Characterization of ST14A Cells for Studying Modulation of Voltage-Gated Calcium Channels

Mandy L. Roberts-Crowley
University of Massachusetts Medical School

Ann R. Rittenhouse
University of Massachusetts Medical School, Ann.Rittenhouse@umassmed.edu

Follow this and additional works at: http://escholarship.umassmed.edu/oapubs

Part of the Cellular and Molecular Physiology Commons, and the Molecular and Cellular Neuroscience Commons

This work is licensed under a Creative Commons Attribution 4.0 License.

Repository Citation
http://escholarship.umassmed.edu/oapubs/2549

This material is brought to you by eScholarship@UMMS. It has been accepted for inclusion in Open Access Articles by an authorized administrator of eScholarship@UMMS. For more information, please contact Lisa.Palmer@umassmed.edu.
Characterization of ST14A Cells for Studying Modulation of Voltage-Gated Calcium Channels

Mandy L. Roberts-Crowley, Ann R. Rittenhouse

Department of Physiology, Program in Neuroscience, University of Massachusetts Medical School, Worcester, Massachusetts, United States of America

¤ Current address: Department of Microbiology and Physiological Systems, Program in Neuroscience, University of Massachusetts Medical School, Worcester, MA, United States of America

* Ann.Rittenhouse@umassmed.edu

Abstract

In medium spiny neurons (MSNs) of the striatum, dopamine D2 receptors (D2Rs) specifically inhibit the Ca\textsubscript{a1.3} subtype of L-type Ca\textsuperscript{2+} channels (LTCs). MSNs are heterogeneous in their expression of dopamine receptors making the study of D2R pathways difficult in primary neurons. Here, we employed the ST14A cell line, derived from embryonic striatum and characterized to have properties of MSNs, to study LTC current and its modulation by neurotransmitters. Round, undifferentiated ST14A cells exhibited little to no endogenous Ca\textsuperscript{2+} current while differentiated ST14A cells expressed endogenous Ca\textsuperscript{2+} current. Transfection with LTC subunits produced functional Ca\textsubscript{a1.3} current from round cells, providing a homogeneous model system compared to native MSNs for studying D2R pathways. However, neither endogenous nor recombinant Ca\textsubscript{a1.3} current was modulated by the D2R agonist quinpirole. We confirmed D2R expression in ST14A cells and also detected D1Rs, D4Rs, Gq, calcineurin and phospholipase A2 using RT-PCR and/or Western blot analysis. Phospholipase C\textsubscript{β-1} (PLC\textsubscript{β-1}) expression was not detected by Western blot analysis which may account for the lack of LTC modulation by D2Rs. These findings raise caution about the assumption that the presence of G-protein coupled receptors in cell lines indicates the presence of complete signaling cascades. However, exogenous arachidonic acid inhibited recombinant Ca\textsubscript{a1.3} current indicating that channels expressed in ST14A cells are capable of modulation since they respond to a known signaling molecule downstream of D2Rs. Thus, ST14A cells provide a MSN-like cell line for studying channel modulation and signaling pathways that do not involve activation of PLC\textsubscript{β-1}.

Introduction

Two classes of L-type Ca\textsuperscript{2+} channel (LTC) α\textsubscript{1} subunits are expressed in the brain: α\textsubscript{1C} (Ca\textsubscript{V1.2}) and α\textsubscript{1D} (Ca\textsubscript{V1.3}) [1] with highest expression in cerebral cortex and striatum [2]. While differing in biophysical properties and pharmacological sensitivities, both LTCs contribute to
membrane excitability, synaptic regulation and gene transcription [3]. In turn, neurotransmitters act via G-protein coupled receptors (GPCRs) to modulate membrane excitability and alter transfer of information within neural circuits. Modulation of L-type 
Ca2+ channels (LTCs) by dopamine GPCR signaling pathways is important in medium spiny neurons (MSN) of the striatum since these neurons are the only source of output from the striatum [4] and are adversely affected in both Parkinson’s and Huntington’s Diseases [5, 6].

Two families of dopamine receptors exist. The D1-like receptor family (D1R, D5R), couples to the G protein Gs, enhancing L-current [7, 8] and the firing rate of MSNs [7]. Conversely, the D2-like receptor family (D2R, D3R, D4R) couples to Gi/o [9], inhibiting L-current [10] and the firing rate of MSNs [11]. Two heterogeneous groups of MSNs respond to dopaminergic input: D1R-expressing MSNs and D2R-expressing MSNs, which are associated with the direct and indirect output, respectively [6]. The balance of output pathways between the opposing D1R- and D2R-expressing MSNs coordinates motor control [12]. Consequently drugs developed to treat Parkinson’s disease target dopamine receptors, particularly D2Rs [13] and more recently LTCs [14, 15]. MSNs express both CaV1.2 and CaV1.3, but D2R activation inhibits only CaV1.3 [11]. In Parkinson’s disease models, loss of D2R modulation of CaV1.3 leads to loss of dendritic spines [16]. Therefore, the pathway underlying D2R modulation of LTC current appears critical for normal function; however due to dopamine receptor heterogeneity in MSNs, the molecular relationship between D2Rs and LTCs has been difficult to elucidate.

Moreover, two different mechanisms may mediate D2R inhibition of LTC current. One characterized pathway involves Gq, phospholipase C (PLC), inositol triphosphate (IP3)-induced Ca2+ release, and protein phosphatase 2B (PP2B) also known as calcineurin [10]. Additionally, D2R activation releases arachidonic acid (AA) in vivo [17–20], in primary neurons [21] and in transfected cell lines [22]. Our laboratory has demonstrated that exogenously applied AA inhibits LTC currents in superior cervical ganglion neurons (SCG) [23–25]. These currents are most likely exclusively due to CaV1.3 current [26]. Additionally, we have shown that AA inhibits recombinant CaV1.3 currents when expressed in HEK293 cells [27]. Therefore, a second D2R signaling pathway inhibiting CaV1.3 may involve activation of Ca2+-dependent cytosolic phospholipase A2 (cPLA2), which cleaves AA from phospholipids, similar to M1 muscarinic receptor (M1R) modulation of LTC current in SCG [25].

In the present study, we developed a model system to probe the D2R signaling pathway inhibiting CaV1.3 using the ST14A cell line, created from embryonic rat striatum [28]. Retroviral transduction of the temperature-sensitive SV40 large T antigen enables ST14A cells to grow and divide at the permissive temperature of 33°C. At higher temperatures the cells differentiate to exhibit general neuronal, as well as specific MSN-like, properties including functional D2-like receptors [28, 29]. We examined whether ST14A cells express identified signaling molecules downstream of D2Rs in an effort to develop a model cell line with MSN properties to study the functional effects of D2R signaling on CaV1.3. We found that round ST14A cells lacked endogenous Ca2+ current and exploited this deficiency by transfecting cells with CaV1.3 and accessory subunits, thus eliminating the need for pharmacological blockers. Recombinant CaV1.3 currents were characterized and tested for current modulation by the D2 dopamine agonist quinpirole and by exogenous AA.

Materials and Methods
Preparation of Cells

The ST14A cell line, created by Elena Cattaneo and described in Cattaneo and Conti, 1998 [28], was given to us as a gift from Dr. Michelle E. Ehrlich (Jefferson Medical School) with permission from Dr. Elena Cattaneo (University of Milano). ST14A cells were propagated at the
permissive temperature of 33°C in Dulbecco’s modified Eagle’s medium (DMEM; Gibco) supplemented with 0.11 g/L sodium bicarbonate (Sigma, St. Louis, MO), 0.29 g/L L-glutamine, 3.9 g/liter HEPES, 100 units/ml penicillin-streptomycin, and 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA). Cells were transferred to 37°C to promote differentiation and used 1–2 days later. The A9L cell line, derived from A9 L cells co-transfected with the human D2R and obtained from the American Type Culture Collection (ATCC, Manassas, VA), were propagated at 37°C in DMEM with 4 mM L-glutamine adjusted to contain 1.5 g/L sodium bicarbonate and 4.5 g/L glucose containing 10% FBS. HEK 293 cells were propagated at 37°C in DMEM/F12 containing 10% FBS. All cells were maintained in a temperature-controlled humidified incubator at 5% CO2 and passaged once flasks became 80–90% confluent. SCG, striatum and cortex of 1 to 4-day old or adult Sprague-Dawley rats (Charles River Laboratories, Wilmington, MA) were isolated following CO2 exposure and decapitation using a protocol (protocol # 822) approved by the Institutional Animal Care and Use Committee (IACUC) of University of Massachusetts Medical School. This study was carried out in strict accordance with the recommendations in the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health. All efforts were made to minimize animal suffering. The Institutional Animal Care and Use Committee (IACUC) of University of Massachusetts Medical School specifically approved animal use for this study.

**Electrophysiology**

Whole-cell currents were recorded at room temperature (RT, 20–24°C) with an Axon 200B patch clamp amplifier (Molecular Devices, Sunnyvale, CA). Currents were filtered at 1–5 kHz and digitized at 5 times the filter cut-off frequency of the 4-pole Bessel filter of the amplifier. Electrodes were pulled from borosilicate glass capillary tubes and each electrode was fire-polished to ~1 m to give the pipette a resistance of 2–3 M. The pipette solution consisted of (in mM): 125 Cs-Aspartate, 10 HEPES, 0.1 BAPTA, 5 MgCl2, 4 ATP and 0.4 GTP brought to pH 7.50 with CsOH. High resistance seals were established in Ca2+ Tyrode’s consisting of (in mM): 5 CaCl2, 145 NaCl, 5.4 KCl, and 10 HEPES brought to pH 7.50 with NaOH. Once a seal was established and the membrane ruptured, the Tyrode’s solution was exchanged for an external bath solution consisting of (in mM): 125 NMG-Aspartate, 20 Ba2+, 10 HEPES brought to pH 7.50 with CsOH.

FPL 64176 and nimodipine were prepared as stock solutions in 100% ethanol and stored at -20°C. AA (5,8,11,14-eicosatetraenoic acid; NuCheck Prep, Elysian, MN) and oleic acid (NuCheck Prep) were dissolved in 100% ethanol and stored under nitrogen as stock solutions at -70°C. ω-conotoxin GVIA (Bachem, Torrence, CA) and quinpirole were prepared as stock solutions in double distilled water and stored at -70°C. Oxotremorine-M (Tocris, Ellisville, MO) was prepared fresh daily by making a 10 mM stock in double distilled water. Working dilutions were made fresh daily by diluting stock solutions at least 1:1,000 with external bath solution. For ethanol prepared stocks, the final ethanol concentration was less than 0.1%. Bovine serum albumin (BSA; fraction V, heat shock, fatty acid ultra-free; Roche Applied Science, Indianapolis, IN) was added directly to the bath solution for a final concentration of 1 mg/ml. All chemicals were purchased from Sigma unless otherwise noted.

Data were acquired using Signal 2.14 software (Cambridge Electronic Design, Cambridge, England) and stored for later analysis on a personal computer. Linear leak and capacitive currents were subtracted from all traces. Data are presented as the mean ± s.e.m. Significance was determined statistically using a two-tailed paired or unpaired t-test, or a one-way ANOVA. Analysis programs include Signal (Cambridge Electronic Design), Excel (Microsoft, Redmond, WA) and Origin (OriginLab, Northampton, MA).
Transfection

ST14A cells were transfected by lipofectamine (Invitrogen, Carlsbad, CA) with a 1:1:1 molar ratio of CaV1.3, β2a, and α2δ subunits [30]. Constructs for CaV1.3b (+exon11, Δexon32, +exon42a; GenBank accession #AF370009), and α2δ-1 (GenBank accession #AF286488) were a gift from Dr. Diane Lipscombe (Brown University) and the construct for CaV β2a (GenBank accession #M80545) was a gift from Dr. Edward Perez-Reyes (University of Virginia). For all transfections, 0.4 μg of DNA was used per well of a 12-well plate. Prior to transfection, cells were washed with DMEM. The DNA mixture was then added dropwise to each well, gently swirled then incubated for 1–3 h at 37°C in a 5% CO2 incubator. Supplemented media, without antibiotics, was returned to the wells to bring the volume up to 1 ml (normal growth medium volume). Cells were washed with full media 2 and 4 h later and assayed for transient gene expression after 24–72 h.

Reverse Transcriptase-Polymerase Chain Reaction

Homogenized tissue samples (50–100 mg) or confluent cells in a 100 mm dish were lysed in 1 ml TRIzol Reagent (Invitrogen). RNA was separated from DNA by phenol-chloroform phase separation. RNA was precipitated with isopropyl alcohol and washed with 75% ethanol. The RNA pellet was dried and resuspended in RNase-free water. RNA samples had an A260/A280 ratio between 1.6 and 1.8 and were treated with DNase to eliminate contamination with genomic DNA. For reverse transcription, cDNA was synthesized from the mRNA by adding 1 μl 10X buffer RT, 1 μl dNTP Mix (5 mM each dNTP), 1 μl Oligo-dT primer (0.5 mg/ml, Promega, Madison, WI), 0.125 μl RNase Inhibitor (40 U/μl, Promega), 0.5 μl Omniscript Reverse Transcriptase (4 U/μl) and RNase-free water for a total volume of 10 μl (all reagents from QIAGEN, Valencia, CA, unless otherwise noted). The mixture was incubated at 37°C for 1 h. The mixture was then heated at 93°C for 5 min and then placed on ice to inactivate the transcriptase. PCR amplification was then performed with a Techgene thermal cycler (Techne Inc, Burlington, NJ) with thin walled PCR tubes.

PCR primers for dopamine receptors D1-D5 (D5 formerly referred to as D1b) were sequences previously published [31]. D1: 5′-GAC AAC TGT GAC ACA AGG TTG AGC-3′ and 5′-ATT ACA GTC CTT GGA GAT GGA GCC-3′ yields a 609 base pair (bp) product. D2: 5′-GCA GTC GAG CTT TCA GAG CC-3′ and 5′-TCT GCG GCT CAT CGT CTT AAG-3′ yields 404 and 317 bp products recognizing the long and short forms of the D2R, respectively. D3: 5′-AGC ATC TGC TCC ATC TCC AAC CC-3′ and 5′-AGA GGT CCG AGT CCT TTC CAC G-3′ yields a 461 bp product. D4: 5′-TC ATG CTA CTG CTT TAC TGG GCC A-3′ and 5′-TCTG AGA GAG GTC TGA TGG TC-3′ yields a 223 bp product. D5: 5′-AGT CGT GGC TAT GAA CCT GAC-3′ and 5′-GCG TCG TTG GAG AGA TTT GAG ACA-3′ yields a 517 bp product. GAD65 and GAD67 primers were from sequences previously published [32]. GAD65: 5′-GCC CCC TGT ATT TGT ACT GCC-3′ yields a 400 bp product. GAD67: 5′-CAC ACC AGT TGC TGG GAG-3′ and 5′-ACA AAC ACC GGT GCA ATT-3′ yields 318 and 238 bp products. GAPDH primers were from sequences previously published [33]. GAPDH: 5′-TGC CAA GGA GGG TGG GGA CAA GG-3′ and 5′-GCT TCA CCA CCT TCT TGA TG-3′ yields a 199 bp product and confirmed equal loading.

Reaction mixtures for PCR contained 200 ng of cDNA, 5 μl of 2.5 X Eppendorf MasterMix [Taq DNA Polymerase (62.5 U/ml), 125 mM KCl, 75 mM Tris-HCl, 3.75 mM Mg(OAc)2, 0.25% Igepal-CA630, 500 μM of each dNTP and stabilizers, Brinkmann, Westbury, NY], 1 μl each of forward and reverse primer (5 pmol/μl, Invitrogen for dopamine receptors and GAD; Qiagen for GAPDH), 2 to 4 mM Mg2+ (Brinkmann) and distilled water to a final volume of
12.5 µl. High (4 mM) Mg2+ concentrations were necessary to amplify dopamine receptor mRNAs. The protocol for dopamine receptor amplification was 94°C for 1 min, 58°C for 1 min, and 72°C for 3 min for 20 cycles [31]. For the D3R, an annealing temperature of 62°C was also tried. For GAD amplification, the protocol was 94°C for 3 min, 53°C for 1 min, 52°C for 1 min, 51°C for 1 min, and 70°C for 2 min for 30 cycles [32]. A 2 µl aliquot was used as a template for a second round of amplification for thirty cycles. PCR products and a 100 bp DNA ladder (Invitrogen; bright band 600 bp or Promega, Madison, WI; bright band 500 bp) were separated by electrophoresis in 2% agarose gels stained with ethidium bromide. Bands excised from gels were sequenced by the UMASS Medical School Nucleic Acid Facility and compared to published sequences (GenBank accession number): D1 (M35077), D2 (M36831), D4 (M84009), D5 (M69118), GAD65 (M74826), and GAD67 (M81883).

Western Blot Analysis

The immunoblot protocol has been previously described [34]. All extracts and buffers contained protease inhibitors (Santa Cruz Biotechnology, Santa Cruz, CA): pepstatin A (1 µg/ml), leupeptin (10 µg/ml), aprotinin (20 µg/ml), phenylmethanesulfonyl fluoride (200 nM), and calpain inhibitor I and II (8 µg/ml each). Day 4 rat brain or cells grown in uncoated 60-mm dishes were solubilized in lysis buffer (0.15 M NaCl, 5 mM EDTA, 1% Triton-X 100, 10 mM Tris-Cl pH 7.4), sonicated, and centrifuged to remove insoluble material. Protein concentration was determined by the RC/DC assay (Bio-Rad, Hercules, CA). 20–30 µg of protein was loaded per lane and separated on 8% SDS-polyacrylamide gels. Equal protein loading was confirmed by staining with β-actin antibodies.

Antibodies used include: monoclonal- D2R (1:400; Santa Cruz Biotechnology), PP2B and β-actin (1:500 and 1:10,000; Sigma); polyclonal- M1R and Gqα (1:1,000; Santa Cruz Biotechnology), cPLA2 (1:200; Cell Signaling Technology, Danvers, MA), PLCβ-1 (1:200; Millipore, Billerica, MA); secondary- HRP-conjugated goat anti-mouse or bovine anti-rabbit (1:15,000 or 1:10,000; Santa Cruz Biotechnology). Primary and secondary antibodies were diluted with PBS.

Results

To determine if ST14A cells express endogenous Ca2+ current, we measured whole-cell currents from cells grown at 33 or 37°C. Two populations of cells were distinguished based on morphology: round or differentiated, with the definition of having neuron-like projections. From a holding potential of -90 mV, round cells exhibited zero to little endogenous peak or tail current measured at +10 mV or -40 mV respectively, regardless of the temperature at which the cells were grown (Fig 1A). Differentiated cells had significantly more endogenous current than round cells at both potentials (Fig 1B). Application of the LTC agonist, FPL 64176 (FPL; 1 µM) enhanced endogenous currents at both potentials (Fig 1A and 1B), indicating that at least a component of the endogenous current was LTC. Since round cells showed little to no LTC current, we transfected the ST14A cells with LTC channel subunits, CaV1.3, β2a, and α2δ-1, along with green fluorescent protein (GFP) and recorded whole-cell currents from green fluorescenting round cells (Fig 1C). Peak and tail currents from round transfected cells were significantly larger than those recorded from round untransfected cells (Fig 1A and 1B). FPL significantly enhanced recombinant current at -40 mV. Fig 1D shows representative individual traces of round endogenous or round recombinant current before and after application of FPL. The recombinant current shows little to no inactivation with 20 mM Ba2+ as the charge carrier [35], typical of LTC current coexpressed with the accessory subunit, β2a [36].

Since CaV1.3 activates at more negative voltages compared to CaV1.2 [30], we measured peak current at -10 mV. Recombinant current recorded at -10 mV was larger than at +10 mV.
Fig. 1. ST14A cells have endogenous Ca\textsuperscript{2+} currents but are capable of expressing recombinant current. (A-B) Currents were recorded from round (undifferentiated, no processes) or differentiated (having processes) endogenous ST14A cells. Note: round cells from ST14A cells grown at either 33°C or 37°C exhibited little to no current and were pooled. Currents were recorded from round ST14A cells grown at 33°C, transfected and grown at 37°C for at least 24 hours before recording. Each set of cells were recorded in the absence (white bars) and presence of 1 μM FPL (black bars). Summary of peak Ca\textsuperscript{2+} currents measured from a holding potential of -90 mV to a test potential of +10 mV (A) and then to a tail potential of -40 mV (B).
Endogenous current from differentiated cells was significantly larger than endogenous current from round cells (†; \( p < 0.05 \)); Transfected current was significantly larger than endogenous current from round cells (‡; \( p < 0.05 \)); FPL significantly increased differentiated endogenous and transfected tail current (\(*; \ p < 0.05\), \( n = 7–15 \)). (C) Transfected ST14A cells expressed GFP throughout the cell soma and in a small percentage of differentiated cells, in the processes. Images (20X magnification) were captured ~24 hours post-transfection. The transfection rate for these cells was ~50%. (D) Top: Protocol for eliciting currents from a holding potential of -90 mV to the test potential of +10 mV for 100 ms before repolarizing to -40 ms. Individual traces from round endogenous (middle) or round transfected (bottom) cells. Dashed lines indicate where peak and tail current were measured 65 ms after depolarization and 15 ms after repolarization, respectively. In the presence of LTC agonist, 1 \( \mu \text{M} \) FPL, both the peak and long-lasting tail current increased (gray trace) and displayed slowed activation and deactivation kinetics, characteristic of FPL-induced L-current.

(Fig 1A) and this increase was also reflected in the tail current amplitude at -40 mV (Fig 1B).

The summary of these results shows that round ST14A cells have little endogenous LTC current but are capable of being transfected with LTCs and expressing functional LTC currents. Therefore, the round cells represent a population of ST14A cells that we used to study recombinant LTC function in isolation from other types of native Ca\(^{2+}\) channels.

We characterized recombinant Ca\(_{\text{V}1.3}\) current further to determine whether transfected channel activity in ST14A cells exhibits biophysical and pharmacological properties of Ca\(_{\text{V}1.3}\) observed in oocytes and HEK 293 cells [30]. First, we measured peak current across a range of voltages to show that channels open at relatively negative voltages compared to Ca\(_{\text{V}1.2}\) [30]. Indeed, in 20 mM Ba\(^{2+}\), Ca\(_{\text{V}1.3}\) currents activated at a test potential of -60 mV and peaked at -10 mV to 0 mV (Fig 2A). Recombinant current in ST14A cells exhibited a Ca\(_{\text{V}1.3}\) LTC pharmacological profile. Fig 2B shows a time course of the effects of Ca\(^{2+}\) channel ligands on Ca\(_{\text{V}1.3}\) current. The current was insensitive to the N-type Ca\(^{2+}\) channel antagonist, \( \omega \)-conotoxin GVIA (1 \( \mu \text{M} \); CTX) but was inhibited in a concentration-dependent manner by the LTC antagonist, nimodipine (0.1–3.0 \( \mu \text{M} \); NIM). The inhibition produced by 1.0 \( \mu \text{M} \) NIM (51.8 ± 4%) is characteristic of the low-voltage sensitivity of Ca\(_{\text{V}1.3}\) compared to Ca\(_{\text{V}1.2}\), which would be fully blocked by 1.0 \( \mu \text{M} \) NIM [30]. Inhibition fully reversed by washing with bath solution. After wash out, channels remained sensitive to FPL. The antagonist data are summarized in the bar graph in Fig 2C. These findings show that recombinant current in ST14A cells displayed both current-voltage and pharmacological profiles specific to Ca\(_{\text{V}1.3}\) channels.

To determine whether D\(_{2}\)R activation by the agonist quinpirole (Quin) inhibits Ca\(_{\text{V}1.3}\) currents in ST14A cells, we recorded recombinant currents in the presence of FPL to enhance current amplitude. At a concentration of 10 \( \mu \text{M} \), Quin had no significant effect on peak or tail current amplitude over time, (Fig 3A left) or in individual traces (Fig 3A right). To determine if only recombinant Ca\(_{\text{V}1.3}\) current was insensitive to D\(_{2}\)R activation, we tested whether endogenous ST14A current from differentiated cells could undergo modulation. Again 10 \( \mu \text{M} \) Quin had no significant effect on endogenous peak or long-lasting tail current amplitude. Fig 3B shows representative current traces in the presence of 1 \( \mu \text{M} \) FPL from a range of voltages before and after Quin. Since a majority of MSNs express muscarinic M\(_{1}\) receptors (M\(_{1}\)Rs) as well as dopamine receptors [37], we tested whether activation of this receptor would inhibit endogenous current. The muscarinic agonist oxotremorine-M (Oxo-M; 10 \( \mu \text{M} \)) had no effect on endogenous peak or long-lasting tail current amplitude. Fig 3C shows representative current traces in the presence of FPL from a range of voltages (-60 mV to -10 mV) tested before and after Oxo-M. Fig 3D summarizes the effect of Quin and Oxo-M on peak (left) and long-lasting tail current (right) from recombinant Ca\(_{\text{V}1.3}\) versus endogenous ST14A current. These results suggest that the D\(_{2}\)R and M\(_{1}\)R signaling pathways, which inhibit LTC current in MSNs, are not intact in ST14A cells. However, application of dopamine or Quin, increases CREB phosphorylation.
Fig 2. Whole-cell currents from transfected ST14A cells have biophysical and pharmacological properties of Ca\(\text{V}_{1.3}\) LTCs. (A) The current-voltage (I-V) relationship shows that recombinant Ca\(\text{V}_{1.3}\) current activates at approximately -60 mV; n = 15. (B) Example time course of peak current at -10 mV. 1 μM ω-conotoxin GVIA (CTX, an N-type Ca\(^{2+}\) channel antagonist) was added for 2 min. Bath solution was exchanged with 1 μM nimodipine (NIM, an LTC antagonist) for 2 min or until a new stable baseline was reached and then washed off to show reversibility. FPL (1 μM) was added at the end of the recording. (C) Summary of pharmacological inhibition of transfected current; n = 3–6.

doi:10.1371/journal.pone.0132469.g002
phosphorylation in ST14A cells, indicating that D2Rs do couple to intact signaling cascades such as the adenylyl cyclase and MAPK pathways [29].

A non-selective D2R-like agonist, Quin also activates D3 and D4Rs [38]. Subsequently, D3 and D4Rs modulate immediate early gene expression [39, 40] raising the question of the identity of dopamine receptors in ST14A cells. Using RT-PCR, we examined dopamine receptor (D1, D2, D3, D4, and D5) mRNA content in ST14A cells, striatum (positive control for all dopamine receptors), cortex, A9L cells (D2R positive control; see Materials and Methods) or HEK

Fig 3. ST14A endogenous or recombinant Ca$_v$1.3 current is not modulated by GPCR activation. (A) Time course (left) and representative sweeps (right) of recombinant Ca$_v$1.3 current before (FPL), 1 min following 10 μM quinpirole application (Quin) and after removing Quin (Wash). (B–C) Endogenous, differentiated current traces in the presence of FPL across a range of test potentials (-50 mV to 0 mV) before (left) and after (right) application of (B) 10 μM Quin or (C) 10 μM oxotremorine M (Oxo-M). (D) Summary of percent inhibition of peak (left) and long-lasting tail (right) recombinant Ca$_v$1.3 or endogenous currents by D2R or M1R agonist; n = 3–7.

doi:10.1371/journal.pone.0132469.g003
293 cells (negative control for all dopamine receptors). PCR products were detected for long and short splice variants of the D2R in ST14A cells (n = 4/11 and 2/11, respectively, striatum (n = 1/2 and 2/2, respectively), cortex (n = 1/2 for both) and A9L cells (n = 2/5 and 2/5, respectively) as shown in Fig 4A. Additionally, D1R (n = 2/10), D3R (7/10), D5R (4/10) (data not shown) mRNAs were also detected in ST14A cells, regardless of the temperature at which cells were grown. D1R, D3R and D5R mRNAs were detected in striatum whereas D1R and D2R mRNAs were detected in the cortex (data not shown). D4R mRNA was also detected in A9L cells (n = 3/4). Experiments for D3R mRNA expression in striatum, cortex, and ST14A cell samples resulted in a smear despite several attempts to adjust the protocol. HEK 293 cells showed no expression of any of the dopamine receptors tested. These results show that ST14A cells express mRNA for more than one D2-like receptor; this finding could account for the previously reported D2R-like changes in pCREB [29].

Since MSNs are GABAergic, a defining characteristic of ST14A cells being MSN-like would be expression of glutamic acid decarboxylase (GAD), the enzyme that catalyzes GABA synthesis from glutamate. Using RT-PCR, we measured whether ST14A cells express GAD2 and GAD1, two genes which encode GAD with molecular weights of 65 and 67 kDa, respectively. The primers used against GAD1 detect both embryonic and adult splice forms of GAD67. All three bands corresponding to GAD65 and embryonic and adult GAD67 were detected in ST14A cells (Fig 4B). Alternate lanes in which RNA samples were not reverse tran-scribed served as controls for genomic DNA contamination and yielded no product. Using striatal and cortical tissues as positive controls for the GAD genes, and in A9L cells, we detected all three forms of GAD as well. In contrast, no bands for GAD65 and only faint bands for GAD67 were detected in HEK 293 cells. The presence of GAD in ST14A cells further confirms that this cell line exhibits GABAergic characteristics of MSNs.

After confirming ST14A cells exhibit a similar mRNA expression profile for dopamine receptors and GAD as MSNs, we examined whether D2Rs and downstream signaling molecules were expressed in ST14A cells. Using Western blot analysis, D2R and M1R expression was confirmed in ST14A cells. D2R antibodies recognized a band at 50 kDa, the expected molecular weight of D2Rs (Fig 5A, top panel) for striatum and cortex (positive controls). A second band at 75 kDa, absent in the A9L cell line, most likely represents a glycosylated form of the receptor [41], further supporting the idea that the ST14A cells are similar to neurons in these brain regions regarding post-translational modification of proteins. Fig 5A (middle panel) shows that ST14A cells, but not A9L cells, express M1Rs, displayed as a 50 kDa band on Western blots. This antibody recognized M1R expression in SCG, striatum, and cortex [34]. Expression of Gqα (Fig 5A, bottom panel), which couples to M1Rs, was also detected in the cell lines and tissue samples at the predicted molecular weight of 42 kDa. These results show that ST14A cells express D2R, M1R and Gqα proteins.

Since D2Rs and M1Rs both couple to PLCβ-1, we examined whether ST14A cells express PLCβ-1. Protein expression of PLCβ-1 was not detected in either the A9L or ST14A cell lines, but was detected in SCG tissue (Fig 5B). The unanticipated absence of PLCβ-1 may account for the lack of CaV1.3 modulation since PLCβ-1 is required for LTC current inhibition by both the D2R and M1R pathways [10, 42]. To determine the presence of molecules downstream of PLCβ-1 reported to participate in LTC modulation by these receptors [10, 25], we tested for cPLA2 and PP2B expression. Fig 5C shows that ST14A cells cultured at 33 and 37°C, as well as A9L cells, express cPLA2. cPLA2 was detected at low expression levels from postnatal day 1 SCG (positive control) as has been reported previously [25]. Fig 5D shows that ST14A cells also express PP2B. Since PLCβ-1 is downstream of both D2 and M1Rs, this result may explain our lack of inhibition of CaV1.3 or endogenous LTC current by Quin or Oxo-M in ST14A cells.
Fig 4. ST14A cells express several D₂R family mRNAs. ST14A mRNA transcripts were amplified by RT-PCR. (A) D₂R mRNA expression in ST14A, A9L, HEK 293 cells and in striatum. A9L cells that express D₂Rs and striatum served as positive controls. Left lane contains a 100 bp ladder; brightest band is 600 bp. The band for ST14A cells was sequenced and BLAST search results matched previously published sequences for D₂Rs. No D₂R bands were detected in HEK 293 cells. (B) GAD65/67 mRNA expression in ST14A, striatum, cortex, A9L and HEK 293 cells. As a control, ST14A mRNA was not reverse-transcribed (-RT).

doi:10.1371/journal.pone.0132469.g004
To circumvent the absence of a key signaling molecule, and determine whether CaV1.3 could be modulated, we directly applied exogenous AA (10 μM) to the bath and measured CaV1.3 recombinant current over several minutes. In the presence of FPL, AA inhibited CaV1.3 peak and long-lasting tail currents by 40 ± 12% and 29 ± 25% respectively after 1 min (n = 4). Fig 6A shows representative sweeps before (FPL) and after AA application (FPL + AA). The large variability in tail current inhibition by AA suggested that voltage may be important for this modulation. However, AA inhibited CaV1.3 current at all voltages when tested over a range of test potentials as shown in the I-V plot in Fig 6B. Bovine serum albumin (BSA), which binds free fatty acids [43], reversed inhibition at all test potentials. To show that inhibition was
not due to AA competing with FPL, we measured CaV1.3 current in the absence of FPL prior to and after application of AA (Fig 6C) and still observed inhibition that could be reversed after adding BSA. Inhibition of CaV1.3 over time by AA (open bars) and recovery by BSA (solid bar) is summarized in Fig 6D. Conversely, oleic acid (10 μM) enhanced current by 7.9 ± 0.7% after 1 minute (Fig 6E; p < 0.001; n = 5–7). (E) Time course of peak CaV1.3 current with 10 μM oleic acid (OA); n = 3.

doi:10.1371/journal.pone.0132469.g006

Discussion

MSNs have a resting membrane potential that oscillates between ~ -85 mV (during the “down” state) and ~-60 mV (during the “up” state) [4, 6]. Inhibition of CaV1.3 is of particular interest in MSNs because it activates at potentials approximately 25 mV more negative than CaV1.2, [30]. Since CaV1.3 activates at the low voltage of -60 mV, this channel may open during the “up” state and contribute to reaching threshold for firing an action potential [4]. Moreover,
increased D2R signaling or inhibition of LTCs in MSNs decreases membrane excitability [10]. MSN activity produces the only output from the striatum and thus the finely tuned regulation of MSN activity is critical for normal motor function. Disruption of MSN regulation results in severe malfunction of the basal ganglia, as seen when MSNs lose dopaminergic input in Parkinson’s disease, or when MSNs undergo cell death as seen in Huntington’s disease [5, 44].

Because studying Ca2+ currents in MSNs has been challenging, we searched for a cell line that might serve as a model system for MSNs. However, remarkably few neuronal cell lines have been developed despite ongoing demand for their use in biophysical studies of ion channels and in high through put systems for therapeutic drug testing. We hypothesized that ST14A cells, compared to HEK 293 cells, would express postsynaptic endogenous D2R signaling microdomains closely matching MSNs, thus making ST14A cells useful for studying CaV1.3 modulation by D2R specific signaling. Therefore we examined ST14A cells to determine whether this cell line exhibits sufficient striatal properties to serve as a suitable model system for studying MSN functioning.

We first tested whether ST14A cells express endogenous voltage-gated Ca2+ current. We characterized a small, endogenous Ca2+ current from differentiated ST14A cells that develop neuronal-like processes when grown at 37°C. However, we found that only 5/15 cells had a current amplitude larger than 200 pA even in the presence of the LTC agonist FPL. This small current and the relatively low number of cells expressing a measurable current, is similar to previous findings [29]. K+, Na+ and HCN-mediated currents, in addition to the ability to fire action potentials, are also reported in a small percentage of ST14A cells [29, 45, 46]. However, when present, the Ca2+ current was within the lower range of current amplitudes recorded in MSNs, and was similar to L-current in MSNs in that the tail current was sensitive to LTC agonists, increasing ~ 6-fold following exposure to FPL. L-current dominates few types of neurons; one of them being MSNs (e.g., [10] [47]). Though the Ca2+ current in differentiated ST14A cells is small, L-current appears to make up much of the whole-cell Ca2+ current. Further optimization of culture conditions and/or recording conditions may increase CaV1.3 expression and consequently L-current amplitudes.

As an alternative strategy to examining native CaV1.3 activity in ST14A cells, we attempted to transiently transfect round ST14A cells, which lack endogenous Ca2+ currents, with LTC channel subunits and GFP. We didn’t know whether ST14A cells would tolerate transfection of the multiple Ca2+ channel subunits as well as express functional channels. However ST14A cells were transfected successfully with channel subunits and exhibited robust voltage-gated Ca2+ currents. We characterized the biophysical properties of isolated, recombinant CaV1.3 current in a striatal-like background without the complications of primary MSNs, i.e., requiring several pharmacological blockers to silence other ionic or multiple types of Ca2+ currents. These large currents appeared ideal for testing whether ST14A cells would support CaV1.3 current inhibition by the D2R agonist.

From the biochemical characterization of ST14A cells as MSN-like due to the expression of dopamine receptors (D1, D2, D4, D5) and GAD65/67 mRNAs as well as expression of D2R, M1R, Gqα, cPLA2 and PP2B protein we anticipated observing LTC modulation. However, no inhibition of CaV1.3 current was observed with Quin or Oxo-M. Since PLCβ-1 expression is so widespread, we tested for its expression in ST14A cells only after finding no LTC modulation by Quin. Lack of PLCβ-1 protein expression was unexpected since several intact signaling cascades are described in the growing literature regarding ST14A cells. Most notably, ST14A cells have functional CREB phosphorylation following D2R stimulation and Ras/MAPK, adenyyl cyclase, Wnt and JAK/STAT signaling pathways [29, 48–51]. Moreover these cells express other enzymes that act on lipids including N-acyl-phosphatidylethanolamine-hydrolyzing phospholipase D (NAPE-PLD), fatty acid amide hydrolase, diacylglyceride lipase, monoacylglycerol lipase (Bari et al., 2013), and cPLA2 (Fig 3C).
The absence of PLCβ-1 expression in ST14A cells coincides with the lack of CaV1.3 current inhibition by either Quin or Oxo-M, supporting the importance of PLCβ-1 for both D2R and M1R signal transduction pathways [10, 52, 53]. Despite PLCβ-1’s absence, it was not obvious that no other variant of PLCβ would substitute for the missing PLCβ-1. The absence of PLCβ-1 and its apparent requirement for D2R signaling will be of interest to researchers who study MSNs. Interestingly, PLCβ-1 has been implicated in regulating growth and proliferation, where its absence results in uncontrolled cell proliferation [54, 55]. Thus a loss of PLCβ-1 may have occurred during the immortalization of ST14A cells.

In these experiments, we used a short splice variant of CaV1.3, CaV1.3b [30]. CaV1.3a, which has a longer C-terminus, was not inhibited by activation of D2Rs when expressed in HEK 293 cells [56]; however, CaV1.3a has been shown to bind the scaffolding protein Shank found in the postsynaptic density of synapses [57]. This association is necessary for CaV1.3 current inhibition by D2Rs in primary MSNs [11]. Although the lack of the long C-terminus also could explain the absence of channel modulation by D2Rs in ST14A cells, CaV1.3b LCTs can be modulated by both IGF-1 and AA. Exposure of SH-SY5Y cells expressing either CaV1.3a or CaV1.3b to IGF-1 enhances both currents and requires phosphorylation of S1486, a residue shared by both splice variants [58]. We have measured significant CaV1.3b inhibition by AA in transiently transfected HEK 293 cells, consistent with a potential transmembrane site of action [27].

Since AA release occurs in the striatum from both neurons and astrocytes [59–61] following stimulation of D2Rs [21] or M1Rs [62], we tested whether bath application of AA modulated LTC in ST14A cells to be certain that CaV1.3b could be modulated by molecules downstream of PLCβ-1. We were unsure whether AA would inhibit CaV1.3b channels in ST14A cells similarly to CaV1.3 channels in HEK 293 cells [27] or native LCTs in SCG neurons [23–25] since a wide range of actions have been reported for AA modulation of a variety of Ca²⁺ currents by other groups (see review by Roberts-Crowley et al [63]). We found that direct application of AA circumvented the lack of receptor-activated channel modulation in ST14A cells and inhibited CaV1.3b recombinant current. This finding demonstrates that CaV1.3 channels are capable of being modulated in ST14A cells as we have observed previously in HEK 293 cells [27]. The properties of CaV1.3 inhibition by AA in HEK 293 and ST14A cells appear similar. Additionally, we have found that Oxo-M inhibits currents from CaV1.3b in HEK 293 cells stably transfected with M1Rs (unpublished data). Lastly, we have found that activation of D2Rs by 10 μM Quin inhibits CaV2.2 currents in HEK 293 cells (unpublished data) demonstrating that this agonist protocol should be sufficient to activate D2Rs in ST14A cells. Therefore, the lack of current modulation by D2Rs or M1Rs, reported here, is supported by the absence of PLCβ-1 rather than an inability of CaV1.3b to respond to modulation.

Conclusions

ST14A cells are used as a model system for both the study and treatment of Huntington’s disease [64–69]. Despite the lack of D2R- or M1R-mediated Ca²⁺ channel modulation in ST14A cells reported here, this cell line was useful for elucidating that AA inhibits CaV1.3 channels. We hypothesize that the consistency of AA’s inhibitory actions on CaV1.3 across cell types will be of interest to researchers who study lipid signaling molecules. Moreover, we anticipate the D2R signaling cascade could be rescued by transfecting ST14A cells with PLCβ-1. Whether D2R signaling would then cause a lipid mediated inhibition of CaV1.3b LCTs awaits future studies. Thus, ST14A cells are a valuable tool for studying the biophysical properties of an isolated Ca²⁺ current and the modulation of these channels by signaling molecules within the context of a striatal background to aid in understanding neuronal malfunctions of the striatum.
Acknowledgments

We thank Michelle Ehrlich and Dr. Elena Cattaneo (Università degli Studi di Milano) for the ST14A cell line, Diane Lipscombe (Brown University) and Edward Perez-Reyes (University of Virginia) for channel cDNAs, Loretta Lee and TzuFeng (Annie) Chung of the UMASS Tissue Culture Facility for maintaining the ST14A, A9L and HEK 293 cells, and Maggie Lee for technical assistance. We also thank Rubing Zhao-Shea and John Heneghan for useful comments and Liwang Liu and Tora Mitra-Ganguli for review of the manuscript.

Author Contributions

Conceived and designed the experiments: MLR ARR. Performed the experiments: MLR. Analyzed the data: MLR. Contributed reagents/materials/analysis tools: ARR. Wrote the paper: MLR ARR.

References


