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Leslie A. Soyka

University of Massachusetts Medical School

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The Effects of Anorexia Nervosa on Bone Metabolism in Female Adolescents*

LESLIE A. SOYKA, STEVEN GRINSPoon, LYNEE L. LEVITSKY, DAVID B. HERZOG, AND ANNE KLIBANSKI

Neuroendocrine Unit (L.A.S., S.G., A.K.), Pediatric Endocrine Unit (L.A.S., L.L.L.), and Eating Disorders Unit (D.B.H.), Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts 02114

ABSTRACT

Osteopenia is a frequent, often persistent, complication of anorexia nervosa (AN) in adolescent girls and occurs during a critical time in bone development. Little is known about bone metabolism in this patient population. Therefore, we measured bone density (BMD) and body composition by dual energy x-ray absorptiometry, nutritional status, bone turnover, calcium, and hormonal status in 19 adolescent girls with AN (mean ± SEM, 16.0 ± 0.4 yr) and 19 age-matched controls. The mean duration of AN was 19 ± 5 months. Spinal (L1–L4) osteopenia was common in AN. Lumbar anteroposterior BMD was more than 1 SD below the mean in 42% of patients, and lateral spine BMD was more than 1 SD below in 63% of patients compared with controls. Lean body mass significantly predicted lumbar bone mineral content (r = 0.75; P < 0.0001) in controls only. In AN, duration of illness was the most significant predictor of spinal BMD (lumbar: r = −0.44; P = 0.06; lateral: r = −0.59; P = 0.008). AN adolescents with mature BA (15 yr and greater) were hypogonadal [estradiol, 16.2 ± 1.9 vs. 23.3 ± 1.6 pg/mL (P = 0.01); free testosterone, 0.70 ± 0.17 vs. 1.36 ± 0.14 pg/mL (P = 0.01)] although dehydroepiandrosterone sulfate and urinary free cortisol levels did not differ. Leptin levels were reduced in AN (2.9 ± 2.1 vs. 16.5 ± 1.8 ng/mL; P < 0.0001). Insulin-like growth factor I (IGF-I) was reduced in AN to 50% of control levels (219 ± 41 vs. 511 ± 35 ng/mL; P < 0.0001) and correlated with all measures of nutritional status, particularly leptin (r = 0.80; P < 0.0001). Surrogate markers of bone formation, serum osteocalcin (OC) and bone-specific alkaline phosphatase (BSAP), were significantly (P = 0.02) reduced in AN vs. controls (OC, 39.1 ± 6.4 vs. 59.2 ± 5.2 ng/mL; BSAP, 27.9 ± 4.0 vs. 40.6 ± 3.4 U/L). The majority of the variation in bone formation in AN was due to IGF-I levels (OC: r² = 0.72; P = 0.002; BSAP: r² = 0.53; P = 0.01) in stepwise regression analyses. Bone resorption was comparable in patients and controls. These data demonstrate that bone formation is reduced and uncoupled to bone resorption in mature adolescents with AN in association with low bone density. Lean body mass was a significant predictor of BMD in controls, but not AN patients. The major correlate of bone formation in AN was the nutritionally dependent bone trophic factor, IGF-I. Reduced IGF-I during the critical period of bone mineral accumulation may be an important factor in the development of osteopenia in adolescents with AN. (J Clin Endocrinol Metab 84: 4489–4496, 1999)

ANOREXIA NERVOSA (AN) is an increasingly common disorder in adolescent girls, with a prevalence of 0.2–1.0% in Western societies (1, 2). Osteopenia is a frequent and often persistent complication of AN in this younger population (3–6), leading to clinical fractures and increased fracture risk throughout life (7, 8). A severe degree of decreased bone mass has been reported even in young adolescents with a brief duration of illness (3). Adolescence represents a critical time in bone metabolism, as most bone mineral is accumulated during the teenage years, and deficits incurred during this time may be permanent. Therefore, an understanding of the factors that impact bone density during this time period is of particular importance.

Bone density and bone metabolism change dramatically during adolescence, and the onset of AN during this critical time may interfere with achievement of peak bone mass. In addition, significant changes in body weight and composition, pubertal development, and pubertal hormones, such as estradiol and insulin-like growth factor I (IGF-I), occur in AN and may impact bone metabolism. The effect of AN on bone turnover in adolescent girls is poorly understood. Limited data are available (9, 10), and prior studies in adolescents with AN have been limited by the small numbers of subjects and the lack of information about specific biochemical markers of bone turnover, including simultaneous measurements of formation and resorption. Moreover, previous studies have included subjects of a broad age range from early adolescence to young adulthood. Finally, pubertal and skeletal maturation have not been measured in assessments of bone density, even though bone density correlates more with pubertal stage and bone age (BA) than chronological age (CA) (11, 12).

The purpose of this study was to further characterize bone metabolism and potential mechanisms of osteopenia in adolescents with AN in a cohort of adolescent females in whom bone density, biochemical indices of bone turnover, nutritional status, and pubertal hormones were determined compared with those in BA-matched controls. Our data demonstrate significant osteopenia in adolescent girls with AN. Bone formation indices are significantly reduced in association with IGF-I and other nutritionally dependent factors in this population of patients, suggesting a profound effect of this disease on osteoblast function.

Subjects and Methods

We studied 38 Caucasian adolescent girls, 19 with AN, and 19 healthy adolescents. The AN patients were 12.8–18.5 yr of age. All met DSM IV...
obtained from all subjects and their parents. Informed consent was

ported in 42% of control subjects, but none occurred within 6 months of

strual periods. None of the controls had a history of an eating disorder.

was premenarchal. All postmenarchal adolescents had regular men-

kilograms per meter squared) between the 25th-90th percentile for age

and through collaboration with eating disorder programs in the greater

Boston area.

A control group of 19 healthy Caucasian female adolescents, aged

13.1–17.8 yr, was recruited by advertisement in primary care providers’
offices and community newspapers. All had a body mass index (BMI; kilograms per meter squared) between the 25th-90th percentile for age
based on published standard charts (13). One control subject, 13.1 yr old,

was premenarchal. All postmenarchal adolescents had regular men-

strual periods. None of the controls had a history of an eating disorder.

Control subjects with a medical condition or receiving hormonal or other
medications known to affect bone metabolism were excluded. A history of
traumatic bone fracture during childhood or adolescence was re-

ported in 42% of control subjects, but none occurred within 6 months of

the study.

The study protocol was approved by the Massachusetts General Hospital subcommittee on human subjects, and informed consent was

obtained from all subjects and their parents.

Experimental protocol

All subjects were evaluated during an out-patient visit at the General Clinical Research Center at Massachusetts General Hospital after a

screening visit to determine eligibility and give instruction on collection of

urine samples and completion of 4-day food records. A medical and

menstrual history and physical examination were performed by a single

investigator. All control subjects were studied during the early follicular

phase of the menstrual cycle to control for the effects of cycle stage on

biochemical and hormonal measurements. All AN and control subjects

had their BA determined by x-ray of the left hand and wrist by a single

investigator using the methods of Greulich and Pyle (14).

Subjects’ height was measured using a single stadiometer (average of
triplicate measurements), and weight was measured in a hospital gown

on an electronic scale. BMI was calculated, and BMI percentile was
determined from published charts (13).

Subjects collected a 24-h and a 2-h second morning urine sample at

home. All had fasting blood samples drawn at the same time of day

(afternoon). Urine and serum samples were obtained for biochemical and

hormonal measurements as outlined below. Bone density and body

composition were assessed in all subjects.

Classification of subjects by bone age

For the analysis of biochemical parameters that are dependent on

pubertal stage, such as gonadal steroids and markers of bone formation

and resorption, we subdivided groups by maturity. Previous studies

reported significant changes in bone turnover markers between girls in

Tanner stages 2–3 and 4–5 (15, 16). Tanner breast staging is difficult to

assess in females with AN who are hypoestrogenemic and have breast

atrophy, whereas BA provides more objective information on past and

current maturation. We therefore subdivided subjects into immature

(BA, <15 yr) and mature (BA, ≥15 yr) groups. We used the cut-off BA

of 15 yr because this BA is mature, in that all epiphyses are near fusion,

and 99% of final adult height has been attained according to the stan-

dards of Greulich and Pyle (14). Biochemical data are available for the

mature group only, because there were insufficient numbers of immu-

nate subjects based on initial power calculations.

Biochemical assessment

Serum ionized calcium, phosphorus, TSH, dehydroepiandrosterone

sulfate, and 24-h urine free cortisol (UFC), calcium, and creatinine were

measured through the hospital laboratory using previously de-

scribed methods (17).

Serum estradiol (E2) was measured by ultrasensitive RIA (Diagnostics

Systems Laboratories, Inc., Webster, TX). The detection limit of the E2

assay was 2.2 pg/mL, and the coefficient of variation is 6.5–8.9%. Total and

free testosterone (T) were measured by RIA (Coat-A-Count, Diagnostic

Products, Los Angeles, CA). Total T had a detection limit of 20 ng/dL

and an intraassay coefficient of variation of 10–15%, and free T had a
detection limit of 0.57 pg/mL and an intraassay coefficient of variation of

3.2–4.3%. Serum IGF-I was measured by acid-alcohol extraction using a

RIA kit (Nichols Institute Diagnostics, San Juan Capistrano, CA), with

a detection limit of 0.06 ng/mL and an intraassay coefficient of variation

2.4–3.0%. Serum leptin was measured by RIA (Linco Research, Inc., St.

Louis, MO) with a sensitivity of 0.5 ng/mL and an intraassay coefficient

of variation of 3.4–8.3%. Serum 25-hydroxyvitamin D was measured by

RIA (DaSorin, Inc., Stillwater, MN), with a detection limit of 3 ng/mL

and a coefficient of variation of 10–15%. Serum intact PTH was mea-
sured by a two-site immunoradiometric assay (Nichols Institute Dia-

gnostics, San Juan Capistrano, CA) with a detection limit of 1 pg/mL and

an intraassay coefficient of variation of 1.8–3.4%. Serum osteocalcin (OC)

was measured using an immunoradiometric assay (Nichols Institute Di-

agnostics) with a sensitivity of 0.5 ng/mL and an intraassay coefficient

of variation of 3.2–5.2%. Bone-specific alkaline phosphatase (BSAP) was

measured by immunoassay (Metra Biosystems, Mountain View, CA)

with a sensitivity of 0.7 U/L and a coefficient of variation of 3.9–5.8%.

Carboxy-terminal propeptide of type 1 procollagen was measured by

RIA with an intraassay coefficient of variation of 2.1–3.2% (INCASTER

Corp., Stillwater, MN). Urinary deoxypyridinoline (DPD) and

N-telopeptide (NTX) were measured in a 2-h second morning urine

sample normalized for creatinine excretion. DPD was measured by

competitive enzyme-linked immunosorbent (EIA) assay with a detection

limit of 1.1 nmol/L and an intraassay coefficient of variation of

4.3–8.4%. NTX was measured by enzyme-linked immuno-

sorbent assay (Ostex International, Inc., Seattle, WA) with a detection

limit of 20 nmol BCE and an intraassay coefficient of variation of 5–19%.

All samples were run in duplicate using a single kit.

Bone density and body composition

Total body and lumbar spine (L1–L4) bone densities were measured using a QDR-4500 (Hologic, Inc., Waltham, MA). The sn for bone density

measurement is 0.01 g/cm² and does not vary with bone density.

Whole body dual energy x-ray absorptiometry (DEXA) was per-

formed using a Hologic, Inc. QDR-4500 densitometer to determine fat

and lean body mass, which has been validated in children and young

adults (18, 19).

Nutritional assessment

Nutritional intake data, including calorie, protein, fat, carbohydrate,

and micronutrient content, were obtained through computer analysis

(version 2, Nutrition Data Systems, Minneapolis, MN) of 4-day food

records and history of dietary intake over the month before the study

visit. Resting metabolic energy expenditure was performed using a

metabolic cart (20). Typical exercise and activity of subjects over the past

year were determined using a validated questionnaire (21).

Statistical analysis

Clinical, bone density, body composition, and biochemical data were

analyzed with Student’s t test and compared by linear regression anal-

ysis. Stepwise regression analyses were performed to determine the

relationship of anthropometric, body composition, and hormonal data
to bone density and bone turnover. Significance was set at P < 0.05 for

all comparisons. Results are the mean ± sem unless otherwise indicated.

Results

Clinical and anthropometric data

Clinical and anthropometric data for patients and controls are summarized in Table 1. Patients were matched for BA (15.7 vs. 15.5 yr) with control subjects. CA was not signifi-
cantly different between the AN and controls groups. How-
ever, BA compared with CA (BA/CA) was significantly
lower in the AN subjects compared with controls (AN, 0.98 ± 0.01; control, 1.03 ± 0.01; P = 0.01). Duration of AN was 19 ± 5 months, and duration of amenorrhea was 24 ± 4 months. Height was not significantly different between the groups. AN patients were of significantly lower weight and BMI. In the AN group, fat mass measured by DEXA was less than 50% that in controls (AN, 7.7 ± 0.8; control, 19.6 ± 0.8 kg; P < 0.0001), and percent body fat was substantially reduced (AN, 0.87 ± 1.3%; control, 33.5 ± 1.3%; P < 0.0001). Lean body mass was also significantly lower in AN subjects (AN, 33.1 ± 1.0; control, 37.1 ± 1.0 kg; P = 0.01) even when controlled for height and body weight.

Historical assessment of activity level over the past year demonstrated that AN subjects were significantly more physically active than controls, with approximately twice as many hours of physical activity per week (AN, 17.0 ± 2.2; control, 9.4 ± 2.2 h/week; P = 0.02). Resting metabolic energy expenditure was significantly lower in AN subjects (AN, 1314 ± 60; control, 1587 ± 60 KCal/day). No subjects regularly smoked cigarettes or consumed alcohol.

Table 1. Clinical and anthropometric data

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa (n = 19)</th>
<th>Control (n = 19)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chronological age (yr)</td>
<td>16.0 ± 0.4 [12.8–18.5]</td>
<td>15.1 ± 0.4 [13.1–17.8]</td>
<td>NS</td>
</tr>
<tr>
<td>Bone age (yr)</td>
<td>15.7 ± 0.3 [13–16]</td>
<td>15.5 ± 0.3 [13–18]</td>
<td>NS</td>
</tr>
<tr>
<td>Bone age/chronological age</td>
<td>0.98 ± 0.01</td>
<td>1.03 ± 0.01</td>
<td>0.01</td>
</tr>
<tr>
<td>Ht (cm)</td>
<td>162.3 ± 1.4</td>
<td>164.7 ± 1.4</td>
<td>NS</td>
</tr>
<tr>
<td>Wt (kg)</td>
<td>43.4 ± 1.5</td>
<td>58.6 ± 1.5</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>16.5 ± 0.4</td>
<td>21.8 ± 0.4</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Duration of illness (months)</td>
<td>19 ± 5 [2–72]</td>
<td>9.4 ± 2.2</td>
<td>0.02</td>
</tr>
<tr>
<td>Amenorrhea</td>
<td>No. premenarchal</td>
<td>5</td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>No. primary</td>
<td>2</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>No. secondary</td>
<td>12</td>
<td>0</td>
</tr>
<tr>
<td>Duration (months)</td>
<td>24 ± 4 [6–48]</td>
<td>9.4 ± 2.2</td>
<td>0.02</td>
</tr>
<tr>
<td>Activity (h/week)</td>
<td>17.0 ± 2.2</td>
<td>9.4 ± 2.2</td>
<td>0.02</td>
</tr>
<tr>
<td>MEE (kCal/day)</td>
<td>1314 ± 60</td>
<td>1587 ± 60</td>
<td>0.003</td>
</tr>
</tbody>
</table>

All values are expressed as the mean ± sem. Ranges are in brackets. The P value is for comparison between anorexia nervosa and control subjects by Student’s t test. MEE, Resting metabolic energy expenditure.

Table 2. Daily nutrient intake

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa (n = 17)</th>
<th>Control (n = 18)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Calories (KCal)</td>
<td>1730 ± 162 [668–2913]</td>
<td>2042 ± 162 [1395–4093]</td>
<td>NS</td>
</tr>
<tr>
<td>Fat (g)</td>
<td>41 ± 7 [7–88]</td>
<td>65 ± 7 [26–162]</td>
<td>0.03</td>
</tr>
<tr>
<td>% Cal from fat</td>
<td>20 ± 2 [16–33]</td>
<td>28 ± 2 [16–41]</td>
<td>0.005</td>
</tr>
<tr>
<td>Protein (g)</td>
<td>72 ± 6 [31–128]</td>
<td>70 ± 6 [40–122]</td>
<td>NS</td>
</tr>
<tr>
<td>Calcium (mg)</td>
<td>1297 ± 131 [580–2333]</td>
<td>1238 ± 128 [589–2708]</td>
<td>NS</td>
</tr>
<tr>
<td>% &lt; RDA</td>
<td>53%</td>
<td>67%</td>
<td>NS</td>
</tr>
<tr>
<td>Vitamin D (IU)</td>
<td>423 ± 62 [112–1212]</td>
<td>302 ± 60 [80–714]</td>
<td>NS</td>
</tr>
<tr>
<td>Phosphorus (mg)</td>
<td>1396 ± 131 [570–2476]</td>
<td>1383 ± 127 [741–2551]</td>
<td>NS</td>
</tr>
<tr>
<td>Magnesium (mg)</td>
<td>368 ± 34 [149–606]</td>
<td>310 ± 33 [173–586]</td>
<td>NS</td>
</tr>
<tr>
<td>Dietary supplementation</td>
<td>Calcium</td>
<td>24%</td>
<td>22%</td>
</tr>
<tr>
<td></td>
<td>Vitamin D</td>
<td>35%</td>
<td>28%</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± sem. Ranges are in brackets. The P value is for comparison between anorexia nervosa and control subjects by Student’s t test.

Bone density data

Bone density data are summarized in Table 3. Lumbar spine (L1–L4) BMD was significantly lower in the AN subjects (AN, 0.87 ± 0.02; control, 0.97 ± 0.02 g/cm²; P = 0.004). Lumbar bone mineral content (BMC) was also reduced in AN (AN, 45 ± 2; control, 51 ± 2 g; P = 0.04), with lumbar area not different between the groups. In 42% of AN subjects lumbar BMD was more than 1 sd below the mean for the healthy, height- and BA-matched controls. Sixteen percent of AN subjects had a lumbar BMD more than 2 sd below the normal mean. Comparison of the subjects’ lumbar spine BMD to the DEXA manufacturer normative data also dem-
TABLE 3. Bone density and body composition data

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa (n = 19)</th>
<th>Control (n = 19)</th>
<th>P value</th>
<th>P value for bt-adjusted BMD</th>
</tr>
</thead>
<tbody>
<tr>
<td>LBMD, L1–4 (g/cm²)</td>
<td>0.87 ± 0.02</td>
<td>0.97 ± 0.02</td>
<td>0.004</td>
<td>0.01</td>
</tr>
<tr>
<td>LBMC, L1–4 (g)</td>
<td>45 ± 2</td>
<td>51 ± 2</td>
<td>0.04</td>
<td>0.03</td>
</tr>
<tr>
<td>Lateral spine BMD (g/cm²)</td>
<td>0.68 ± 0.02</td>
<td>0.81 ± 0.02</td>
<td>&lt;0.0001</td>
<td>0.0002</td>
</tr>
<tr>
<td>Total BMD (g/cm²)</td>
<td>1.01 ± 0.02</td>
<td>1.04 ± 0.02</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>BMC (g)</td>
<td>1700 ± 53</td>
<td>1899 ± 53</td>
<td>0.02</td>
<td>0.02</td>
</tr>
<tr>
<td>Fat (kg)</td>
<td>7.7 ± 0.8</td>
<td>19.6 ± 0.8</td>
<td>&lt;0.0001</td>
<td>NA</td>
</tr>
<tr>
<td>% Body fat</td>
<td>17.7 ± 1.3</td>
<td>33.5 ± 1.3</td>
<td>&lt;0.0001</td>
<td>NA</td>
</tr>
<tr>
<td>Lean (kg)</td>
<td>33.1 ± 1.0</td>
<td>37.1 ± 1.0</td>
<td>0.01</td>
<td>NA</td>
</tr>
</tbody>
</table>

All values are expressed as the mean ± SEM. The P value is for comparison between anorexia nervosa and control subjects by Student’s t test. NA, Not applicable.

onstrated that the AN group had significantly lower lumbar BMD (AN z-score, −1 ± 0.3; controls z-score, 0.2 ± 0.3; P = 0.002), with a z-score below −1 in 50%, and below −2 in 21% of AN patients. The reduction in lateral spine BMD was even more severe in AN (AN, 0.68 ± 0.02; control, 0.81 ± 0.02 g/cm²; P < 0.0001), with BMD more than 1 SD below the normal mean in 63% of patients and more than 2 SD below the normal mean in 26% of patients. Total body BMC (TBMC) was reduced in AN (AN, 1700 ± 53; control, 1899 ± 53 g; P = 0.02) and total body BMD (TBMD) was lower in AN, but not significantly. Differences in spinal BMD and TBMC remained significant when controlled for height. Similar differences in bone mineral apparent density were demonstrated (AN, 0.122 ± 0.003; control, 0.132 ± 0.003 g/cm³; P = 0.02), using the formula of Katzman et al. (22).

In the control group, lean body mass was the single most significant predictor of lumbar BMC (r = 0.75; P < 0.0001) and lateral spine (r = 0.55; P = 0.003) and total (r = 0.54; P = 0.004) BMD. These correlations remained significant when controlled for height and weight, but weight rather than lean body mass was a more significant determinant of lumbar BMD when controlled for height. The relationships between body weight, composition, BMC, and BMD were not present in AN. In AN, duration of illness was the most important determinant of lumbar BMD (r = −0.44; P = 0.06) and BMC (r = −0.42; P = 0.07) and lateral spine BMD (r = −0.59; P = 0.008), but not TBMD or TBMC. In addition, duration of amenorrhea (r = −0.58; P < 0.05) was predictive of lateral spine BMD.

Biochemical data

TSH and serum ionized calcium and phosphorus were normal in all subjects. Calcium excretion in a 24-h urine collection was normal in all subjects, with the exception of one AN subject who was ingesting 2100 mg calcium daily with a dietary supplement. The median 24-h urinary calcium was 144 mg in both groups (range in AN, 71–450 mg; in control, 39–217 mg). UFC in a 24-h collection was not significantly different between groups (AN, 59 ± 7; control, 51 ± 7 μg), although 33% of AN subjects (6 of 18) had modest elevations in UFC.

Serum PTH levels were normal in all subjects and were not different between the groups (Table 4). Serum 25-hydroxyvitamin D levels were lower in the control group, but normal in all subjects, and correlated with dietary vitamin D intake (r = 0.41; P = 0.03).

TABLE 4. Serum hormone concentrations

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa (n = 11)</th>
<th>Control (n = 15)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>25-Hydroxyvitamin D (ng/mL)</td>
<td>37 ± 3</td>
<td>29 ± 3</td>
<td>0.05</td>
</tr>
<tr>
<td>PTH (pg/mL)</td>
<td>24.8 ± 3.6</td>
<td>32.8 ± 3.1</td>
<td>NS</td>
</tr>
<tr>
<td>Estradiol (pg/mL)</td>
<td>16.2 ± 1.9</td>
<td>23.3 ± 1.6</td>
<td>0.01</td>
</tr>
<tr>
<td>Total testosterone (ng/dL)</td>
<td>23.6 ± 3.4</td>
<td>25.6 ± 2.9</td>
<td>0.06</td>
</tr>
<tr>
<td>Free testosterone (pg/mL)</td>
<td>0.70 ± 0.17</td>
<td>1.36 ± 0.14</td>
<td>0.01</td>
</tr>
<tr>
<td>DHEAS (μg/dL)</td>
<td>223 ± 31</td>
<td>212 ± 26</td>
<td>NS</td>
</tr>
<tr>
<td>IGF-I (ng/mL)</td>
<td>219 ± 41</td>
<td>511 ± 35</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Leptin (ng/mL)</td>
<td>2.9 ± 2.1</td>
<td>16.5 ± 1.8</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

Serum hormone data in mature (bone age of 15 yr or greater) subjects. Values are expressed as the mean ± SEM. The P value is for comparison between anorexia nervosa and control subjects by Student’s t test.

Serum E₅ levels were significantly reduced, even in comparison with controls sampled in the early follicular phase (Table 4). There was a trend toward lower total T levels in AN, and free T levels were markedly reduced (AN, 0.70 ± 0.17; control, 1.36 ± 0.14 pg/mL; P = 0.006), with levels below the lower limit of detection of the assay in 82% of AN subjects.

Serum IGF-I levels were reduced significantly in AN, to less than 50% of those in the control subjects (AN, 219 ± 41; control, 511 ± 35 ng/mL; P < 0.0001). Leptin was also markedly reduced in AN (AN, 2.9 ± 2.1; control, 16.5 ± 1.8 ng/mL; P < 0.0001), to less than 20% of control levels. In all subjects, IGF-I levels correlated highly with nutritional status, including BMI (r = 0.73; P < 0.0001), percent body fat (r = 0.61; P = 0.0008), lean body mass (r = 0.49; P = 0.005), and leptin (r = 0.80; P < 0.0001). In contrast, there was no relationship between IGF-I and gonadal steroid concentrations. In stepwise regression analysis including all anthropometric and biochemical measurements, leptin accounted for 65% of the variation in IGF-I levels (r² = 0.65; P < 0.00001). Leptin was highly dependent on fat mass (r = 0.78; P < 0.0001).

Bone turnover marker results are summarized in Table 5 and Fig. 1. OC and BSAP were significantly lower in AN patients than in normal subjects. DPD and NTX were not significantly different between the groups. However, there was a trend toward lower NTX levels in the AN group. There was a strong correlation between IGF-I and OC in AN subjects (r = 0.71; P = 0.01; Fig. 2), whereas in the control group a similar trend was found, but the correlation did not reach statistical significance (r = 0.46; P = 0.07).
TABLE 5. Bone turnover markers

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa (n = 11)</th>
<th>Control (n = 15)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>OC (ng/mL)</td>
<td>39.1 ± 6.4</td>
<td>59.2 ± 5.2</td>
<td>0.02</td>
</tr>
<tr>
<td>BSAP (U/L)</td>
<td>27.9 ± 4.0</td>
<td>40.6 ± 3.4</td>
<td>0.02</td>
</tr>
<tr>
<td>PICP (ng/mL)</td>
<td>139 ± 12</td>
<td>154 ± 10</td>
<td>NS</td>
</tr>
<tr>
<td>DPD (nmol/mmol creatinine)</td>
<td>13.4 ± 1.4</td>
<td>15.1 ± 1.2</td>
<td>NS</td>
</tr>
<tr>
<td>NTX (nmol BCE/mmol creatinine)</td>
<td>138 ± 25</td>
<td>200 ± 21</td>
<td>NS</td>
</tr>
</tbody>
</table>

Bone turnover markers in mature (bone age of 15 yr or more) subjects. Values are expressed as the mean ± SEM. The P value if for comparison between anorexia nervosa and control subjects by Student’s t test.

regression analysis, IGF-I accounted for 72% ($r^2 = 0.72; P = 0.002$), and IGF-I and free T together accounted for 95% ($r^2 = 0.95; P = 0.0006$), of the variation in OC levels in AN. There was a similar strong relationship between IGF-I and BSAP in the AN group ($r = 0.73; P = 0.01$; Fig. 2) and the control group ($r = 0.59; P = 0.02$). In stepwise regression analysis, IGF-I was the only significant predictor of BSAP, accounting for 53% ($r^2 = 0.53; P = 0.01$) of the variation in BSAP in AN and 35% ($r^2 = 0.35; P = 0.01$) in controls.

Adult height

Subjects in the mature group, who had a minimum BA of 15 yr, were determined to be at adult height. When adult height was compared to midparental target height, height was significantly reduced in the AN group, on the average 3.0 cm below midparental height (AN, −3.0 ± 1.0; control, 0.2 ± 0.9 cm; $P = 0.02$). Furthermore, only one subject in the AN group exceeded her midparental target height, compared with eight (53%) control subjects.

Discussion

In this study, we have characterized bone density and bone metabolism in adolescent girls with AN compared with those in BA-matched controls. Bone formation indices and bone density are significantly reduced in adolescents with AN. Moreover, low bone formation in this population was strongly associated with markedly diminished IGF-I, a nutritionally dependent endogenous bone trophic factor. In addition, we have shown that in these osteopenic girls the decrease in bone formation is uncoupled from bone resorption.

Our data demonstrate that spinal osteopenia is common in adolescent girls with AN compared with a group of BA- and CA-matched healthy adolescents. In the patient group, 42% of AN subjects had a lumbar spine BMD more than 1 sd, and 16% had a lumbar spinal BMD more than 2 sd, below the mean of the BA- and height-matched controls despite an average duration of illness of only 1.5 yr. The results of lateral spine BMD, which may be more representative of trabecular bone (23), demonstrated even more severe osteopenia, with BMD more than 1 sd below the mean in 63% and more than 2 sd in 26% of AN patients compared with controls. Furthermore, we obtained similar results for height-adjusted and bone mineral apparent density calculations, suggesting that the low BMD was not an artifact of size.

Previous studies of bone metabolism in adolescents with AN have been limited by the small numbers of subjects, inclusion of a broad age range, and lack of information about specific biochemical markers of bone turnover, including measures of both formation and resorption. To assess further the mechanisms of reduced BMD in AN we characterized bone metabolism in adolescents with AN compared with that in a CA- and BA-matched group of healthy adolescents. Studies of adult women with AN demonstrate a severe imbalance in bone turnover, with a decrease in markers of bone formation combined with an increase in markers of bone resorption (24). However, data obtained from women with AN cannot be extrapolated to adolescent patients who are in an active period of bone growth and mineral accretion as well as remodeling. Abrams et al. demonstrated that adolescents with AN have decreased calcium absorption and increased urinary calcium excretion; however, bone markers were not measured (9). Saggese et al. reported low levels of carboxy-terminal propeptide of type I procollagen, a bone formation marker, in six adolescent AN patients ranging from 11–21 yr of age, but did not investigate other markers of formation or markers of resorption in these patients (10).

Matching patients with controls based on stage of maturation is critical in the evaluation of bone metabolism, as both bone formation and resorption markers change dramatically with the transition from early (Tanner stage 2–3) to late (Tanner stage 4–5) puberty (15, 16). Furthermore, levels of bone formation and resorption remain above adult levels even in mature (Tanner stage 5) adolescents (15). We chose to classify maturation in adolescents based on BA because assessment of Tanner stage in adolescents with AN who are hypoestrogenemic is difficult and likely to be inaccurate. In our study, all healthy controls who had a BA of 15 yr or greater had Tanner stage 4–5 development and therefore were classified as mature. In contrast, 35% of AN patients with a mature BA had what appeared clinically to be Tanner stage 3 breast tissue. Therefore, Tanner staging as a measure of pubertal maturation in AN can be inaccurate, whereas BA provides a more objective classification.

Bone formation markers, OC and BSAP, were significantly reduced in mature adolescents with AN in contrast to those in healthy mature adolescents. In contrast, markers of bone resorption, DPD and NTX, were not significantly different, although there was a trend toward lower NTX in AN subjects. Therefore, in healthy adolescents, high levels of bone formation and resorption were seen. In contrast, patients with AN had low bone formation with normal resorption. Although biochemical markers are indirect measures of bone growth and remodeling, resorption indexes are generally thought to reflect degradation of existing bone, whereas formation indices in adolescents are thought to reflect skeletal growth and mineralization. We speculate that in a low metabolic state such as AN, remodeling continues, and bone formation associated with growth and bone mineral accumulation is reduced. We were unable to measure bone metabolism in the immature adolescents due to insufficient numbers of subjects and therefore could not determine whether the balance of bone formation and resorption is also altered in immature patients with AN. Further studies in younger patients will provide insight as to whether reduced
bone formation in AN is dependent on the stage of maturation.

The primary correlate of bone formation in AN was the level of IGF-I, which was markedly reduced in AN subjects, in contrast to the high levels expected and found in normal adolescents. IGF-I functions as a bone trophic hormone, which positively affects bone growth and bone turnover by stimulating osteoblasts, collagen synthesis, and longitudinal bone growth (25, 26). Low levels of serum IGF-I are present in undernutrition and correlate with low BMI in AN (24, 27, 28). We found a high correlation between all indexes of nutritional status, including body composition variables and particularly leptin, with IGF-I levels, indicating that undernutrition is a major cause of reduced IGF-I levels in adolescents with AN. These data indicate that IGF-I deficiency due to undernutrition could significantly contribute to low bone formation in adolescents with AN.

In healthy adolescents, E2 is thought to contribute significantly to bone mineral acquisition, either by acting directly on bone or by stimulating other mediators of bone growth (12, 29). In this study, in contrast to other studies (15, 29), we determined bone turnover during the early follicular phase of the menstrual cycle when E2 is at low levels to control for effects of cycle stage on bone metabolism. Significant effects of the menstrual cycle on OC and BSAP have been described in women, showing that osteoblast activity is higher during the luteal phase (30). We have described a reduction in bone formation between AN and controls at a point in the menstrual cycle when levels of bone formation in controls are relatively reduced. Therefore, the degree of reduction in indexes of bone formation might be even greater after the follicular phase.

Hypoestrogenemia has been proposed as a contributing factor to the low bone density in AN, because primary or secondary amenorrhea is a nearly universal feature. In adolescents, lower lumbar bone density is seen in amenorrheic teens compared with those with normal menses, but is not significant when patients are controlled for body weight (31, 32). In fact, gains in bone density may precede the return of menses in patients recovering from AN (4). Estrogen treatment in young women with AN has not caused improvement in bone density in a randomized, prospective trial (33), but has not been investigated in adolescent AN patients. In our study, duration of illness was the most predictive of AP and lateral spine BMD. Our data suggest that nutritional status may be an independent determinant of bone density.

This study demonstrates that in addition to estrogen deficiency, adolescents with AN have markedly diminished levels of total and free T, with free T undetectable in the majority of patients. In contrast, dehydroepiandrosterone sulfate levels were not different between the groups, suggesting a relatively greater deficiency in gonadal than ad-
renal androgen production. These data are in agreement with recent data demonstrating that in pubertal adolescents the ovaries are the major source of serum T, in contrast to prepuberty when the adrenals are the primary source (34). Positive correlations between total and free T and bone formation (OC) have been demonstrated in adult women during the normal menstrual cycle, including the follicular phase (35), although this has not been reported in adolescents. In our group of AN adolescents, OC was determined in part by free T in stepwise regression analysis.

Nutritional factors, particularly body weight and lean body mass, are critical determinants of bone mass in adolescent girls (36, 37). Our data demonstrate markedly reduced fat mass, percent body fat, and lean body mass in AN adolescents even when controlled for height and weight. In healthy normal weight and obese children (5–18 yr), lean body mass had the highest correlation with total bone mineral content (36). Bachrach et al. reported that BMI correlated highly with bone density when adolescents with AN and controls were analyzed together, but in the AN group alone there was no correlation (3). In our study, lean body mass was the only body composition variable that contributed significantly to total and regional (total and lumbar BMC and lateral spine BMD) bone density in healthy adolescent girls. However, determinants of bone mass in AN may differ from those in healthy girls. Similar to the relationship between BMI and BMD described by Bachrach et al. (3), our data show that when the AN and control groups were analyzed together, lean body mass significantly correlated with lumbar BMD (r = 0.49; P = 0.005); however, in the AN group alone this relationship was not present.

An important finding in our study is that dietary calcium and vitamin D deficiency are prevalent in our study population of both AN patients and normal adolescent girls. As has been reported in other studies of adolescent girls (38, 39), a majority of girls were deficient in dietary calcium intake during a time when calcium requirements are maximal (40), even though 23% of all girls studied were receiving dietary supplements. Calcium intake has been shown to correlate with bone density in healthy children and adolescents (11, 41), and calcium supplementation has been shown to increase bone density in studies of healthy adolescents, although the effect may be dependent on pubertal stage (38, 39). Most of our adolescent subjects also had deficient vitamin D intake. Although all of our study subjects had normal serum 25-hydroxyvitamin D and PTH levels, the AN group had higher 25-hydroxyvitamin D levels that correlated with vitamin D intake. Our data indicate that most adolescent girls may not get sufficient amounts of these important nutrients with food intake alone; therefore, routine supplementation in many adolescent girls may be warranted. No correlation between calcium intake and BMD in adolescents with AN was demonstrated in this study, and it is unknown whether BMD could be improved by calcium and vitamin D supplementation in adolescents with AN.

Physical activity, particularly weight-bearing exercise, is known to have positive effects on bone density in children and adolescents (11, 41), although heavy exercise during adolescence may lead to delayed puberty and low bone density (42). In this study, physical activity was significantly higher in the AN group, probably contributing to their low weight and amenorrhea. However, there was no relationship between amount of physical activity and BMD in either group. Some patients may have relatively normal BMD despite low weight due to a previous environmental exposure (i.e. prepubertal exercise) or inherited factors, such as vitamin D receptor or COLA1A polymorphisms, which may be protective (43–46) but were not systematically evaluated in this study.

Adolescents with AN have been reported to have decreased linear growth rate and adult height (47, 48); however, adult height in this population of patients in comparison with genetic target height has not been reported. We found that mature AN adolescents were, on the average, 3.0 cm below their genetic target height compared with healthy adolescents who, on the average, reached their genetic target height. The reason for reduced growth in AN could not be evaluated in this cross-sectional cohort and will require longitudinal study, but reduced circulating IGF-I resulting in low bone formation is one potential mechanism.

This is the first study to describe low bone formation in BA- and CA-matched adolescents with AN, in whom bone formation is highly correlated with serum levels of IGF-I. Low bone formation may result in reduced peak bone accumulation and eventual severe osteopenia, as seen in adult women with AN. Low IGF-I due to undernutrition in addition to gonadal steroid deficiency during the critical adolescent period may be an important factor contributing to the pathogenesis of osteopenia in AN.

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References


