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# $\beta$ 1,3-*N*-Acetylglucosaminyltransferase 1 Glycosylation Is Required for Axon Pathfinding by Olfactory Sensory Neurons

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During embryonic development, axons from sensory neurons in the olfactory epithelium (OE) extend into the olfactory bulb (OB) where they synapse with projection neurons and form glomerular structures. To determine whether glycans play a role in these processes, we analyzed mice deficient for the glycosyltransferase  $\beta$ 1,3-*N*-acetylglucosaminyltransferase 1 ( $\beta$ 3GnT1), a key enzyme in lactosamine glycan synthesis. Terminal lactosamine expression, as shown by immunoreactivity with the monoclonal antibody 1B2, is dramatically reduced in the neonatal null OE. Postnatal  $\beta$ 3GnT1<sup>-/-</sup> mice exhibit severely disorganized OB innervation and defective glomerular formation. Beginning in embryonic development, specific subsets of odorant receptor-expressing neurons are progressively lost from the OE of null mice, which exhibit a postnatal smell perception deficit. Axon guidance errors and increased neuronal cell death result in an absence of P2, I7, and M72 glomeruli, indicating a reduction in the repertoire of odorant receptor-specific glomeruli. By ~2 weeks of age, lactosamine is unexpectedly reexpressed in sensory neurons of null mice through a secondary pathway, which is accompanied by the regrowth of axons into the OB glomerular layer and the return of smell perception. Thus, both neonatal OE degeneration and the postnatal regeneration are lactosamine dependent. Lactosamine expression in  $\beta$ 3GnT1<sup>-/-</sup> mice is also reduced in pheromone-receptive vomeronasal neurons and dorsal root ganglion cells, suggesting that  $\beta$ 3GnT1 may perform a conserved function in multiple sensory systems. These results reveal an essential role for lactosamine in sensory axon pathfinding and in the formation of OB synaptic connections.

**Key words:** olfactory development; sensory neurons; odorant receptors; axon guidance; glycosyltransferase; lactosamine

## Introduction

Odor stimuli are transmitted by olfactory sensory neurons that are widely dispersed in the nasal cavity to focal sites of synaptic contact in the olfactory bulb (OB), termed glomeruli. Populations of neurons expressing a given odorant receptor project axons that converge on one or a few glomeruli in each OB where they synapse with the dendritic arbors of mitral and tufted cells, forming the first central relay in olfactory perception (Ressler et al., 1993; Vassar et al., 1994; Mombaerts et al., 1996). The convergence of axons expressing the same odorant receptor to specific glomerular loci in the OB creates a representative map of receptor activation that is thought to be essential for encoding

odorant information for processing by higher centers in the brain.

A role for odorant receptors in glomerular targeting has emerged from genetic ablation and receptor swap studies in mice (Mombaerts et al., 1996; Wang et al., 1998), although a mechanism through which these receptors might affect the behavior of growth cones remains elusive and does not seem to be evolutionarily conserved (Dobritsa et al., 2003). However, known guidance factors are differentially expressed by olfactory neurons where they influence the patterning of olfactory projections. Semaphorin 3A (Sema3A), for example, is expressed by ensheathing glial cells and restricts the innervation of neuropilin-1<sup>+</sup> axons to medial and lateral glomerular domains (Pasterkamp et al., 1998; Schwarting et al., 2000, 2004). In addition, genetic manipulation of ephrin expression levels alters the positioning of glomeruli (Cutforth et al., 2003). Thus, it is likely that olfactory axons are at least partially guided by cues on cells and axons in adjacent pathways. In addition, other molecules, such as *N*-cadherin, regulate axodendritic interactions required for protoglomerular formation but do not appear to directly influence pathfinding to glomeruli (Hummel and Zipursky, 2004).

Cell-surface carbohydrates are an additional source of neuronal heterogeneity (Crandall et al., 2000; Lipscomb et al., 2002;

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Gibson et al., 2004; Storan and Key, 2004). The stereotyped distribution of glycan-expressing glomeruli has led to the hypothesis that glycans may directly influence targeting of sensory axons (Lipscomb et al., 2003). One such glycan is terminal lactosamine, which is expressed by neurons that preferentially innervate ventromedial targets in embryonic rodents, consistent with a role in patterning olfactory connectivity (Schwartz and Crandall, 1991). Interestingly, Dodd and Jessell (1985) showed that lactosamine was also expressed by subsets of dorsal root ganglion (DRG) neurons, where it was suggested to influence the organization of projections to specific dorsal horn laminae.  $\beta$ 1,3-N-acetylglucosaminyltransferase 1 ( $\beta$ 3GnT1) is a member of a family of  $\beta$ 3GnTs that regulate the initiation and extension of lactosamine chains on glycoproteins and glycolipids. We show here that  $\beta$ 3GnT1 determines lactosamine expression by sensory neurons in both the olfactory epithelium (OE) and the vomeronasal organ, in addition to nociceptive neurons in DRGs. Furthermore, we demonstrate that  $\beta$ 3GnT1 is required for the formation of sensory axon connections and normal glomerular formation during olfactory development.

## Materials and Methods

**Animals.**  $\beta$ 3GnT1 null mice were established from the KST308 embryonic stem cell line, which harbors a secretory trap vector insertion in the  $\beta$ 3GnT1 locus (available from the Baygenomics Consortium) (Mitchell et al., 2001). The pGT1TMPfs plasmid used for producing KST308 cells encodes a splice acceptor site that directs the expression of a fusion transcript consisting of the  $\beta$ 3GnT1 exon 1 sequence upstream of the integration site spliced to a vector-derived rat CD4 transmembrane domain sequence fused in frame with the  $\beta$ -geo reporter gene. 5'-Rapid amplification of cDNA ends analysis determined that the vector integrated in the second  $\beta$ 3GnT1 exon at position 686 of the coding region (GenBank accession number NM\_016888), predicting the expression of a severely truncated  $\beta$ 3GnT1 fusion product from the disrupted locus.  $\beta$ 3GnT1 mice were generated on a mixed 129Ola  $\times$  C57BL/6 background and backcrossed to C57BL/6 for four generations before our analysis of heterozygous intercrosses. I7-Internal ribosomal entry site (IRES)-tau-green fluorescent protein (GFP), M72-IRES-tau-GFP, and P2-IRES-tau-lacZ mice were generated by Dr. Peter Mombaerts (Rockefeller University, New York, NY). P2-IRES-tau-lacZ mice have been previously backcrossed over several generations to the background of the  $\beta$ 3GnT1 line. Mice were housed according to standard National Institutes of Health and institutional care guidelines.

**In situ hybridization.** Antisense and sense riboprobes corresponding to the 1100 bp of  $\beta$ 3GnT1 coding sequence immediately downstream of the transmembrane domain were subcloned into pBluescript-II SK<sup>+</sup>. Digoxigenin-labeled riboprobes (Roche Applied Science, Indianapolis, IN) were synthesized and hybridized to 4% paraformaldehyde-fixed, cryosectioned tissue as described previously (Henion et al., 2001). Odorant receptor riboprobes, prepared according to Iwema et al. (2004), were provided by Dr. James Schwob (Tufts University, Boston, MA).

**Histology and immunocytochemistry.** Histochemical and immunocytochemical analysis was conducted on free-floating sections prepared by 4% paraformaldehyde fixation and sectioning on a sliding microtome at 50  $\mu$ m thickness. Placental alkaline phosphatase labeling of axons in genetrapp mouse lines was performed using nitroblue-tetrazolium-chloride/5-bromo-4-chloro-3-indolyl-phosphate substrate, as described previously (Leighton et al., 2001). The mouse monoclonal antibody 1B2 (IgM) (Young et al., 1981) against the lactosamine epitope was obtained from American Type Culture Collection (Manassas, VA). Monoclonal antibody to olfactory marker protein (OMP) was a generous gift from Dr. Frank Margolis (University of Maryland, Baltimore, MD). Unless otherwise noted, 1B2 and OMP immunoreactivity was localized with biotinylated secondary antibodies in combination with avidin–biotin–horseradish peroxidase (Vectastain standard ABC kit; Vector Laboratories, Burlingame, CA), visualized with a diaminobenzidine–nickel reaction, as described previously (Schwartz et al., 2000). P2 glomeruli in

$\beta$ 3GnT1/P2-IRES-tau-LacZ mice were visualized with 1:2000 rabbit anti- $\beta$ -galactosidase (Cortex Biochem) and cyanine 3-conjugated secondary antibody against rabbit IgG (Jackson ImmunoResearch, West Grove, PA).

**Cell death and proliferation analysis.** Assays for apoptosis in the OE were performed using protocols specified in the DeadEnd terminal deoxynucleotidyl transferase-mediated biotinylated UTP nick end labeling (TUNEL) system (Promega, Madison, WI) with the modification of using Vectastain ABC kit amplification with diaminobenzidine–nickel substrate detection. Relative levels of cell death between  $\beta$ 3GnT1<sup>+/-</sup> and  $\beta$ 3GnT1<sup>-/-</sup> mice were determined by counting the number of TUNEL<sup>+</sup> cells per square millimeter of coronal dorsal OE sections using three animals per group. To assay OE basal cell proliferation, mice were administered a single 12.5 mg/kg dose of 5-bromo-2'-deoxyuridine (BrdU) 30 min before killing. Tissues were fixed in 4% paraformaldehyde and reacted with biotinylated rat anti-BrdU antibody (Axyll) and biotinylated anti-rat secondary antibody before visualization with Vectastain ABC and diaminobenzidine–nickel substrate.

**Buried food olfactory test.** Mice were tested for their ability to find buried food according to Wrenn et al. (2003) with minor modifications. Briefly, mice were weaned at day 14 and tested beginning on day 16. Testing was conducted for 3 consecutive days each week over the course of 4 weeks. The test was also conducted on 16-week-old adult mice. Three mutant, three heterozygote, and three wild-type littermates were habituated to the food reward by placing pieces of cookies in the cage overnight and then depriving the mice of food for 6 h. The test was conducted in 44  $\times$  25 cm rat cages, and a piece of cookie was hidden under 2.5 cm of clean standard bedding. One mouse was placed in the cage and timed for the latency to find and begin eating the cookie. A maximum of 10 min was allowed.

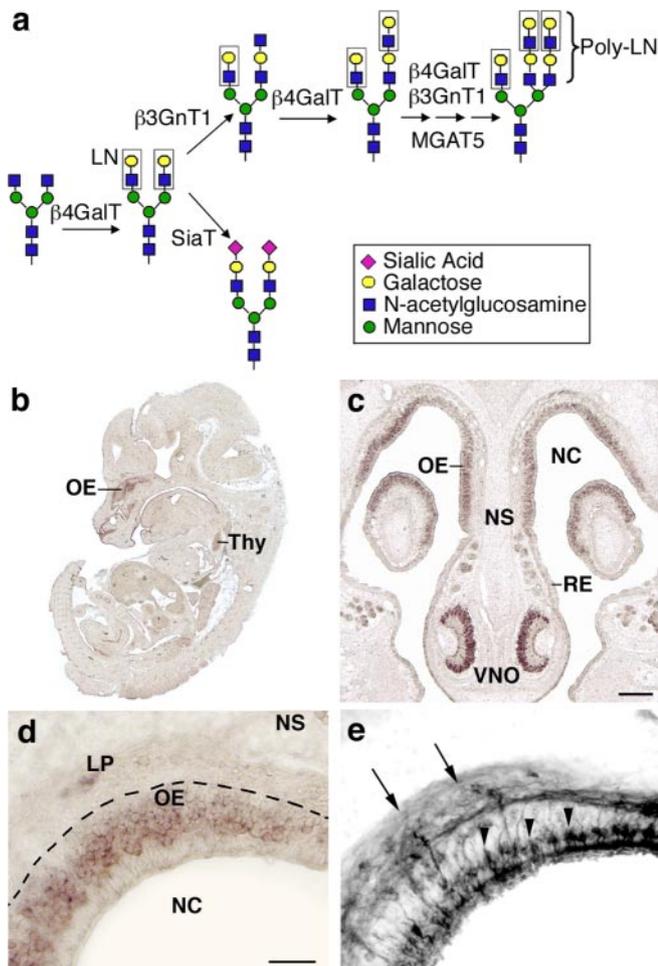
## Results

### $\beta$ 3GnT1 mRNA expression during nervous system development

Lactosamine (Gal $\beta$ 1,4-GlcNAc) is a common core component of complex N-linked glycans and some O-glycans and glycolipids (Fig. 1a). Olfactory sensory neurons are unique in the expression of a form of terminal lactosamine that is recognized by the monoclonal antibody 1B2.  $\beta$ 3GnT enzyme activities play a key role in regulating lactosamine initiation and extension in different tissues. To investigate how lactosamine synthesis is determined during olfactory development, we used *in situ* hybridization to map the expression of several  $\beta$ 3GnTs with established roles in lactosamine extension. Of five potential candidates examined,  $\beta$ 3GnT1,  $\beta$ 3GnT3,  $\beta$ 3GnT5,  $\beta$ 3GnT6, and iGnT (Sasaki et al., 1997; Zhou et al., 1999; Henion et al., 2001; Yeh et al., 2001; Iwai et al., 2002), only  $\beta$ 3GnT1 was broadly transcribed throughout all OE regions, similar to the established pattern of 1B2 reactivity (Crandall et al., 2000). At embryonic day 13 (E13),  $\beta$ 3GnT1 was mostly restricted to olfactory sensory neurons, endothelial cells, and the thymus, with no detectable expression by any neuronal population in the brain (Fig. 1b).  $\beta$ 3GnT1 was also transcribed in other developing regions of the peripheral nervous system, including dorsal root, trigeminal, and IXth cranial ganglia (data not shown). Expression of  $\beta$ 3GnT1 was strikingly high in vomeronasal neurons of the accessory olfactory system, although most neurons in the OE transcribed  $\beta$ 3GnT1 at relatively uniform levels (Fig. 1c,d).  $\beta$ 3GnT1 expression continued throughout postnatal development in sensory neurons adjacent to the mature population of 1B2<sup>+</sup> neurons located near the apical OE surface (Fig. 1d,e).

### Lactosamine-deficient $\beta$ 3GnT1<sup>-/-</sup> mice exhibit neuronal loss

Direct regulation of lactosamine expression by  $\beta$ 3GnT1 was established using null mice generated from embryonic stem cells identified in a secretory trap insertional mutagenesis screen (Mitchell et al., 2001). Integration of the secretory trap vector



**Figure 1.**  $\beta 3\text{GnT1}$  and lactosamine are strongly expressed in the developing olfactory system. **a**, The lactosamine disaccharide recognized by 1B2 antibodies (boxes) occurs widely on *N*-glycans but is typically capped by sialic acid or other glycans. Lactosamine can be extended by tissue-specific  $\beta 3\text{GnT1}$  and  $\beta 4\text{GalT}$  activities to form poly-lactosamines, which frequently occur on MGAT5 branched glycans. The  $\beta 3\text{GnT1}$  dependence of 1B2 reactivity may indicate preferential recognition of poly-lactosamine in tissues. **b–d**, *In situ* hybridization analysis of  $\beta 3\text{GnT1}$  expression in the mouse in a sagittal section through an E13 embryo (**b**), E15 coronal nose (**c**), and P1 OE (**d**). The dashed line in **b** denotes the border between the OE and the underlying lamina propria (LP). **e**, ICC showing 1B2<sup>+</sup> sensory neurons (arrowheads) and axons (arrows) in the section adjacent to **d**. NC, Nasal cavity; NS, nasal septum; RE, respiratory epithelium; Thy, thymus; VNO, vomeronasal organ. Scale bars: **c**, 200  $\mu\text{m}$ ; **d**, **e**, 50  $\mu\text{m}$ .

encoding  $\beta$ -geo and human placental alkaline phosphatase (PLAP) truncates  $\beta 3\text{GnT1}$  after 228 aa, removing the “DxD” and other conserved C-terminal motifs required for  $\beta 1,3$ -glycosyltransferase activity (Malissard et al., 2002). Mutant mice had no obvious phenotypic abnormalities and survived postnatally similar to control littermates. Immunocytochemical (ICC) analysis of postnatal day 1 (P1)  $\beta 3\text{GnT1}^{-/-}$  OEs revealed that 1B2 reactivity on cell bodies was widely abolished, with only residual lactosamine expression by scattered neurons (Fig. 2*a*). Mature OMP<sup>+</sup> neurons, which normally occupy several layers of apical cells near the OE surface, were reduced to a single cell layer in  $\beta 3\text{GnT1}^{-/-}$  mice (Fig. 2*a*). This loss of mature neurons correlated with a >20% decrease in relative OE thickness between  $\beta 3\text{GnT1}^{-/-}$  and  $\beta 3\text{GnT1}^{+/+}$  mice from P1 ( $0.77 \pm 0.05$ ;  $52.8 \mu\text{m}$   $\beta 3\text{GnT1}^{-/-}$  vs  $68.3 \mu\text{m}$   $\beta 3\text{GnT1}^{+/+}$ ) through P25 ( $0.78 \pm 0.01$ ;  $46.3 \mu\text{m}$   $\beta 3\text{GnT1}^{-/-}$  vs  $59.0 \mu\text{m}$   $\beta 3\text{GnT1}^{+/+}$ ).

The PLAP enzyme in  $\beta 3\text{GnT1}$  mice is expressed from the disrupted genomic locus as a bicistronic RNA with  $\beta$ -geo under

translational control of an IRES. PLAP histochemical activity selectively labels axon processes of neurons transcribing secretory-trapped genes (Leighton et al., 2001) and was used to map the projections of olfactory neurons expressing  $\beta 3\text{GnT1}$ . At P1, protoglomeruli were visible in  $\beta 3\text{GnT1}^{+/+}$  mice but absent from mutant OEs (data not shown). By P10, glomeruli were morphologically prominent in control animals, but the glomerular layer of  $\beta 3\text{GnT1}^{-/-}$  mice was widely devoid of innervation by both PLAP-labeled and 1B2<sup>+</sup> axons (Fig. 2*b*). ICC analysis with OMP revealed that the OE nerve layer was reduced in thickness compared with control mice, indicating a decrease in the number of axons innervating the OE (Fig. 2*b*). The OMP<sup>+</sup> axons that remained in the mutant nerve layer mostly failed to enter the glomerular neuropil or segregate into discrete glomeruli, suggesting that  $\beta 3\text{GnT1}$  loss caused defects in axon extension through the nerve layer into the developing glomerular layer. Residual glomeruli that did form in  $\beta 3\text{GnT1}^{-/-}$  mice were excluded from the dorsal OE, a region that appears particularly sensitive to innervation defects (Wang et al., 2004; Yu et al., 2004). The failure of glomerulogenesis correlated with an overall 22% reduction in OE size in null adults (data not shown).

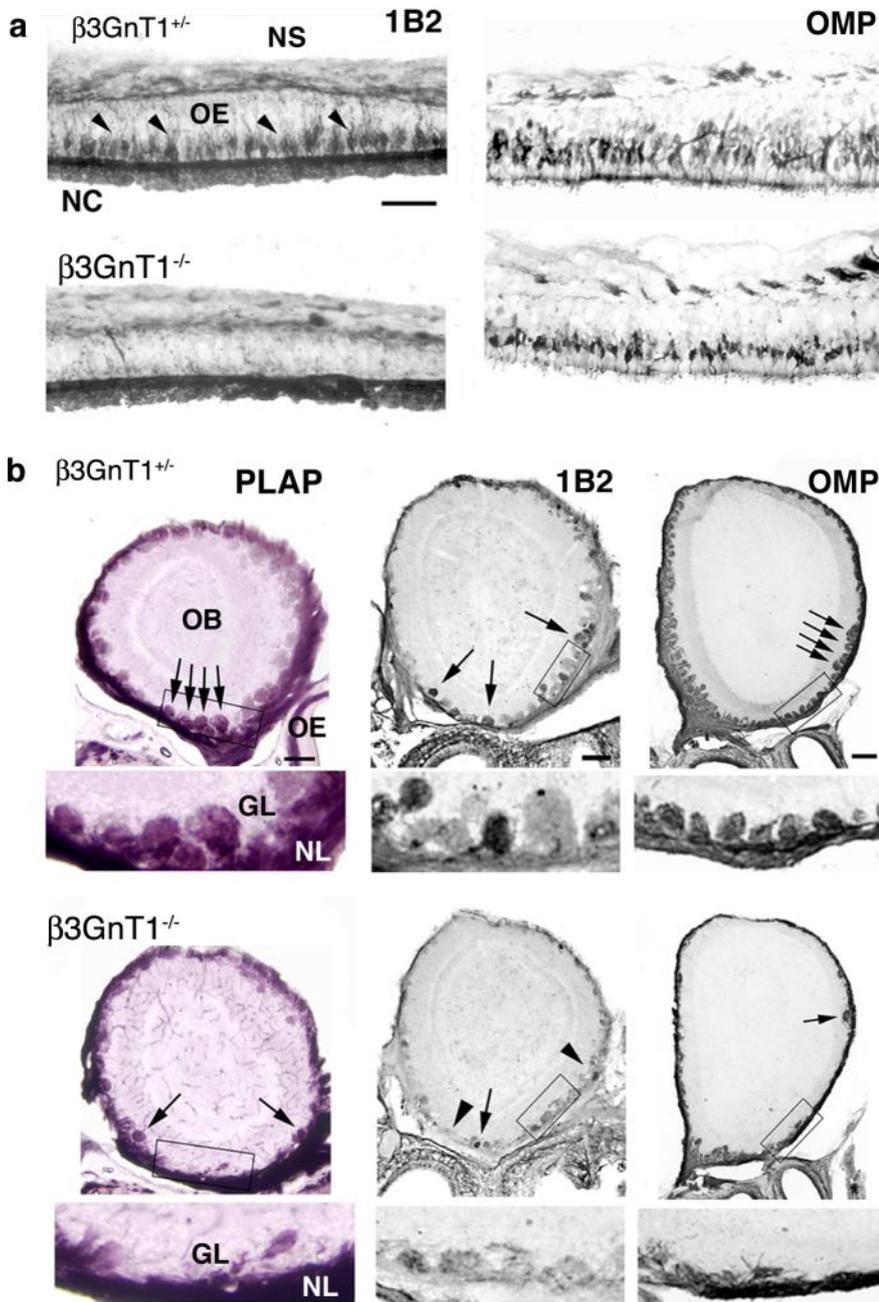
#### Increased cell death and precursor proliferation in $\beta 3\text{GnT1}^{-/-}$ mice

We examined whether the decreases observed in OE innervation and number of 1B2/OMP<sup>+</sup> neurons in the OE of early postnatal mutant mice could be related to changes in neuronal cell death. Developmental apoptosis in the mouse OE normally peaks at approximately E16 and declines toward birth (Voyron et al., 1999). In P1 null mice, a significant elevation in the number of cells undergoing apoptosis was detected by TUNEL analysis ( $161 \pm 45/\text{mm}^2$   $\beta 3\text{GnT1}^{-/-}$  vs  $60 \pm 2/\text{mm}^2$   $\beta 3\text{GnT1}^{+/+}$ ;  $p > 0.01$ ) (Fig. 3*a*). This paralleled the failure in glomerular formation and suggested that olfactory neuron cell death may result from failed connectivity with the OE. By P10, the number of apoptotic cells in all mice had decreased significantly and was not substantially increased in null mice relative to controls. BrdU-labeling analysis, however, revealed a striking increase in the rate of basal progenitor cell proliferation in P10 null OEs (Fig. 3*b*). Cell division was not obviously altered at P1, P25, or adult ages. The transient increase in neurogenesis observed >1 week after the peak of cell death is consistent with delayed neurogenic responses elicited after bulbectomy or methyl bromide-induced OE lesions (Schwartz-Levy et al., 1991; Schwob et al., 1995).

#### Specific odorant receptor subsets are lost from the $\beta 3\text{GnT1}^{-/-}$ OE

Survival of specific olfactory neuronal populations was examined by analyzing odorant receptor gene expression. During late embryogenesis and the first two postnatal weeks, the number of neurons expressing the P2 odorant receptor normally undergoes a rapid expansion, paralleling the increased size and complexity of the nasal cavity (Royal and Key, 1999). We used *in situ* hybridization to examine odorant gene expression throughout development. P2 neurons were progressively lost from the OE in  $\beta 3\text{GnT1}^{-/-}$  mice, beginning in embryonic development (Fig. 4). At E17, the number of P2 neurons in the  $\beta 3\text{GnT1}^{-/-}$  OE was 72% that of control littermates. The relative number of neurons declined to 58% by P1 and further decreased to 45% at P10, whereas in adult mutants, <15% of P2 neurons survived.

To further investigate the fate of odorant receptor-specific neurons, we crossed  $\beta 3\text{GnT1}$  mice to I7-IRES-tau-GFP odorant receptor reporter strain and compared GFP<sup>+</sup> neurons in mutant



**Figure 2.** Mature olfactory sensory neurons and OB glomeruli are lost in lactosamine  $\beta 3GnT1$  null mice. **a**, 1B2 and OMP ICC analysis of P1 OE. 1B2 expression in sensory neurons (arrowheads) is virtually absent in  $\beta 3GnT1$  null mice. Residual 1B2 labeling of the luminal surface of the OE is mucin dependent and is not present in Core-2 O-glycan-deficient mice (data not shown). The number of OMP<sup>+</sup> neurons is greatly reduced in null mice, indicating that lactosamine is required for maintenance of the majority of fully differentiated sensory neurons. NC, Nasal cavity; NS, nasal septum. Scale bar, 50  $\mu m$ . **b**, Histochemical staining for PLAP activity and ICC analysis of 1B2 and OMP expression in coronal sections from P10 OBs. In null mice, the number of glomeruli (arrows) is reduced, and most 1B2 reactivity is lost (arrowheads). Higher-magnification views of boxed areas, shown below each figure, demonstrate the severe loss of normal glomerular structures at this age. GL, Glomerular layer; NL, nerve layer. Scale bar, 200  $\mu m$ .

mice and control littermates.  $\beta 3GnT1$  null mice displayed a loss of I7-GFP neurons over time similar to that observed for P2 neurons. A noticeable deletion of I7 neurons began by E17 and progressed with postnatal development until <20% of I7 neurons remained in the adult null OE (data not shown).

#### Odorant receptor-specific axon trajectories

P2 axon projections were visualized by crossing  $\beta 3GnT1$  mice to the P2-IRES-tau-LacZ reporter strain (Mombaerts et al., 1996)

and staining for  $\beta$ -galactosidase protein expression. I7 and M72 axon trajectories were visualized by crossing  $\beta 3GnT1$  and I7- and M72-IRES-tau-GFP mice, respectively. ICC analysis of OEs from P2 mice at different developmental ages confirmed the loss of  $\beta$ -galactosidase<sup>+</sup> P2 neurons observed by *in situ* hybridization. Serial sections from E17  $\beta 3GnT1^{+/+}$  OBs revealed labeled fibers that converged on the ventral nerve layer and formed a typical P2 protoglomerulus (Fig. 5a). In contrast, axons from  $\beta 3GnT1^{-/-}$  mice failed to penetrate the developing glomerular layer, but instead remained in the nerve layer adjacent to their normal protoglomerular position. At later time points, when P2 axons had condensed to form a mature glomerulus in  $\beta 3GnT1^{+/+}$  mice, P2 axons were not detected in the mutant nerve layer, indicating that axons reaching the OB at P1 were not maintained (Fig. 5a).

A glomerular formation defect analogous to that for P2 also occurred for I7 axons of mutant mice. By P7, very few GFP<sup>+</sup> I7 fibers reached appropriate positions in the ventral nerve layer, and only rarely did I7 axons grow into the glomerular layer (Fig. 5b). Axons from the residual neurons expressing I7 in adult mutant OEs were not found in the nerve layer, although this could reflect a decrease in the number of remaining I7 axons below a critical threshold required to sustain glomerular formation (Ebrahimi and Chess, 2000).

We crossed  $\beta 3GnT1$  mice with M72-IRES-tau-GFP mice to quantify the number of M72-GFP neurons in null mice and to analyze axon trajectories of neurons originating in the dorsal OE and projecting to the dorsal, caudal OB. Compared with wild-type controls at P1, the OE of mutant mice displayed an ~60% reduction in M72 neurons (Fig. 6a), which increased over early postnatal development similar to the depletion of P2 and I7 neuron populations. Despite the ongoing loss of M72-GFP neurons in the OE, bundles of GFP<sup>+</sup> axons were still detectable in the

nerve layer of null mice, growing toward their targets in the dorsal, caudal OB. Unlike the P2 and I7 crosses to  $\beta 3GnT1$  null mice, in which few labeled axons were seen

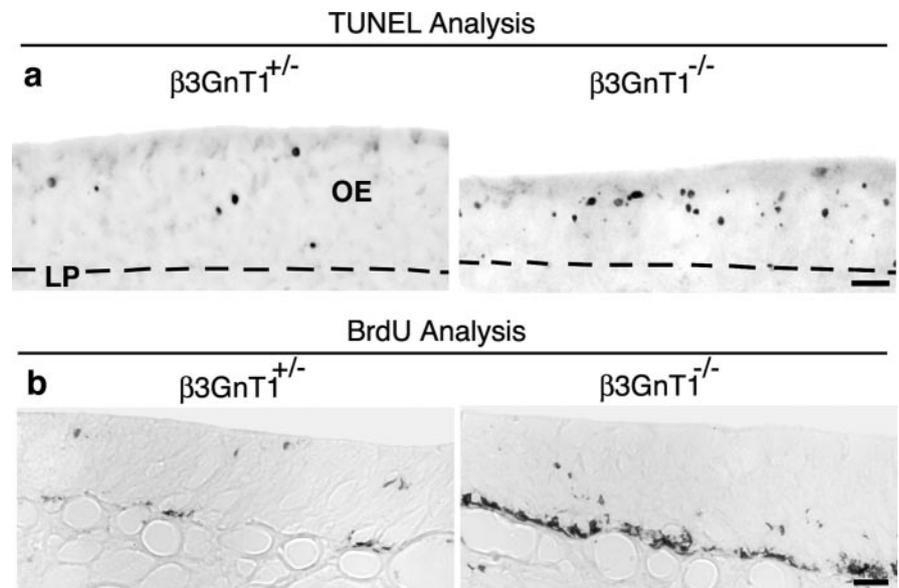
in the nerve layer at P10, axon guidance appeared to be perturbed in  $\beta 3GnT1^{-/-}$ /M72-IRES-tau-GFP mice. In control mice, the medial M72 glomerulus was typically positioned in the dorsomedial quadrant of the extreme caudal end of the OB (Fig. 6b). In  $\beta 3GnT1$  mutant mice, M72 axons failed to form a glomerulus in the normal M72 position (Fig. 6b, right panel, arrows), and in some examples, axons continued to grow to the dorsal extent of the OB. Thus, although there were a decreased number of M72

neurons in  $\beta 3\text{GnT1}^{-/-}$  OE, M72 axons grew into the dorsal nerve layer but appeared to bypass their intended target and failed to form normal glomeruli.

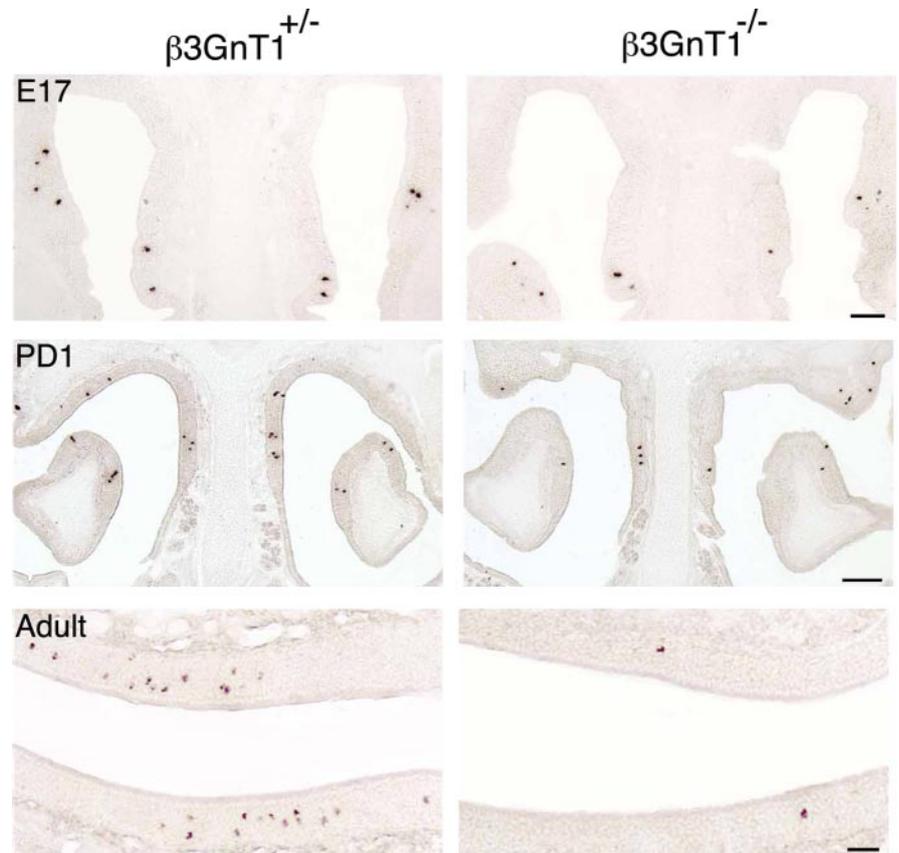
#### OB reinnervation in $\beta 3\text{GnT1}^{-/-}$ mice is associated with lactosamine reexpression

The above results demonstrate a unique requirement for  $\beta 3\text{GnT1}$  activity in sensory axon pathfinding and subsequent survival during olfactory development. In adult  $\beta 3\text{GnT1}^{-/-}$  OBs, however, an unexpected reinnervation of OMP and 1B2-reactive axons occurred (Fig. 7*a*). After P15, OMP-labeled axons entered the ventral OB, forming morphologically abnormal glomeruli that were primarily excluded from the most dorsal OB regions. In parallel to the reinnervation, the ratio of OE thicknesses between null and control animals also returned to normal ( $0.97 \pm 0.03$ ;  $39.2 \mu\text{m}$   $\beta 3\text{GnT1}^{-/-}$  vs  $40.5 \mu\text{m}$   $\beta 3\text{GnT1}^{+/-}$ ). Surprisingly, double-label immunofluorescence revealed that all reinnervating OMP<sup>+</sup> fibers in  $\beta 3\text{GnT1}^{-/-}$  OBs expressed lactosamine, unlike the mosaic distribution of 1B2-reactive glomeruli typical of control animals (Fig. 7*a,b*). This expanded distribution suggests that lactosamine was required for either regrowth into the OB or survival of sensory neurons. The mechanism for this lactosamine reexpression remains unclear. Reverse transcription-PCR analysis of mRNA from mutant OEs did not identify alternatively spliced  $\beta 3\text{GnT1}$  transcripts lacking  $\beta$ -geo (data not shown), indicating that secondary changes in other glycosyltransferase activities are a likely mechanism for the lactosamine neosynthesis. The reexpression of lactosamine in  $\beta 3\text{GnT1}^{-/-}$  mice is unique to olfactory neurons, because sensory neurons that lose lactosamine in other regions of the peripheral nervous system do not undergo such compensatory changes (see below).

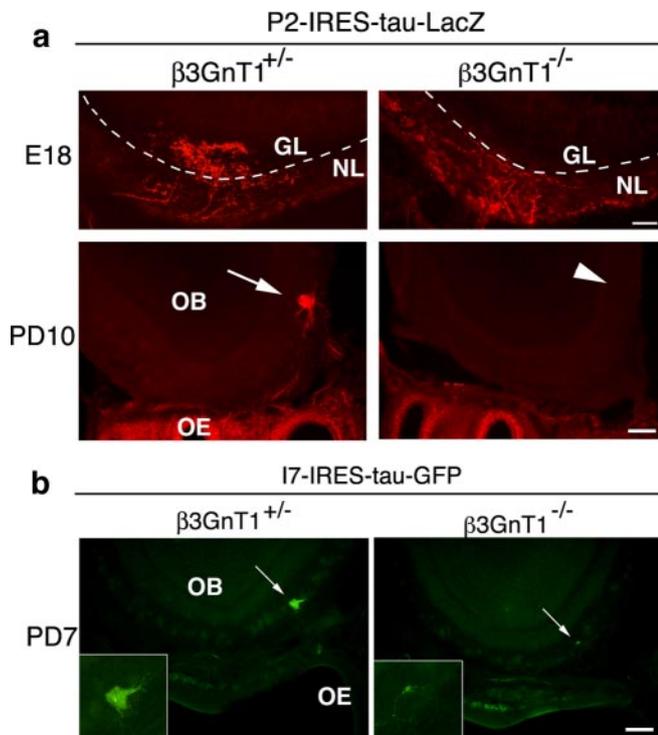
Mice with targeted gene deletions in odorant receptor signaling components frequently die from an inability to recognize olfactory cues required for suckling (Brunet et al., 1996; Belluscio et al., 1998; Wong et al., 2000). In contrast, newborn  $\beta 3\text{GnT1}^{-/-}$  pups nursed normally, indicating some level of olfactory function. The aberrant innervation of the glomerular layer in mutant mice, however, affected smell perception in food-finding tests of olfactory function. Between P14 and P28, fasted null mice took significantly longer than control littermates to locate hidden food rewards (Fig. 7*c*). Interestingly, mutant mice regained near-normal perfor-



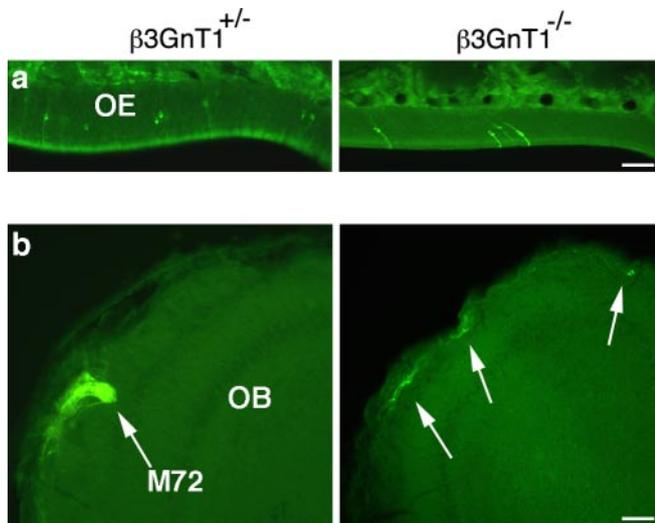
**Figure 3.** Apoptosis and compensatory neurogenesis in  $\beta 3\text{GnT1}$  null mice. *a*, TUNEL analysis of neonatal OE cell death, showing increased sensory neuron apoptosis in P1 null mice. Most TUNEL<sup>+</sup> cells are found at more apical levels of the OE, suggesting that loss of lactosamine does not induce cell death of neuronal progenitors located in the basal cell layer. The dashed line marks the boundary between OE and the underlying lamina propria (LP). *b*, BrdU analysis of cell division in the P10 OE. The proliferation of neural progenitors in the basal OE is dramatically increased in  $\beta 3\text{GnT1}^{-/-}$  mice 1 week after the peak of apoptosis shown in *a*. Scale bars: *a, b*, 25  $\mu\text{m}$ .



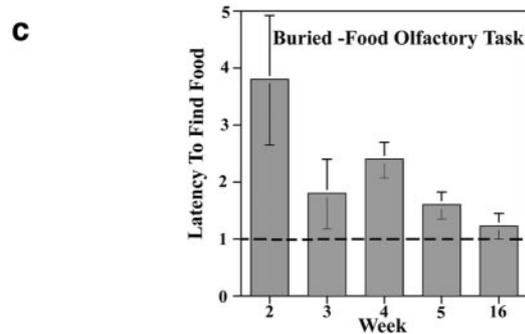
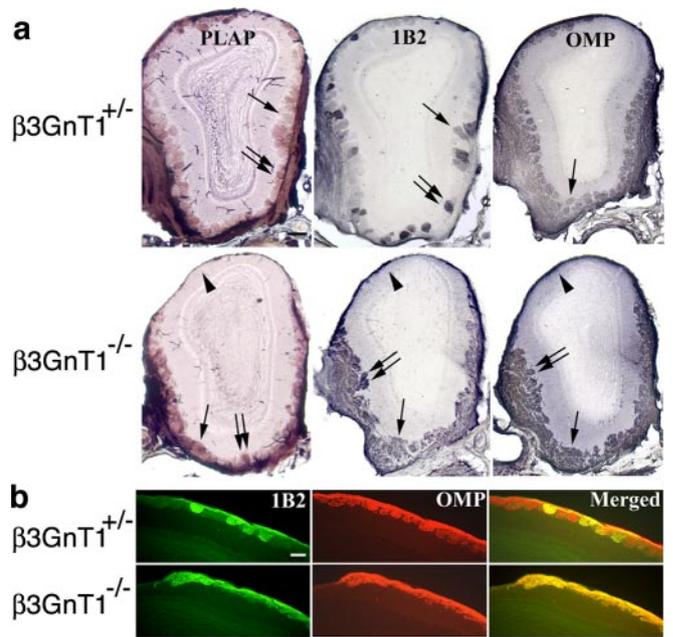
**Figure 4.** P2 odorant receptor-expressing sensory neurons are progressively lost in null mice. *In situ* hybridization for P2 odorant receptor mRNA expression in the OE of  $\beta 3\text{GnT1}$  mice at different ages. The P2 odorant receptor-expressing subset is restricted to zone III of the OE, located at an intermediate position along the dorsoventral axis of the nasal cavity (Wang et al., 1998). The number of neurons expressing the P2 odorant receptor declines progressively, beginning in late embryonic development and continuing into adulthood. In adult null mice, <15% of P2 neurons remain. Scale bars: top, 100  $\mu\text{m}$ ; middle, 200  $\mu\text{m}$ ; bottom, 100  $\mu\text{m}$ .



**Figure 5.** P2 and I7 glomeruli fail to form in  $\beta3GnT1^{-/-}$  mice. **a**, Targeting of the OB by P2 axons in E18 and P10  $\beta3GnT1$ /P2-IRES-tau-LacZ mice, visualized by anti- $\beta$ -galactosidase immunoreactivity. At E18,  $\beta3GnT1^{-/-}$  P2 axons fail to grow through the nerve layer (NL) into the forming glomerular layer (GL) (top). P2 axons innervate the medial OB glomerulus (arrow) in P10  $\beta3GnT1^{+/-}$  mice but are completely absent from the glomerular layer of  $\beta3GnT1^{-/-}$  mice. The arrowhead denotes the approximate position of the missing P2 glomerulus in the mutant OB. **b**, Medial I7-glomeruli in P7  $\beta3GnT1$ /I7-IRES-tau-GFP mice, visualized by GFP fluorescence. Arrows indicate the position of the I7 glomerulus in  $\beta3GnT1^{+/-}$  mice and several innervating I7 fibers in the same region of  $\beta3GnT1^{-/-}$  mice. Inset shows higher-magnification of images in each panel of **b**. Scale bars: **a**, top, 25  $\mu$ m; bottom, 200  $\mu$ m; **b**, 200  $\mu$ m.



**Figure 6.** Axon guidance defects in the M72-odorant receptor neuron subset of  $\beta3GnT1^{-/-}$  mice. **a**, GFP<sup>+</sup> M72 neurons along the P10 dorsal septum of the M72-IRES-tau-GFP reporter line crossed to  $\beta3GnT1^{+/-}$  and  $\beta3GnT1^{-/-}$  mice. Note that the OE is significantly thinner at this age in mutant mice compared with the control OE. **b**, In a coronal section through the wild-type OB, M72 axons converge on a glomerulus on the dorsomedial surface of the extreme caudal OB. In  $\beta3GnT1$  mutant mice, at the same caudal OB position as controls, M72 axons primarily remain in the nerve layer and fail to converge on their typical glomerular position (arrows). Some M72 axons continue to grow past their normal termination site to the dorsalmost extent of the OB. Scale bars: **a**, 25  $\mu$ m; **b**, 50  $\mu$ m.

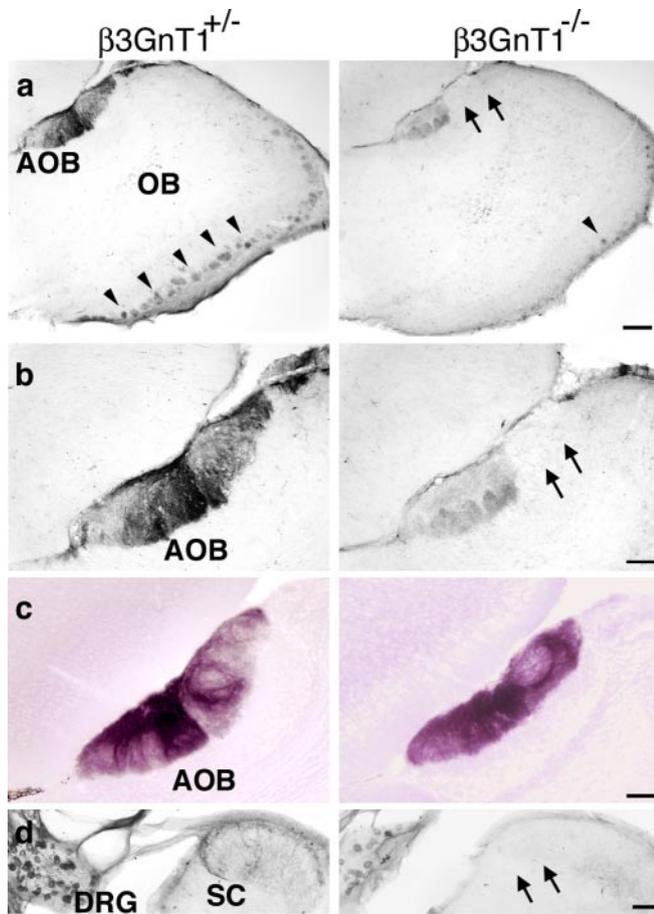


**Figure 7.** Expression of lactosamine in adult  $\beta3GnT1$  null mice permits OB reinnervation and normal olfactory perception. **a**, Histochemical staining for PLAP activity and ICC analysis of 1B2 and OMP expression. Glomeruli (arrows) are grossly abnormal in mutant mice and mostly excluded from the dorsal OB (arrowheads). Scale bar, 200  $\mu$ m. **b**, Double-label immunofluorescence of 1B2 (green), OMP (red), and both (overlay) in sagittal P25 OB sections. 1B2 is heavily expressed by all reinnervating fibers in null mice. Dorsal is up, and rostral is to the right. Scale bar, 100  $\mu$ m. **c**, Olfactory performance of  $\beta3GnT1^{-/-}$  mice relative to control littermates in food-finding task. Null mice remain significantly impaired at this task until adult ages, after lactosamine reexpression permits OB innervation. The dashed line denotes the baseline performance level of control littermates at each age.

mance in this test over a time course that paralleled lactosamine reexpression. By P35, null mice were only slightly delayed in food finding, and adult  $\beta3GnT1^{-/-}$  mice were not statistically different from controls. Therefore,  $\beta3GnT1$  ablation induced a transient smell perception deficit that abated after lactosamine reexpression, despite the formation of an extremely disorganized glomerular layer and the loss of some odorant receptor subsets. It is likely that the regeneration of other OMP<sup>+</sup> populations allows sufficient recovery of olfactory perception for the detection of complex food odors that are central to survival. More extensive analysis with refined odorant compounds may, however, reveal residual deficits in olfactory perception in mutant mice, as the disorganization in the adult regenerated OB would suggest.

**Loss of lactosamine and defects in other sensory areas of  $\beta3GnT1^{-/-}$  mice**

$\beta3GnT1$  disruption also perturbed normal development in other sensory systems. Vomeronasal neurons at all developmental



**Figure 8.**  $\beta 3\text{GnT1}$  determines lactosamine expression in other sensory areas. *a*, 1B2 labeling in sagittal P10 OB sections. Positions of  $1\text{B2}^+$  glomeruli in the ventral OB (arrowheads) and absent 1B2-labeling in  $\beta 3\text{GnT1}$  null sections (arrows) are noted. *b*, Higher-magnification view of the AOB from *a*, showing detail of innervation by lactosamine-expressing axons. *c*, PLAP histochemical labeling of axons in AOB sections from the mouse shown in *b*, showing complete correspondence between  $\beta 3\text{GnT1}$ -expressing fibers and 1B2 reactivity. *d*, 1B2 analysis of P10 DRG neurons and fibers innervating the dorsal horn of the spinal cord, which remains unlabeled in null mice. Unlike the OE, lactosamine is not reexpressed in either the AOB or DRG in adult null mice. Scale bars: *a*, 200  $\mu\text{m}$ ; *b–d*, 100  $\mu\text{m}$ .

stages strongly expressed  $\beta 3\text{GnT1}$  (Fig. 1*c*). In control mice, axons from apical and basal vomeronasal compartments, which project to anterior and posterior accessory olfactory bulb (AOB) domains, respectively, were 1B2-reactive (Fig. 8*a,b*). Lactosamine expression in  $\beta 3\text{GnT1}^{+/-}$  AOBs colocalized completely with PLAP-expressing vomeronasal axons (Fig. 8*c*). In  $\beta 3\text{GnT1}^{-/-}$  mice, residual  $1\text{B2}^+$  fibers remained only in the posterior AOB. The area innervated by PLAP-expressing axons was 25% smaller in  $\beta 3\text{GnT1}^{-/-}$  mice, implying a defect in AOB morphogenesis or, potentially, in AOB glomerular formation.

1B2 reactivity in DRGs was also primarily dependent on  $\beta 3\text{GnT1}$  expression. Lactoseries carbohydrates define a subpopulation of small and intermediate-diameter DRG neurons involved in transmission of cutaneous sensory information (Dodd and Jessell, 1985).  $\beta 3\text{GnT1}$  was strongly expressed in a subpopulation of DRG neurons at P10.  $\beta 3\text{GnT1}$  null mice had reduced 1B2 expression in DRG neurons and an absence of  $1\text{B2}^+$  axon collaterals projecting into the dorsal horn of the spinal cord (Fig. 8*d*).

## Discussion

### Lactosamine glycans have novel functions in sensory system development

We have identified a glycosyltransferase,  $\beta 3\text{GnT1}$ , which is expressed predominantly by subsets of sensory neurons in the olfactory system, in cranial ganglia, and in DRGs. We also show that the expression of 1B2-reactive lactosamine glycans in the OE and these other sensory neurons is directly determined by  $\beta 3\text{GnT1}$  enzymatic activity. The loss of lactosamine in  $\beta 3\text{GnT1}$  mutant mice prevents olfactory projections from making connections with their intended synaptic partners in the OB. The results suggest that lactosamine may play a direct adhesive or signaling role in mediating axon–axon or axon–matrix interactions during development, and they provide strong genetic evidence for a cell-intrinsic function for glycans in axon pathfinding.

A plausible mechanism for such a function could involve galectins, a family of lactosamine-binding proteins, because several members of this family are expressed in the axon pathways (Mahanthappa et al., 1994; Storan and Key, 2004). In the olfactory system, galectin-1 is expressed by ensheathing glial cells in axon tracts between the OE and OB and is capable of modulating olfactory axon–axon interactions *in vitro* (Mahanthappa et al., 1994). Interestingly, galectin-1 was also postulated to participate in guidance of lactosamine-expressing DRG neurons to specific laminae of the spinal cord (Regan et al., 1986).

Alternatively, it is also possible that the loss of lactosamine affects the function of glycoproteins required for olfactory signal transduction. Genetic deletion of type-3 adenylyl cyclase, olfactory cyclic nucleotide-gated channel subunit 1, and odorant receptors each produce OB innervation defects that are similar in some respects to those reported here for  $\beta 3\text{GnT1}^{-/-}$  mice. Although it is clear from our analysis that some odorant receptors do not express lactosamine (see below), each of these signaling proteins is glycosylated (Gat et al., 1994; Bonigk et al., 1999; Wong et al., 2000). The particular glycans they express and their functional significance are now only beginning to be addressed. For example, complete removal of the single N-terminal glycosylation site of mOR-EG affects surface presentation of this odorant receptor *in vitro* (Katada et al., 2004). The loss of terminal lactosamine would be unlikely to perturb protein folding required for trafficking to the cell surface, however, because it would only be expected to affect late Golgi glycan processing. A more plausible lactosamine requirement could be in surface signaling complex assembly or dynamics. In this regard, decreases in lactosamine expression have been shown recently to influence the half-life of some growth factor receptors through the inhibition of endocytosis, an effect also ascribed to galectin interactions (Partridge et al., 2004). The possibility that lactosamine regulates surface expression of some components of the olfactory signaling apparatus awaits a detailed biochemical analysis of the glycosylation and stability of these glycoproteins in  $\beta 3\text{GnT1}$  null mice.

It is unclear whether glycosylation defects in one or multiple glycoproteins leads to the olfactory phenotype in  $\beta 3\text{GnT1}^{-/-}$  mice. Lactosamine is a common constituent of complex N-glycans in all cells but is typically capped by sialic acid or other saccharides that prevent 1B2 binding and contribute to masking of reactivity in tissues such as the CNS (Fig. 1*a*). Desialylation of tissue sections leads to broad 1B2 reactivity in the OB and brain (Schwartz et al., 1992), suggesting that the selective expression of lactosamine by sensory neurons is unique. This is consistent with specialized functions for lactosamine in sensory develop-

ment, which may reflect both tissue-specific  $\beta 3\text{GnT1}$  expression and the overt absence of additional lactosamine processing.

### **$\beta 3\text{GnT1}$ loss has heterogeneous effects on olfactory axon targeting**

1B2 reactivity in the postnatal OE is restricted to a population of differentiated apical sensory neurons and their axons, which suggests that lactosamine regulates connectivity with targets in the OB by affecting either axon sorting within the nerve layer or penetration into the glomerular layer. Severe deficits in OB innervation and glomerular formation were observed in three separate odorant receptor subsets, for which the timing and nature of the defects were heterogeneous. P2 axons from embryonic  $\beta 3\text{GnT1}^{-/-}$  mice primarily targeted the appropriate region of the OB but did not penetrate into the glomerular layer. Similarly, only a subpopulation of I7 axons reached the correct region of the nerve layer by P7, although a few fibers did innervate the presumptive I7 glomerulus. In both subsets, projections were not maintained after P10, suggesting that the absence of synaptic contact with mitral and tufted cells destabilized existing OB connections, compromising glomerular formation and olfactory neuron survival throughout postnatal life.

M72 neurons, however, exhibited a more specific axon guidance defect compared with P2 and I7 neurons. GFP-labeled M72 fibers projected diffusely to a large area of the dorsal posterior nerve layer and did not innervate glomeruli, indicating that lactosamine is specifically required for the targeting of some olfactory axon subsets. Heterogeneous effects on glomerular formation have been reported previously among different neuron subsets, which may reflect intrinsic differences in OE zone of receptor expression or the timing and region of OB innervation (Zheng et al., 2000; Potter et al., 2001; Yu et al., 2004). In  $\beta 3\text{GnT1}$  null mice, differences between subsets may also result from differential expression of lactosamine by neuron populations. The pattern of 1B2 reactive glomeruli is normally stereotyped in adult mice, with large variations in lactosamine expression among individual glomeruli (Fig. 7a) (Crandall et al., 2000). For example, P2 axons, which are lactosamine-positive, project to the anterior, ventral OB, and form glomeruli early in development. In contrast, M72 axons, which are lactosamine negative, project to the posterior, dorsal OB and form glomeruli several days after P2 glomeruli (Potter et al., 2001).

Interestingly, M72 neurons and their projections appear more resilient in postnatal  $\beta 3\text{GnT1}^{-/-}$  mice than neurons of the P2 and I7 subsets, because GFP-labeled M72 fibers are at least detectable in the postnatal nerve layer. M72 axons grow to and beyond their intended targets in the OB, suggesting that lactosamine influences guidance of axon populations that do not directly express lactosamine. This is very similar to axon guidance mistakes in  $\text{Sema3A}^{-/-}$  mice, in which axons that do not express neuropilin-1, the  $\text{Sema3A}$  receptor, are also often misguided because of perturbations caused by severe misrouting of neuropilin-1<sup>+</sup> axons (Schwarting et al., 2004). It may be that M72 axons fail to converge on their correct target because of secondary changes in the nerve and glomerular layers after the loss of other neuron populations and their projections early in development.

### **Factors that influence survival and regeneration of $\beta 3\text{GnT1}^{-/-}$ sensory neurons**

In P10 mutant mice, several residual, 1B2-immunoreactive glomeruli are visible in the OB (Fig. 2b). This implies that an alternative mechanism for producing lactosamine exists in a small

subset of olfactory neurons, potentially the same pathway that is upregulated during the regeneration phase. This observation also suggests that lactosamine confers the ability of olfactory axons to grow into the OB, to form glomeruli, and to survive. It also raises the question of how many odorant receptors remain and what factors are responsible for reconstituting the adult olfactory system in  $\beta 3\text{GnT1}$  adult mutants, in which all axons express lactosamine. Perhaps olfactory neurons in adult  $\beta 3\text{GnT1}$  mutants express a reduced repertoire of odorant receptor subtypes, which survive and form connections, in part, because they are able to activate this alternative lactosamine synthesis mechanism.

The defects in embryonic development described here show similarities to other rodent olfactory degeneration models that show that survival of olfactory sensory neurons requires trophic signals derived from the OB (Schwob et al., 1992; Holcomb et al., 1995; Weiler and Farbman, 1999). In  $\beta 3\text{GnT1}$  null mice, ~30% of P2 and I7 neurons are lost embryonically, before mature glomerular formation and exposure to the external odor environment. This prenatal deletion is distinct from null phenotypes reported for global ablation of olfactory signal transduction genes, which widely fail to alter neuronal subsets or disrupt initial glomerular targeting (Brunet et al., 1996; Belluscio et al., 1998; Zhao and Reed, 2001), and suggests that lactosamine interactions are required for axons to connect with and obtain trophic support from targets in the embryonic OB. Postnatally, the absence of synaptic contact in  $\beta 3\text{GnT1}^{-/-}$  sensory neurons precludes odor-evoked activity for the majority of axons that do not enter the glomerular layer, which likely further compromises neuronal survival.

It is less clear why the three specific subsets of neurons we examined in reporter mice crossed to  $\beta 3\text{GnT1}$  nulls continue to be lost during the regeneration phase, whereas other OMP<sup>+</sup> neuron populations survive to repopulate the OE. Genetic studies have shown that neuron subsets that are deficient in synaptic activity, either through loss of olfactory signal transduction (Zhao and Reed, 2001) or through inhibition of synaptic vesicle release (Yu et al., 2004), are gradually deleted from the odorant receptor repertoire through failed competition with functionally active neurons. These observations are relevant to the recovery of individual neuronal populations observed in  $\beta 3\text{GnT1}$  null mice, in which, despite the widespread reexpression of lactosamine, the population of P2 and I7 odorant receptor neurons decreased postnatally until <15% of the normal number remained in adults. One possibility is that variations in sensory experience attributable to differences in environmental odor ligand exposure could lead to variations in recovery, a hypothesis previously suggested from chemical lesioning models of olfactory regeneration (Iwema et al., 2004). It is also possible that some subsets of sensory neurons are unable to *de novo* express lactosamine during the regeneration phase. Finally, we cannot exclude the possibility that some sensory neurons are lost from detection through odorant receptor switching instead of cell death (Serizawa et al., 2003).

### **$\beta 3\text{GnT1}$ and other sensory systems**

The retinotectal system has often been used as a model to better understand mechanisms used to construct connections in the developing olfactory system (Key and St John, 2002). Studies presented here suggest that it may be helpful to consider the patterning of DRG projections to the dorsal horn of the spinal cord as an alternative model. Interestingly, there are parallels between axon targeting by OE neurons to the OB and nociceptive DRG neurons to the spinal cord. Both create spatial order by converging input from dispersed sensory neurons to specific tar-

get zones (OB glomeruli and dorsal horn laminae). It will be informative to determine whether the topography of DRG projections in  $\beta 3\text{GnT1}^{-/-}$  mice is disrupted similarly to the abnormal olfactory connectivity described here. Furthermore, because  $\beta 3\text{GnT1}$  is expressed by subsets of other cranial neurons, including trigeminal and glossopharyngeal neurons, it will be of additional significance to contemplate a conserved function for lactosamine in building circuitry in other sensory systems.

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