyme-linked immunosorbent assay measurements**

Whole cell lysates were extracted from liver tissue as previously described\(^22\). These lysates were assayed for mature IL-1β (R and D Systems, Minneapolis, MN, United States), TNF-α, total IL-18, and total IL-1β (BD Biosciences, United States). Serum was also assayed for mature IL-1β (R and D Systems).

**Statistical analysis**

Statistical significance was determined using Student’s /
RESULTS

LPS induces liver injury and inflammatory cytokines

Gut-derived LPS plays a role in several inflammatory liver diseases\[9\]. To evaluate the effects of LPS on the liver, we injected wild-type mice with LPS for 2, 4, 6, 18 or 24 h. Serum was separated from whole blood and analyzed for alanine aminotransferase (ALT), which was significantly increased after 4 h of LPS stimulation. Mean ± SD are shown. *P < 0.01.

ALT (IU/mL) vs Hours after LPS

![Graph showing ALT levels over time](image)

Figure 1 Lipopolysaccharide stimulation results in liver damage. C57Bl/6 wild-type chow-fed mice (three per group) were injected intraperitoneally with lipopolysaccharide (LPS) for 2, 4, 6, 18 or 24 h. Serum was separated from whole blood and analyzed for alanine aminotransferase (ALT), which was significantly increased after 4 h of LPS stimulation. Mean ± SD are shown. *P < 0.01.

LPS upregulates mRNA expression of Nalp3 inflammasome components

For complete function, the inflammasome requires expression of its various components including Nalp3, ASC (the adapter molecule), and caspase-1. We investigated the mRNA levels of the different types of inflammasomes, including the Nalp1 and Nlr4 inflammasomes. Our examination revealed a significant increase in liver mRNA of Nalp3 after ≥ 4 h LPS stimulation, with a peak at 6 h (Figure 3A). Upregulation of the inflammasome was not limited to Nalp3, and there was a significant increase in Nalp1 (sevenfold) and Nlr4 (3.5-fold) mRNA at 24 h compared to the unstimulated controls (P < 0.01, P = 0.04, respectively, data not shown). We also analyzed the mRNA levels of other components of the inflammasome including the pyrin and CARD-domain-containing adaptor ASC; caspase-1, an inflammatory caspase; and pannexin-1, a hemi-channel that is recruited upon activation of the P2X7 receptor and is a necessary component for inflammasome activation. The mRNA levels of ASC gradually increased at each time point after LPS stimulation (Figure 3B), whereas caspase-1 levels peaked at 6 h and pannexin-1 mRNA levels peaked at 6 h (Figure 3C and D).
Activation of the inflammasome results in increased levels of total IL-18, total IL-1β, and mature IL-1β.

LPS stimulation has been shown to increase levels of pro-IL-1β transcripts\(^{26}\). However, production of the biologically active IL-1β and IL-18 requires the mature caspase-1 to cleave the precursor forms into their mature forms\(^{27}\). First, we evaluated mRNA levels of IL-1β and IL-18 and found that IL-1β mRNA levels were significantly increased at all time points post-LPS stimulation peaking at 2 h (Figure 4A). Induction of IL-18 mRNA by LPS was modest but reached statistical significance at the 6-h time point (Figure 4B). There was also induction of the mRNA for the IL-1 receptor antagonist after LPS stimulation in the liver (Figure 4C).

We sought to determine whether the increase in mRNA correlated with increased protein production for IL-1β and IL-18. We found basal expression of pro-IL-1β protein in the livers of unstimulated control mice and this was significantly upregulated at 2 h after LPS stimulation (Figure 5A). More importantly, we observed a significant increase in mature IL-1β protein in liver tissue at all time points post-LPS stimulation compared to the controls, in which no mature IL-1β was detected (Figure 5B). Consistent with inflammasome activation, we found increased serum IL-1β levels in mice after stimulation with LPS for 2 and 6 h (Figure 5C). Investigation of IL-18, which is also cleaved by caspase-1, revealed a significant increase in total IL-18 in the liver at all time points following LPS injection (Figure 5D).

**DISCUSSION**

Although many molecules and compounds activate the
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Figure 5 Lipopolysaccharide increased interleukin-1β and interleukin-18 protein. Protein level was detected in liver tissue by enzyme-linked immuno-sorbent assay for total-interleukin (IL)-1β (pro-IL-1β) and cleaved IL-1β (A), cleaved IL-1β in the liver tissue (B), cleaved IL-1β in the serum (C) and total IL-18 (pro-IL-18 and cleaved IL-18) (D) in liver tissue. The values shown are the fold change compared to non-Lipopolysaccharide (LPS) stimulated control. Protein levels were normalized to total protein concentrations in each tissue sample. Mean ± SD are shown. n = 3 per group (except at 2 h LPS stimulation, where n = 2 due to an outlier), *P < 0.01.

Nalp3 inflammasome both in vitro and in vivo[3], the role of Nalp3 inflammasome in a healthy liver following LPS stimulation has not been determined. Our novel findings demonstrate upregulation of the Nalp3 inflammasome in the liver in response to LPS stimulation, both at the mRNA and protein levels. We also show for the first time that the components of the inflammasome pathway, including ASC and caspase-1, are upregulated in the liver after LPS challenge and this results in functional activation of the inflammasome and caspase-1, indicated by IL-1β and IL-18 secretion.

LPS is recognized by TLR4, which induces an intracellular signaling cascade leading to activation of NF-κB, as well as the production of pro-IL-1β and pro-IL-18[28]. Previous studies have suggested that signaling through the TLRs alone is insufficient to generate the production of mature IL-1β and IL-18, and that a second signal is needed that leads to ionic perturbation[29]. These perturbations may include ATP signaling through the P2X7 channel, leading to changes in intracellular potassium concentrations and membrane perturbations. The physiological importance of ATP in IL-1β secretion is unclear and the second signal that leads to cleavage of pro-IL-1β in the liver is still under investigation[30]. In vitro studies have shown that a second signal is needed in addition to LPS to obtain cleavage of pro-IL-1β[31]. It is possible that a DAMP released by injured cells could act as the second signal. In vivo studies, however, have shown that LPS alone is sufficient to increase cleavage of IL-1β and IL-18[31]. Our novel data demonstrate that, after in vivo LPS challenge, the inflammasome is activated in the liver and this results in increased inflammasome function and IL-1β secretion. We found that LPS stimulation not only increased IL-1β mRNA levels and the expression of pro-IL-1β protein, but it significantly increased the levels of the 18-kDa form of mature IL-1β in the liver. Secretion of mature IL-1β also resulted in elevated IL-1β in the serum. Interestingly, we found significant levels of pro-IL-1β protein expressed in the liver without exogenous stimulation. This suggests that the exogenous LPS probably provided the second signal for IL-1β production. The mechanisms for the presence of pro-IL-1β protein in the liver remain to be determined, however, one possibility is stimulation via PAMPs, such as LPS, which are being constantly supplied by the gut through the portal vein to the liver[31].

Cleavage of pro-IL-1β is mediated by activated caspase-1; consistent with this, we found increased levels of IL-1β protein in the serum after LPS challenge. The other target of the caspase-1 complex and inflammasome activation is IL-18. Consistent with LPS-induced inflammasome and caspase-1 activation, we found an increase in IL-18 mRNA and total IL-18 levels in the liver. Due to limitations of the detection assay, we could not distinguish between the mature form of IL-18 and pro-IL-18, but the increase in total IL-18 after LPS stimulation was most likely due to an increase in both.

We found elevated basal levels of pro-IL-1β in the liver, and pro-IL-1β was further increased by LPS with the highest levels at 2 h following stimulation. It is likely that LPS-induced inflammasome activation can cleave the pre-existing pro-IL-1β to the mature form by caspase-1. In addition to activation of the Nalp3 inflammasome, our data suggests that other types of caspase-1-activating inflammasomes, Nalp1 and Nlrc4, may also play a role in caspase-1 activation and subsequent IL-1β cleavage in the liver. Our data showed that both Nalp1 and Nlrc4 mRNA...
levels were upregulated in the liver following LPS stimulation. This suggests that other inflammasome-activating signals lead to full activation in the liver.

The contribution of the Nalp3 inflammasome to inflammation in the liver in response to LPS stimulation may have implications for several different liver diseases. Increased gut-derived LPS has been shown to contribute to liver inflammation in alcoholic and non-alcoholic fatty liver diseases, as well as other forms of liver damage such as hepatitis C virus infection. In healthy subjects, the liver plays a central role in elimination of gut-derived endotoxins and other pathogens to maintain immune homeostasis. Thus, it is tempting to speculate that gut-derived ligands of TLRs constantly entering the liver could contribute to the high basal expression of pro-IL-1β found in the liver in our experiments. The high basal expression of pro-IL-1β represents the first step of inflammasome activation, and this appears to provide a pre-activated state where even single LPS stimulation can result in inflammasome activation, as seen in the liver after in vivo LPS challenge in our experiments. The specific role of Nalp3 and the inflammasome remains to be evaluated in different types of liver diseases where inflammation is related to LPS and other pathogen-derived or endogenous danger signals.

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REFERENCES

18. Imamura M, Tsutsui H, Yasuda K, Uchiyama R, Yumikura-


Yang SQ, Lin HZ, Lane MD, Clemens M, Diehl AM. Obesity increases sensitivity to endotoxin liver injury: implications for the pathogenesis of steatohepatitis. Proc Natl Acad Sci USA 1997; 94: 2557-2562


Pålsson-McDermott EM, O’Neill LA. Signal transduction by the lipopolysaccharide receptor, Toll-like receptor-4. Immunology 2004; 113: 153-162


Hogquist KA, Nett MA, Unanue ER, Chaplin DD. Interleukin 1 is processed and released during apoptosis. Proc Natl Acad Sci USA 1991; 88: 8485-8489


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