The effects of anorexia nervosa on bone metabolism in female adolescents

Leslie A. Soyka

University of Massachusetts Medical School

Let us know how access to this document benefits you.

Follow this and additional works at: https://escholarship.umassmed.edu/peds_endocrinology

Part of the Endocrinology, Diabetes, and Metabolism Commons, and the Pediatrics Commons

Repository Citation


This material is brought to you by eScholarship@UMassChan. It has been accepted for inclusion in Endocrinology/Diabetes by an authorized administrator of eScholarship@UMassChan. For more information, please contact Lisa.Palmer@umassmed.edu.
The Effects of Anorexia Nervosa on Bone Metabolism in Female Adolescents*

LESLIE A. SOYKA, STEVEN GRINSPoon, LYNEE L. LEVITSKY, DAVID B. HERZoG, AND ANNE KLIBANSKI

Neuroendocrine Unit (L.A.S., S.G., A.K.), Pediatric Endocrine Unit (L.A.S., L.L.L.), and Eating Disorders Unit (D.B.H.), Massachusetts General Hospital and Harvard Medical School, Boston, Massachusetts 02114

ABSTRACT

Osteopenia is a frequent, often persistent, complication of anorexia nervosa (AN) in adolescent girls and occurs during a critical time in bone development. Little is known about bone metabolism in this patient population. Therefore, we measured bone density (BMD) and body composition by dual energy x-ray absorptiometry, nutritional status, bone turnover, calcium, and hormonal status in 19 adolescent girls with AN (mean ± SEM, 16.0 ± 0.4 yr) and 19 age-matched controls. The mean duration of AN was 19 ± 5 months. Spinal (L1–L4) osteopenia was common in AN. Lumbar anteroposterior BMD was more than 1 SD below the mean in 42% of patients, and lateral spine BMD was more than 1 SD below in 63% of patients compared with controls. Lean body mass significantly predicted lumbar bone mineral content (r = 0.75; P < 0.0001) in controls only. In AN, duration of illness was the most significant predictor of spinal BMD (lumbar: r = −0.44; P = 0.06; lateral: r = −0.59; P = 0.008). AN adolescents with mature BA (15 yr and greater) were hypogonadal [estradiol, 16.2 ± 6.6 vs. 1.9 ± 1.3 ng/mL; P < 0.0001] and urinary free cortisol levels did not differ. Leptin levels were 2.1 ± 1.8 vs. 0.75 ± 0.53 ng/mL; P < 0.0001) and correlated with all measures of nutritional status, particularly leptin (r = 0.80, P < 0.0001). Surrogate markers of bone formation, serum osteocalcin (OC) and bone-specific alkaline phosphatase (BSAP), were significantly (P = 0.02) reduced in AN vs. controls (OC, 39.1 ± 6.4 vs. 59.2 ± 5.2 ng/mL; BSAP, 27.9 ± 4.0 vs. 40.6 ± 3.4 U/L). The majority of the variation in bone formation in AN was due to IGF-I levels (OC: r² = 0.72; P = 0.002; BSAP: r² = 0.53; P = 0.01) in stepwise regression analyses. Bone resorption was comparable in patients and controls. These data demonstrate that bone formation is reduced and uncoupled to bone resorption in mature adolescents with AN in association with low bone density. Lean body mass was a significant predictor of BMD in controls, but not AN patients. The major correlate of bone formation in AN was the nutritionally dependent bone trophic factor, IGF-I. Reduced IGF-I during the critical period of bone mineral accumulation may be an important factor in the development of osteopenia in adolescents with AN. (J Clin Endocrinol Metab 84: 4489–4496, 1999)

Received June 11, 1999. Accepted September 17, 1999.

Address all correspondence and requests for reprints to: Dr. Anne Klibanski, Neuroendocrine Unit, Bulfinch 457, Massachusetts General Hospital, Boston, Massachusetts 02114.

* This work was supported in part by NIH Grants R01-DK-52625, M01-RR-01066, R01–3833, the Genentech Foundation for Growth and Development, and the Harvard Eating Disorders Center.

Bone density and bone metabolism change dramatically during adolescence, and the onset of AN during this critical time may interfere with achievement of peak bone mass. In addition, significant changes in body weight and composition, pubertal development, and pubertal hormones, such as estradiol and insulin-like growth factor I (IGF-I), occur in AN and may impact bone metabolism. The effect of AN on bone turnover in adolescent girls is poorly understood. Limited data are available (9, 10), and prior studies in adolescents with AN have been limited by the small numbers of subjects and the lack of information about specific biochemical markers of bone turnover, including simultaneous measurements of formation and resorption. Moreover, previous studies have included subjects of a broad age range from early adolescence to young adulthood. Finally, pubertal and skeletal maturation have not been measured in assessments of bone density, even though bone density correlates more with pubertal stage and bone age (BA) than chronological age (CA) (11, 12).

The purpose of this study was to further characterize bone metabolism and potential mechanisms of osteopenia in adolescents with AN in a cohort of adolescent females in whom bone density, biochemical indices of bone turnover, nutritional status, and pubertal hormones were determined compared with those in BA-matched controls. Our data demonstrate significant osteopenia in adolescent girls with AN. Bone formation indices are significantly reduced in association with IGF-I and other nutritionally dependent factors in this population of patients, suggesting a profound effect of this disease on osteoblast function.

Subjects and Methods

Subjects

We studied 38 Caucasian adolescent girls, 19 with AN, and 19 healthy adolescents. The AN patients were 12.8–18.5 yr of age. All met DSM IV...
criteria for AN, with duration since diagnosis ranging from 2–72 months. Only 1 patient had concomitant bulimia. All patients had either primary (n = 2) or secondary (n = 12; duration, 6–48 months) amenorrhea or were premenarchal (n = 5). Two patients with secondary amenorrhea had taken estrogen previously (for 6–18 months), but not within 3 months of the study. Patients receiving hormonal or other medications known to affect bone metabolism were excluded. A history of traumatic bone fracture during childhood or adolescence was re-

Strathal Strom, 1906. None of the controls had a history of an eating disorder.

Based on published standard charts (13). One control subject, 13.1 yr old, was premenarchal. All postmenarchal adolescents had regular menstral periods. None of the controls had a history of an eating disorder. Control subjects with a medical condition or receiving hormonal or other medications known to affect bone metabolism were excluded. A history of traumatic bone fracture during childhood or adolescence was re-

ported in 42% of control subjects, but none occurred within 6 months of the study.

The study protocol was approved by the Massachusetts General Hospital subcommittee on human subjects, and informed consent was obtained from all subjects and their parents.

Experimental protocol

All subjects were evaluated during an out-patient visit at the General Clinical Research Center at Massachusetts General Hospital after a screening visit to determine eligibility and give instruction on collection of urine samples and completion of 4-day food records. A medical and menstrual history and physical examination were performed by a single investigator. All control subjects were studied during the early follicular phase of the menstrual cycle to control for the effects of cycle stage on biochemical and hormonal measurements. All AN and control subjects had their BA determined by x-ray of the left hand and wrist by a single investigator using the methods of Greulich and Pyle (14).

Subjects’ height was measured using a single stadiometer (average of triplicate measurements), and weight was measured in a hospital gown on an electronic scale. BMI was calculated, and BMI percentile was determined from published charts (13).

Subjects collected a 24-h and a 2-h second morning urine sample normalized for creatinine excretion. DPD was measured by competitive enzyme-linked immunosassay (Mitra Biosystems, Mountain View, CA) with a minimum detection limit of 1.1 nmol/L and an intraassay coefficient of variation of 4.3–8.4%. NTX was measured by enzyme-linked immunoassorbent assay (Oxist International, Inc., Seattle, WA) with a detection limit of 20 nmol BCE and an intraassay coefficient of variation of 5–19%.

All samples were run in duplicate using a single kit.

Bone density and body composition

Total body and lumbar spine (L1–L4) bone densities were measured using a QDR-4500 (Hologic, Inc., Waltham, MA). The sn for bone density measurement is 0.01 g/cm² and does not vary with bone density.

Whole body dual energy x-ray absorpmiometry (DEXA) was performed using a Hologic, Inc. QDR-4500 densitometer to determine fat and lean body mass, which has been validated in children and young adults (18, 19).

Nutritional assessment

Nutritional intake data, including calorie, protein, fat, carbohydrate, and micronutrient content, were obtained through computer analysis (version 2, Nutrition Data Systems, Minneapolis, MN) of 4-day food records and history of dietary intake over the month before the study visit. Resting metabolic energy expenditure was performed using a metabolic cart (20). Typical exercise and activity of subjects over the past year were determined using a validated questionnaire (21).

Statistical analysis

Clinical, bone density, body composition, and biochemical data were analyzed with Student’s t test and compared by linear regression analysis. Stepwise regression analyses were performed to determine the relationships of anthropometric body composition, and hormonal data to bone density and bone turnover. Significance was set at P < 0.05 for all comparisons. Results are the mean ± sem unless otherwise indicated.

Results

Clinical and anthropometric data

Clinical and anthropometric data for patients and controls are summarized in Table 1. Patients were matched for BA (15.7 vs. 15.5 yr) with control subjects. CA was not signifi-

Results

Clinical and anthropometric data

Clinical and anthropometric data for patients and controls are summarized in Table 1. Patients were matched for BA (15.7 vs. 15.5 yr) with control subjects. CA was not signifi-

Biochemical assessment

Serum ionized calcium, phosphorus, TSH, dehydroepiandrosterone sulfate, and 24-hour urinary free cortisol (UCP), calcium, and creatinine were measured through the hospital laboratory using previously described methods (17).

Serum estradiol (E₂) was measured by ultrasensitive RIA (Diagnosti-
lower in the AN subjects compared with controls (AN, 0.98 ± 0.01; control, 1.03 ± 0.01; \( P = 0.01 \)). Duration of AN was 19 ± 5 months, and duration of amenorrhea was 24 ± 4 months. Height was not significantly different between the groups. AN patients were of significantly lower weight and BMI. In the AN group, fat mass measured by DEXA was less than 50% that in controls (AN, 0.87 ± 0.3; control, 1.5 ± 0.3; \( P < 0.0001 \)), and percent body fat was substantially reduced (AN, 1314 ± 60 KCal/day). No subjects regularly smoked cigarettes or consumed alcohol.

Historical assessment of activity level over the past year demonstrated that AN subjects were significantly more physically active than controls, with approximately twice as many hours of physical activity per week (AN, 17.0 ± 2.2; control, 9.4 ± 2.2 h/week; \( P = 0.02 \)). Resting metabolic energy expenditure was significantly lower in AN subjects (AN, 1314 ± 60; control, 1587 ± 60 KCal/day). No subjects regularly smoked cigarettes or consumed alcohol.

### Nutritional data

Analysis of intake by 4-day food record and diet history produced similar results; therefore, only the data for food records are provided in Table 2. Patients were in various stages of illness, and there was considerable variability in intake reported by the two groups. The major difference in dietary intake between the groups was reduced fat intake (AN, 21 ± 4; control, 51 ± 2; \( P = 0.04 \)) and percentage of calories from fat (AN, 20 ± 2%; control, 28 ± 2%; \( P = 0.005 \)) in AN subjects. Fifty-three percent of AN patients and 67% of controls had a calcium intake less than the Recommended Daily Allowance despite calcium supplementation in approximately 23%. In 53% of AN patients and 78% of controls, vitamin D intake was less than the Recommended Daily Allowance despite vitamin D supplementation in approximately 30% of subjects.

### Bone density data

Bone density data are summarized in Table 3. Lumbar spine (L1–L4) BMD was significantly lower in the AN subjects (AN, 0.87 ± 0.02; control, 0.97 ± 0.02 g/cm²; \( P = 0.004 \)). Lumbar bone mineral content (BMC) was also reduced in AN (AN, 45 ± 2; control, 51 ± 2 g; \( P = 0.04 \)), with lumbar area not different between the groups. In 42% of AN subjects lumbar BMD was more than 1 sd below the mean for the healthy, height- and BA-matched controls. Sixteen percent of AN subjects had a lumbar BMD more than 2 sd below the normal mean. Comparison of the subjects’ lumbar spine BMD to the DEXA manufacturer normative data also dem-

---

**TABLE 1.** Clinical and anthropometric data

<table>
<thead>
<tr>
<th>Measure</th>
<th>Anorexia nervosa (n = 19)</th>
<th>Control (n = 19)</th>
<th>( P ) value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chronological age (yr)</td>
<td>16.0 ± 0.4 [12.8–18.5]</td>
<td>15.1 ± 0.4 [13.1–17.8]</td>
<td>NS</td>
</tr>
<tr>
<td>Bone age (yr)</td>
<td>15.7 ± 0.3 [13–18]</td>
<td>15.5 ± 0.3 [13–18]</td>
<td>NS</td>
</tr>
<tr>
<td>Bone age/chronological age</td>
<td>0.98 ± 0.01</td>
<td>1.03 ± 0.01</td>
<td>0.01</td>
</tr>
<tr>
<td>Ht (cm)</td>
<td>162.3 ± 1.4</td>
<td>164.7 ± 1.4</td>
<td>NS</td>
</tr>
<tr>
<td>Wt (kg)</td>
<td>43.4 ± 1.5</td>
<td>58.6 ± 1.5</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>16.5 ± 0.4</td>
<td>21.8 ± 0.4</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Duration of illness (months)</td>
<td>19 ± 5 [2–72]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amenorrhea</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No. premenarchal</td>
<td>5</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>No. primary</td>
<td>2</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>No. secondary</td>
<td>12</td>
<td>0</td>
<td></td>
</tr>
<tr>
<td>Duration (months)</td>
<td>24 ± 4 [6–48]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Activity (h/week)</td>
<td>17.0 ± 2.2</td>
<td>9.4 ± 2.2</td>
<td>0.02</td>
</tr>
<tr>
<td>MEE (kCal/day)</td>
<td>1314 ± 60</td>
<td>1587 ± 60</td>
<td>0.003</td>
</tr>
</tbody>
</table>

All values are expressed as the mean ± SEM. Ranges are in brackets. The \( P \) value is for comparison between anorexia nervosa and control subjects by Student’s \( t \) test. MEE, Resting metabolic energy expenditure.

**TABLE 2.** Daily nutrient intake

<table>
<thead>
<tr>
<th>Nutrient</th>
<th>Anorexia nervosa (n = 17)</th>
<th>Control (n = 18)</th>
<th>( P ) value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Calories (KCal)</td>
<td>1730 ± 162 [668–2913]</td>
<td>2042 ± 162 [1395–4093]</td>
<td>NS</td>
</tr>
<tr>
<td>Fat (g)</td>
<td>41 ± 7 [7–88]</td>
<td>65 ± 7 [26–162]</td>
<td>0.03</td>
</tr>
<tr>
<td>% Cal from fat</td>
<td>20 ± 2 [16–33]</td>
<td>28 ± 2 [16–41]</td>
<td>0.005</td>
</tr>
<tr>
<td>Protein (g)</td>
<td>72 ± 6 [31–128]</td>
<td>70 ± 6 [40–122]</td>
<td>NS</td>
</tr>
<tr>
<td>Calcium (mg)</td>
<td>1297 ± 131 [580–2333]</td>
<td>1238 ± 128 [589–2708]</td>
<td>NS</td>
</tr>
<tr>
<td>% &lt; RDA</td>
<td>53%</td>
<td>67%</td>
<td>NS</td>
</tr>
<tr>
<td>Vitamin D (IU)</td>
<td>423 ± 62 [112–1212]</td>
<td>302 ± 60 [80–714]</td>
<td>NS</td>
</tr>
<tr>
<td>% &lt; RDA</td>
<td>53%</td>
<td>78%</td>
<td>NS</td>
</tr>
<tr>
<td>Phosphorus (mg)</td>
<td>1396 ± 131 [570–2476]</td>
<td>1383 ± 127 [741–2551]</td>
<td>NS</td>
</tr>
<tr>
<td>Magnesium (mg)</td>
<td>368 ± 34 [149–606]</td>
<td>310 ± 33 [173–586]</td>
<td>NS</td>
</tr>
<tr>
<td>Dietary supplementation</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Calcium</td>
<td>24%</td>
<td>22%</td>
<td>NS</td>
</tr>
<tr>
<td>Vitamin D</td>
<td>35%</td>
<td>28%</td>
<td>NS</td>
</tr>
</tbody>
</table>

Values are expressed as the mean ± SEM. Ranges are in brackets. The \( P \) value is for comparison between anorexia nervosa and control subjects by Student’s \( t \) test.
onstrated that the AN group had significantly lower lumbar BMD (AN z-score, \(-1 \pm 0.3\); controls z-score, \(0.2 \pm 0.3\); \(P = 0.002\)), with a z-score below \(-1\) in 50\%, and below \(-2\) in 21\% of AN patients. The reduction in lateral spine BMD was even more severe in AN (AN, \(0.68 \pm 0.02\); control, \(0.81 \pm 0.02\) g/cm\(^2\); \(P < 0.0001\)), with BMD more than 1 sd below the normal mean in 63\% of patients and more than 2 sd below the normal mean in 26\% of patients. Total body BMC (TBMC) was reduced in AN (AN, \(1700 \pm 53\) g; control, \(1889 \pm 53\) g; \(P = 0.02\)) and total body BMD (TBMD) was lower in AN, but not significantly. Differences in spinal BMD and TBMC remained significant when controlled for height. Similar differences in bone mineral apparent density were demonstrated (AN, \(0.122 \pm 0.003\); control, \(0.132 \pm 0.003\) g/cm\(^2\); \(P = 0.02\)), using the formula of Katzman et al. (22).

In the control group, lean body mass was the single most significant predictor of lumbar BMC (\(r = 0.75\); \(P < 0.0001\)) and lateral spine (\(r = 0.55\); \(P = 0.003\)) and total (\(r = 0.54\); \(P = 0.004\)) BMD. These correlations remained significant when controlled for height, but weight rather than lean body mass was a more significant determinant of lumbar BMD when controlled for height. The relationships between body weight, composition, BMC, and BMD were not present in AN. In AN, duration of illness was the most important determinant of lumbar BMD (\(r = -0.44\); \(P = 0.06\)) and BMC (\(r = -0.42\); \(P = 0.07\)) and lateral spine BMD (\(r = -0.59\); \(P = 0.008\)), but not TBMD or TBMC. In addition, duration of amenorrhea (\(r = -0.58\); \(P < 0.05\)) was predictive of lateral spine BMD.

**Biochemical data**

TSH and serum ionized calcium and phosphorus were normal in all subjects. Calcium excretion in a 24-h urine collection was normal in all subjects, with the exception of one AN subject who was ingesting 2100 mg calcium daily with a dietary supplement. The median 24-h urinary calcium was 144 mg in both groups (range in AN, 71–450 mg; in control, 39–217 mg). UFC in a 24-h collection was not significantly different between groups (AN, 59 ± 7; control, 51 ± 7 μg), although 33% of AN subjects (6 of 18) had modest elevations in UFC.

Serum PTH levels were normal in all subjects and were not different between the groups (Table 4). Serum 25-hydroxyvitamin D levels were lower in the control group, but normal in all subjects, and correlated with dietary vitamin D intake (\(r = 0.41\); \(P = 0.03\)).

**TABLE 3. Bone density and body composition data**

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa ((n = 19))</th>
<th>Control ((n = 19))</th>
<th>(P) value</th>
<th>(P) value for het-adjusted BMD</th>
</tr>
</thead>
<tbody>
<tr>
<td>LBMD, L1–4 (g/cm(^2))</td>
<td>0.87 ± 0.02</td>
<td>0.97 ± 0.02</td>
<td>0.004</td>
<td>0.01</td>
</tr>
<tr>
<td>LBMC, L1–4 (g)</td>
<td>45 ± 2</td>
<td>51 ± 2</td>
<td>0.04</td>
<td>0.03</td>
</tr>
<tr>
<td>Lateral spine BMD (g/cm(^2))</td>
<td>0.68 ± 0.02</td>
<td>0.81 ± 0.02</td>
<td>&lt;0.0001</td>
<td>0.0002</td>
</tr>
<tr>
<td>Total BMD (g/cm(^2))</td>
<td>1.01 ± 0.02</td>
<td>1.04 ± 0.02</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>TBMC (g)</td>
<td>1700 ± 53</td>
<td>1889 ± 53</td>
<td>0.02</td>
<td>0.02</td>
</tr>
<tr>
<td>Fat (kg)</td>
<td>7.7 ± 0.8</td>
<td>19.6 ± 0.8</td>
<td>&lt;0.0001</td>
<td>NA</td>
</tr>
<tr>
<td>% Body fat</td>
<td>17.7 ± 1.3</td>
<td>33.5 ± 1.3</td>
<td>&lt;0.0001</td>
<td>NA</td>
</tr>
<tr>
<td>Lean (kg)</td>
<td>33.1 ± 1.0</td>
<td>37.1 ± 1.0</td>
<td>0.01</td>
<td>NA</td>
</tr>
</tbody>
</table>

All values are expressed as the mean ± SEM. The \(P\) value is for comparison between anorexia nervosa and control subjects by Student’s \(t\) test. NA, Not applicable.

**TABLE 4. Serum hormone concentrations**

<table>
<thead>
<tr>
<th></th>
<th>Anorexia nervosa ((n = 11))</th>
<th>Control ((n = 15))</th>
<th>(P) value</th>
</tr>
</thead>
<tbody>
<tr>
<td>25-Hydroxyvitamin D (ng/mL)</td>
<td>37 ± 3</td>
<td>29 ± 3</td>
<td>0.05</td>
</tr>
<tr>
<td>PTH (pg/mL)</td>
<td>24.8 ± 3.6</td>
<td>32.8 ± 3.1</td>
<td>NS</td>
</tr>
<tr>
<td>Estradiol (pg/mL)</td>
<td>16.2 ± 1.9</td>
<td>23.3 ± 1.6</td>
<td>0.01</td>
</tr>
<tr>
<td>Total testosterone (ng/dL)</td>
<td>23.6 ± 3.4</td>
<td>25.2 ± 2.9</td>
<td>0.06</td>
</tr>
<tr>
<td>Free testosterone (pg/mL)</td>
<td>0.70 ± 0.17</td>
<td>1.36 ± 0.14</td>
<td>0.01</td>
</tr>
<tr>
<td>DHEAS (μg/dL)</td>
<td>223 ± 31</td>
<td>212 ± 26</td>
<td>NS</td>
</tr>
<tr>
<td>IGF-I (ng/mL)</td>
<td>219 ± 41</td>
<td>511 ± 35</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>Leptin (ng/mL)</td>
<td>2.9 ± 2.1</td>
<td>16.5 ± 1.8</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

Serum hormone data in mature (bone age of 15 yr or greater) subjects. Values are expressed as the mean ± SEM. The \(P\) value if for comparison between anorexia nervosa and control subjects by Student’s \(t\) test.

Serum \(E_2\) levels were significantly reduced, even in comparison with controls sampled in the early follicular phase (Table 4). There was a trend toward lower total T levels in AN, and free T levels were markedly reduced (AN, 0.70 ± 0.17; control, 1.36 ± 0.14 pg/mL; \(P = 0.006\)), with levels below the lower limit of detection of the assay in 82\% of AN subjects.

Serum IGF-I levels were reduced significantly in AN, to less than 50\% of those in the control subjects (AN, 219 ± 41; control, 511 ± 35 ng/mL; \(P < 0.0001\)). Leptin was also markedly reduced in AN (AN, 2.9 ± 2.1; control, 16.5 ± 1.8 ng/mL; \(P < 0.0001\)), to less than 20\% of control levels. In all subjects, IGF-I levels correlated highly with nutritional status, including BMI (\(r = 0.73\); \(P < 0.0001\)), percent body fat (\(r = 0.61\); \(P = 0.0008\)), lean body mass (\(r = 0.49\); \(P = 0.005\)), and leptin (\(r = 0.80\); \(P < 0.0001\)). In contrast, there was no relationship between IGF-I and gonadal steroid concentrations. In stepwise regression analysis including all anthropometric and biochemical measurements, leptin accounted for 65\% of the variation in IGF-I levels (\(r^2 = 0.65\); \(P < 0.00001\)). Leptin was highly dependent on fat mass (\(r = 0.78\); \(P < 0.0001\)).

Bone turnover marker results are summarized in Table 5 and Fig. 1. OC and BSAP were significantly lower in AN patients than in normal subjects. DPD and NTX were not significantly different between the groups. However, there was a trend toward lower NTX levels in the AN group. There was a strong correlation between IGF-I and OC in AN subjects (\(r = 0.71\); \(P = 0.01\); Fig. 2), whereas in the control group a similar trend was found, but the correlation did not reach statistical significance (\(r = 0.46\); \(P = 0.07\)). In stepwise re-
TABLE 5. Bone turnover markers

<table>
<thead>
<tr>
<th>Anorexia nervosa (n = 11)</th>
<th>Control (n = 15)</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>OC (ng/mL)</td>
<td>39.1 ± 6.4</td>
<td>59.2 ± 5.2</td>
</tr>
<tr>
<td>BSAP (U/L)</td>
<td>27.9 ± 4.0</td>
<td>40.6 ± 3.4</td>
</tr>
<tr>
<td>PICP (ng/mL)</td>
<td>139 ± 12</td>
<td>154 ± 10</td>
</tr>
<tr>
<td>DPD (nmol/mmol creatinine)</td>
<td>13.4 ± 1.4</td>
<td>15.4 ± 1.2</td>
</tr>
<tr>
<td>NTX (nmol BCE/mmol creatinine)</td>
<td>138 ± 25</td>
<td>200 ± 21</td>
</tr>
</tbody>
</table>

Bone turnover markers in mature (bone age of 15 yr or more) subjects. Values are expressed as the mean ± SEM. The P value if for comparison between anorexia nervosa and control subjects by Student's t test.

Discussion

In this study, we have characterized bone density and bone metabolism in adolescent girls with AN compared with those in BA-matched controls. Bone formation indices and bone density are significantly reduced in adolescents with AN. Moreover, low bone formation in this population was strongly associated with markedly diminished IGF-I, a nutritionally dependent endogenous bone trophic factor. In addition, we have shown that in these osteopenic girls the decrease in bone formation is uncoupled from bone resorption.

Our data demonstrate that spinal osteopenia is common in adolescent girls with AN compared with a group of BA- and CA-matched healthy adolescents. In the patient group, 42% of AN subjects had a lumbar spine BMD more than 1 sd, and 16% had a lumbar spinal BMD more than 2 sd, below the mean of the BA- and height-matched controls despite an average duration of illness of only 1.5 yr. The results of lateral spine BMD, which may be more representative of trabecular bone (23), demonstrated even more severe osteopenia, with BMD more than 1 sd below the mean in 63% and more than 2 sd in 26% of AN patients compared with controls. Furthermore, we obtained similar results for height-adjusted and bone mineral apparent density calculations, suggesting that the low BMD was not an artifact of size.

Previous studies of bone metabolism in adolescents with AN have been limited by the small numbers of subjects, inclusion of a broad age range, and lack of information about specific biochemical markers of bone turnover, including measures of both formation and resorption. To assess further the mechanisms of reduced BMD in AN we characterized bone metabolism in adolescents with AN compared with that in a CA- and BA-matched group of healthy adolescents. Studies of adult women with AN demonstrate a severe imbalance in bone turnover, with a decrease in markers of bone formation combined with an increase in markers of bone resorption (24). However, data obtained from women with AN cannot be extrapolated to adolescent patients who are in an active period of bone growth and mineral accretion as well as remodeling. Abrams et al. demonstrated that adolescents with AN have decreased calcium absorption and increased urinary calcium excretion; however, bone markers were not measured (9). Saggese et al. reported low levels of carboxy-terminal propeptide of type 1 procollagen, a bone formation marker, in six adolescent AN patients ranging from 11–21 yr of age, but did not investigate other markers of formation or markers of resorption in these patients (10).

Matching patients with controls based on stage of maturation is critical in the evaluation of bone metabolism, as both bone formation and resorption markers change dramatically with the transition from early (Tanner stage 2–3) to late (Tanner stage 4–5) puberty (15, 16). Furthermore, levels of bone formation and resorption remain above adult levels even in mature (Tanner stage 5) adolescents (15). We chose to classify maturation in adolescents based on BA because assessment of Tanner stage in adolescents with AN who are hypoestrogenic is difficult and likely to be inaccurate. In our study, all healthy controls who had a BA of 15 yr or greater had Tanner stage 4–5 development and therefore were classified as mature. In contrast, 35% of AN patients with a mature BA had what appeared clinically to be Tanner stage 3 breast tissue. Therefore, Tanner staging as a measure of pubertal maturation in AN can be inaccurate, whereas BA provides a more objective classification.

Bone formation indexes, OC and BSAP, were significantly reduced in mature adolescents with AN in contrast to those in healthy mature adolescents. In contrast, markers of bone resorption, DPD and NTX, were not significantly different, although there was a trend toward lower NTX in AN subjects. Therefore, in healthy adolescents, high levels of bone formation and resorption were seen. In contrast, patients with AN had low bone formation with normal resorption. Although biochemical markers are indirect measures of bone growth and remodeling, resorption indexes are generally thought to reflect degradation of existing bone, whereas formation indexes in adolescents are thought to reflect skeletal growth and mineralization. We speculate that in a low metabolic state such as AN, remodeling continues, and bone formation associated with growth and bone mineral accumulation is reduced. We were unable to measure bone metabolism in the immature adolescents due to insufficient numbers of subjects and therefore could not determine whether the balance of bone formation and resorption is also altered in immature patients with AN. Further studies in younger patients will provide insight as to whether reduced
Bone formation in AN is dependent on the stage of maturation.

The primary correlate of bone formation in AN was the level of IGF-I, which was markedly reduced in AN subjects, in contrast to the high levels expected and found in normal adolescents. IGF-I functions as a bone trophic hormone, which positively affects bone growth and bone turnover by stimulating osteoblasts, collagen synthesis, and longitudinal bone growth (25, 26). Low levels of serum IGF-I are present in undernutrition and correlate with low BMI in AN (24, 27, 28). We found a high correlation between all indexes of nutritional status, including body composition variables and particularly leptin, with IGF-I levels, indicing that undernutrition is a major cause of reduced IGF-I levels in adolescents with AN. These data indicate that IGF-I deficiency due to undernutrition could significantly contribute to low bone formation in adolescents with AN.

In healthy adolescents, E2 is thought to contribute significantly to bone mineral acquisition, either by acting directly on bone or by stimulating other mediators of bone growth (12, 29). In this study, in contrast to other studies (15, 29), we determined bone turnover during the early follicular phase of the menstrual cycle when E2 is at low levels to control for effects of cycle stage on bone metabolism. Significant effects of the menstrual cycle on OC and BSAP have been described in women, showing that osteoblast activity is higher during the luteal phase (30). We have described a reduction in bone formation between AN and controls at a point in the menstrual cycle when levels of bone formation in controls are relatively reduced. Therefore, the degree of reduction in indexes of bone formation might be even greater after the follicular phase.

Hypoestrogenemia has been proposed as a contributing factor to the low bone density in AN, because primary or secondary amenorrhea is a nearly universal feature. In adolescents, lower lumbar bone density is seen in amenorrheic teens compared with those with normal menses, but is not significant when patients are controlled for body weight (31, 32). In fact, gains in bone density may precede the return of menses in patients recovering from AN (4). Estrogen treatment in young women with AN has not caused improvement in bone density in a randomized, prospective trial (33), but has not been investigated in adolescent AN patients. In our study, duration of illness was the most predictive of AP and lateral spine BMD. Our data suggest that nutritional status may be an independent determinant of bone density.

This study demonstrates that in addition to estrogen deficiency, adolescents with AN have markedly diminished levels of total and free T, with free T undetectable in the majority of patients. In contrast, dehydroepiandrosterone sulfate levels were not different between the groups, suggesting a relatively greater deficiency in gonadal than ad-
renal androgen production. These data are in agreement with recent data demonstrating that in pubertal adolescents the ovaries are the major source of serum T, in contrast to prepuberty when the adrenals are the primary source (34). Positive correlations between total and free T and bone formation (OC) have been demonstrated in adult women during the normal menstrual cycle, including the follicular phase (35), although this has not been reported in adolescents. In our group of AN adolescents, OC was determined in part by free T in stepwise regression analysis.

Nutritional factors, particularly body weight and lean body mass, are critical determinants of bone mass in adolescent girls (36, 37). Our data demonstrate markedly reduced fat mass, percent body fat, and lean body mass in AN adolescents even when controlled for height and weight. In healthy normal weight and obese children (5–18 yr), lean body mass had the highest correlation with total bone mineral content (36). Bachrach et al. reported that BMI correlated highly with bone density when adolescents with AN and controls were analyzed together, but in the AN group alone there was no correlation (3). In our study, lean body mass was the only body composition variable that contributed significantly to total and regional (total and lumbar BMC and lateral spine BMD) bone density in healthy adolescent girls. However, determinants of bone mass in AN may differ from those in healthy girls. Similar to the relationship between BMI and BMD described by Bachrach et al. (3), our data show that when the AN and control groups were analyzed together, lean body mass significantly correlated with lumbar BMD (r = 0.49; P = 0.005); however, in the AN group alone this relationship was not present.

An important finding in our study is that dietary calcium and vitamin D deficiency are prevalent in our study population of both AN patients and normal adolescent girls. As has been reported in other studies of adolescent girls (38, 39), a majority of girls were deficient in dietary calcium intake during a time when calcium requirements are maximal (40), even though 23% of all girls studied were receiving dietary supplements. Calcium intake has been shown to correlate with bone density in healthy children and adolescents (11, 41), and calcium supplementation has been shown to increase bone density in studies of healthy adolescents, although the effect may be dependent on pubertal stage (38, 39). Most of our adolescent subjects also had deficient vitamin D intake. Although all of our study subjects had normal serum 25-hydroxyvitamin D and PTH levels, the AN group had higher 25-hydroxyvitamin D levels that correlated with vitamin D intake. Our data indicate that most adolescent girls may not get sufficient amounts of these important nutrients with food intake alone; therefore, routine supplementation in many adolescent girls may be warranted. No correlation between calcium intake and BMD in adolescents with AN was demonstrated in this study, and it is unknown whether BMD could be improved by calcium and vitamin D supplementation in adolescents with AN.

Other factors that may affect BMD, including UFC and exercise, were investigated, but did not impact BMD in this study. We did not find a relationship between UFC and lumbar BMD as has been described in adult women with AN (6), although 33% of the AN patients had elevations in UFC. Physical activity, particularly weight-bearing exercise, is known to have positive effects on bone density in children and adolescents (11, 41), although heavy exercise during adolescence may lead to delayed puberty and low bone density (42). In this study, physical activity was significantly higher in the AN group, probably contributing to their low weight and amenorrhea. However, there was no relationship between amount of physical activity and BMD in either group. Some patients may have relatively normal BMD despite low weight due to a previous environmental exposure (i.e., prepubertal exercise) or inherited factors, such as vitamin D receptor or COLA1A polymorphisms, which may be protective (43–46) but were not systematically evaluated in this study.

Adolescents with AN have been reported to have decreased linear growth rate and adult height (47, 48); however, adult height in this population of patients in comparison with genetic target height has not been reported. We found that mature AN adolescents were, on the average, 3.0 cm below their genetic target height compared with healthy adolescents who, on the average, reached their genetic target height. The reason for reduced growth in AN could not be evaluated in this cross-sectional cohort and will require longitudinal study, but reduced circulating IGF-I resulting in low bone formation is one potential mechanism.

This is the first study to describe low bone formation in BA- and CA-matched adolescents with AN, in whom bone formation is highly correlated with serum levels of IGF-I. Low bone formation may result in reduced peak bone accumulation and eventual severe osteopenia, as seen in adult women with AN. Low IGF-I due to undernutrition in addition to gonadal steroid deficiency during the critical adolescent period may be an important factor contributing to the pathogenesis of osteopenia in AN.

Acknowledgments

The investigators wish to thank the skilled research nurses on the General Clinical Research Center for their dedicated patient care, Ellen Anderson and the Clinical Research Center nutrition staff for assessment of dietary intake, and Mr. Gregory Neubauer for technical assistance.

References


