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Rapid in vivo forward genetic approach for identifying axon death genes in *Drosophila*

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Axons damaged by acute injury, toxic insults, or neurodegenerative diseases execute a poorly defined autodestruction signaling pathway leading to widespread fragmentation and functional loss. Here, we describe an approach to study Wallerian degeneration in the *Drosophila* L1 wing vein that allows for analysis of axon degenerative phenotypes with single-axon resolution in vivo. This method allows for the axotomy of specific subsets of axons followed by examination of progressive axonal degeneration and debris clearance alongside uninjured control axons. We developed new Flippase (FLP) reagents using proneural gene promoters to drive FLP expression very early in neural lineages. These tools allow for the production of mosaic clone populations with high efficiency in sensory neurons in the wing. We describe a collection of lines optimized for forward genetic mosaic screens using MARCM (mosaic analysis with a repressible cell marker; i.e., GFP-labeled, homozygous mutant) on all major autosomal arms (~95% of the fly genome). Finally, as a proof of principle we screened the X chromosome and identified a collection eight recessive and two dominant alleles of *highwire*, a ubiquitin E3 ligase required for axon degeneration. Similar unbiased forward genetic screens should help rapidly delineate axon death genes, thereby providing novel potential drug targets for therapeutic intervention to prevent axonal and synaptic loss.

**neurodegeneration | glial response**

Widespread axonal degeneration and synapse loss occurs during neurodegenerative disease and after neural trauma. These degenerative events result in disruption of neural circuit connectivity and ultimately functional impairment of the nervous system. Identifying molecular cascades that actively promote axonal self-destruction is a key goal. However, despite decades of work, remarkably little is known about the molecular pathways that drive the degeneration of neurites or synapses in any context (1, 2).

Axotomy-induced axon degeneration (termed Wallerian degeneration, WD) serves as a useful model to study the mechanisms of axonal self-destruction. When axons are severed, the portion of the axon distal to the injury site and its synapses undergo catastrophic fragmentation after a defined latent phase, and the resulting debris is eventually cleared by surrounding glial cells. The discovery of the spontaneous Wallerian degeneration slow (WldS) mouse revealed, surprisingly, that severed axons can in fact survive for weeks in the absence of a cell body (3). It also led to the proposal that “axon death” signaling cascades might exist, akin to apoptotic cell death programs, which actively drive the destruction of the axon (4).

Interestingly, WldS provides significant suppression in mouse models of progressive motor neuron disease and glaucoma (5–8), and moderate protection from chemotherapy-induced axon degeneration (9). These observations argue that defining the molecular mechanisms of axon degeneration in the context of WD could have an important therapeutic impact on the treatment of neurological disorders.

WD has not been subjected to comprehensive forward genetic screens, despite the fact that such approaches in model organisms, including *Drosophila melanogaster* and *Caenorhabditis elegans*, were critical for defining cellular degradation signaling pathways, such as autophagy (10) and apoptosis (11). Recently, we described the first forward genetic screen for loss-of-function mutations capable of blocking WD in *Drosophila*, and identified the kinase scaffolding molecule dSarm/Sarm1 (sterile alpha/Armadillo/Toll-interleukin receptor homology domain protein) as essential for WD in both the fly and mouse in vivo (12). Thus, unbiased genetic screens in *Drosophila* can lead to the identification of key genes required for mammalian axon degeneration after injury. This point is further supported by the recent discovery of the E3 ubiquitin ligase Highwire/Phr1, which is required for WD (13, 14).

The labor-intensive nature of current screens is a critical and limiting factor in identifying additional WD genes (F2 loss-of-function, dissections, immunohistochemistry, and so forth). Here, we describe a powerful forward genetic mosaic approach to identify loss-of-function mutations that suppress WD/axon death signaling. Our method also uses the *Drosophila* wing (15), but allows for axotomy and visualization of single axons, and the use of F1 forward genetic screening approaches. The collection of tools we describe should prove extremely useful in defining the molecular mechanisms of axon degeneration and we envision many additional uses in exploring the basic biology of the axon in vivo.

**Results**

**Visualizing WD with Single-Axon Resolution in the L1 Wing Vein.** To rapidly screen for mutants defective in axon degeneration, we sought to develop an in vivo system that contains the following characteristics: (i) rapid observation of individual axons (rather than an entire axon bundle) with minimal dissection; (ii) the opportunity to sever only a subset of axons so that one could

**Significance**

The elimination of neurites is a widespread event during the assembly of the nervous system, and in the mature brain after injury or in neurological disease. However, molecular pathways underlying neurite/axon degeneration in any context remain poorly defined. We have developed an in vivo genetic approach with single-axon precision in the *Drosophila* wing, allowing for the observation of the onset, progression, and clearance of axon degeneration alongside control axons. We generated genetic reagents to facilitate rapid in vivo isolation of ethyl methane sulphonate-induced mutants that are defective in axon degeneration. Additional unbiased and genome-wide forward genetic screens will provide novel potential drug targets for therapeutic intervention to prevent axonal and synaptic loss after injury or disease.

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observe axon death alongside healthy, uninjured axons; and (iii) the ability to initiate and visualize axon death without killing the fly (e.g., thereby allowing for F1-based mutagenesis screens). These criteria led us to explore the Drosophila nerve housed in the marginal (L1) wing vein, which contains both sensory neurons and glia (Fig. 1 and Fig. S1; see also below).

When screening through a number of peripheral nervous system Gal4 drivers, we identified candidate OK371-Gal4, which labels ~40 glutamatergic sensory neurons in the wing (Fig. L4) (16). The cell bodies of neurons expressing OK371-Gal4 are aligned along the length of the L1 vein (arrow in Fig. L4), and project their axons within the L1 vein into the thorax. Some of these axons project through the entire wing and are among the longest axons in Drosophila (17). We focused our imaging of these axons in a region of the L1 vein immediately posterior to the costal vein (Fig. L4, Inset). Hereafter, we will refer to this region as the proximal L1 vein.

The L1 vein allows for a graded level of axotomy, which results in two populations of axons: severed axons, whose cell bodies are distal to the injury site; and intact axons, whose cell bodies are proximal to the injury site. Axotomies were performed with nonlethal surgical cuts through the middle of the L1 vein with microdissection scissors in one wing; the other wing served as an intact control (Fig. L1). Flies were then placed back in vials for 0–7 d. Eventually, both wings were removed, and imaged by confocal microscopy (Fig. 1C). To determine the number of uninjured axons, we counted the number of residual cell bodies that remained in the proximal region of the injured wing. We found that 100% of the severed axons initiated WD (i.e., fragmentation) within 1 d postaxotomy (dpa), and that all axonal debris was absent from the L1 vein by 7 dpa (Fig. 1C and D). In contrast, uninjured axons remained morphologically intact.

### Conserved Molecular Mechanisms Modulate WD in the L1 Wing Vein

We reasoned that if the L1 wing vein nerve was to be a useful model to study WD, similar molecular programs should be in place as observed in other Drosophila or mouse axotomy models including protection by Wld<sup>D</sup> (18, 19). Wld<sup>D</sup> expression levels correlate with protection/conservation in severed axons, where high levels of Wld<sup>D</sup> result in strong protection (18). Recently, work by Fang et al. showed that axon degeneration in different wing sensory neurons are protected by Wld<sup>D</sup> (15). We tested the glutamatergic neurons for their sensitivity to Wld<sup>D</sup> by either strongly (5×UAS) overexpressing Wld<sup>D</sup> or an enzymatically impaired version, Wld<sup>D</sup>(dead20), or by low level (1×UAS) expression of Wld<sup>D</sup> (Fig. 1E, strong, intermediate, and mild protection, respectively). Whereas GFP<sup>D</sup> axonal debris was absent from severed wild-type axons from the L1 vein at 7 dpa (Fig. 1E, wild-type control), we found graded protection levels ranging from strong protection by high levels of Wld<sup>D</sup> (i.e., many severed but intact axons); intermediate protection by high levels of Wld<sup>D</sup>(dead2) and mild protection by low levels of Wld<sup>D</sup> (where fragmentation in severed axons had already occurred, but axonal debris was not yet fully cleared). These observations support the model of a dosage-dependent protection by Wld<sup>D</sup> of wing sensory neurons, similar to what has been described in mouse (18). Moreover, our production of the 1× Wld<sup>D</sup> construct demonstrates the wing can serve as a sensitized system in which to screen for modifiers of Wld<sup>D</sup> neuroprotection in vivo.

We also confirmed that programmed cell death is not involved in WD in the wing vein, as neuronal-specific overexpression of P35, a broad apoptosis inhibitor in Drosophila (21), did not alter the rate of axon degeneration at 7 dpa (Fig. 1E, apoptosis-defective). Local engulfing cells were required for active clearance of axonal debris: we assayed homozygous viable draper mutant animals [where clearance of axonal debris in the CNS is fully impaired after olfactory receptor neuron (ORN) ablation (19)], and found that GFP<sup>D</sup> axonal debris persisted at 7 dpa at similar signal intensities as compared side-by-side to an intact axon (arrowhead in Fig. 1E, engulfment-defective). Thus, Draper-dependent signaling mechanisms are required for efficient clearance of axonal debris in the L1 vein.

### Gial Cells in the L1 Wing Vein Are Diverse and Engulf Axonal Debris

The most likely candidate cell types executing engulfment of axon debris are local glial cells, which are known to reside in the Drosophila wing (22). To explore the morphological and functional diversity of glial subtypes in the L1 vein, we combined...
a number of Gal4- and LexA-based drivers, as well as GFP traps that label specific subsets of Drosophila glial cells (23–25). We also took advantage of another binary expression system and generated a number of new QF/QUAS-based tools to differentially label axons and glia side-by-side (Fig. 2A, Fig. S2, and see below).

To examine the histological relationship between neurons and glia, we first labeled all glia (pan-glial driver repo-LexA) and found the entire L1 vein was fully covered by glial membranes (Fig. 2B, Top). Glial cells could be divided into two main subtypes: wrapping glia (anr2-GFP protein trap) that ensheathe axons directly, and subperineurial glia (L82-Gal4) that surround both axons and wrapping glia (Fig. 2B, Middle and Bottom). To approximate the density of cells within these populations, we drove a nuclear marker in individual glial subtypes and counted the number of nuclei (Fig. 2C and D). Wrapping glial nuclei were found at a density of 2.6/100 μm, subperineurial glia at 4.1/100 μm, and total glial density was 6.6/100 μm. We suspect that these subsets represent the complete complement of glia in the L1 vein, as the sum of ensheathing and subperineurial glial nuclei is comparable to the number found when all glial nuclei were labeled (Fig. 2C and D).

To determine whether glial cells engulfed axonal debris after axotomy we used RNAi-mediated knockdown of draper using glial-specific Gal4 drivers (Fig. 2E and F). Depleting Draper from all glia suppressed clearance of axonal debris at levels equivalent to those found in homozygous draper-null mutants at 7 dpa. Knockdown of draper in wrapping glial also resulted in a strong suppression of clearance of axonal debris, whereas knockdown in subperineurial glia had a much more modest effect (Fig. 2E and F). Thus, glial cells—and in particular wrapping glia—are the key cell type required for clearance of axonal debris in the L1 wing vein. This tissue represents a histologically simple system in which to visualize axonal degeneration and neuron-glia interactions after injury with high cellular resolution (Fig. 2G).

A Tool Kit for Rapid Genome-Wide Forward Genetic Screens with Single-Axon Resolution. The observation of fine axonal structures within an axon bundle where 40 axons are labeled is challenging. Ideally one would label individual axons. We therefore combined the above-described Gal4-based markers for subsets of glutamatergic neurons with mosaic analysis with a repressible cell marker (MARM) (26). First, we optimized heat-shock–triggered Flippase expression (hs-FLP), which was sufficient to generate GFP+ MARM clones with single-axon resolution (Fig. 3A and B and Fig. S3). Based on these initial hs-FLP analyses, we concluded that labeling between three and five axons was optimal for visualizing axonal degeneration. Next, we cloned several promoters of pronuclear genes: we reasoned that these promoters were highly active during wing sensory neuron development to help pattern sensory organ precursors (SOPs), and therefore they could be excellent candidates for specific FLP expression (Fig. S4A and B). We created transgenic flies harboring three different SOP candidates, promotor-FLP transgenes: senseless-FLP (sense-FLP), achaete-FLP, and asense-FLP (sense-FLP), and extracellular matrix-FLP (emc-FLP), which is broadly active in epithelial cells before pronuclear cluster determination, thus less specific to SOP lineages (Table S1).

We prescreened these transgenic stocks to determine cell-type FLP activity (i.e., neurons, glia, or epithelial cells), and rescreened promising candidates for their relative strength in FLP activity in glutamatergic neurons (Fig. S4C), hs- and emc-FLP sources contained strong and broad (e.g., glial, neuronal, and epidermal) activity, whereas both asense-FLP and achaete-FLP showed strong cell-type restricted activity to neurons. However, asense-FLP (inserted on chromosomes 2 and 3, respectively) contained strong neuronal activity (Fig. S4C). We therefore decided to test asense-FLP (a sense-FLP (a)) in a MARCM background for both arms of chromosome 3. asense-FLP (a) was sufficient to induce GFP-labeled clones, and the induction of clone numbers was not only dependent on the chromosomal arm used for MARCM (1.2 ± 0.9 vs. 5.2 ± 1.7, FRT2A vs. FRT82B, respectively, in Table S2), but also on the copy number of asense-FLP (a), because increased dosage of asense-FLP leads to the production of increased numbers of MARCM clones in L1 wing vein sensory neurons (Fig. S5).

Transgene activity in the Drosophila genome is influenced by the genomic area surrounding the transgene insertion site. We therefore mobilized asense-FLP (a) to create random asense-FLP (a) insertions. These insertions were screened in a MARCM background to ultimately determine “optimally” paired FLP-sources and FRT lines with which one could generate reliably high (∼5) or low (1 or 2) MARCM clone induction rates, using any major chromosome arm (Fig. 3 C and D). Each newly isolated FLP source was tested in a MARCM assay (chromosomes 3, 2, and X in Tables S2, S3, and S4, respectively). This established collection of tools allows for genetic access to the vast majority of the Drosophila genome for studies of axon degeneration or basic axon biology using the L1 wing vein.

The ability to label individual axons with GFP in the intact wing allowed us to observe fine structural changes in axonal

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Alleles mutants, at least for 7 dpa. Whereas in highwire mutants could also protect other hs-FLP axons (24). (Bottom) Distribution of heat-shock MARCM clones on chromosomes 3L and 3R. Detailed distribution is shown in Fig. S5. (D) as-FLP sources as a tool for genome-wide clonal analysis. as-FLP source and Gal4 driver combinations induce many or few clones on each chromosomal arm. Individual as-FLP clones are termed MARCM clones (chromosome X, FRT19A; 2L: FRT40A; 2R: FRT1G13; 3L: FRT2A; 3R: FRT82B). (E) Examples of uninjured (ctl), as well as injured axon clones 6 and 12 hpa side-by-side of healthy axons, respectively (arrows, pronounced blebbing, swelling, and loss of axonal integrity, respectively). (F) Quantification of severed axons undergoing degeneration (n ≥ 60). (Scale bars, 5 μm.) Genotypes are described in SI Materials and Methods.

Fig. 3. Genome-wide clonal tool for analyses at single-axon resolution. (A) Examples of heat-shock-induced hs-FLP MARCM axon clones. (Top) GFP+ glutamatergic axon bundle. (Middle and Bottom) Examples of five and two MARCM clones, respectively. Cross-sections are shown on the right. Heat-shock treatment is shown in Fig. S3. (B) Distribution of heat-shock-induced MARCM clones (chromosome X, FRT19A). Dots indicate clone numbers per wing (n = 30). (C) Examples of as-FLP MARCM clones on chromosomes 3L and 3R. Detailed distribution is shown in Fig. S5. (D) as-FLP sources as a tool for genome-wide clonal analysis. as-FLP source and Gal4 driver combinations induce many or few clones on each chromosomal arm. Individual as-FLP clones are termed MARCM clones (chromosome X, FRT19A; 2L: FRT40A; 2R: FRT1G13; 3L: FRT2A; 3R: FRT82B). (E) Examples of uninjured (ctl), as well as injured axon clones 6 and 12 hpa side-by-side of healthy axons, respectively (arrows, pronounced blebbing, swelling, and loss of axonal integrity, respectively). (F) Quantification of severed axons undergoing degeneration (n ≥ 60). (Scale bars, 5 μm.) Genotypes are described in SI Materials and Methods.

integrity over time after axotomy, and compare them to uninjured control axons in the same preparation (Fig. 3 E and F). Within 6 h postaxotomy (hpa), 42% of all severed axons (n ≥ 60) underwent morphological changes that included the formation of spheroids [i.e., local swellings or focal axonal degeneration (27)] and the first signs of fragmentation (Fig. 3E, arrow). At 12 hpa, severed axons began to lose their integrity and contained discontinuous axon shafts, whereas control axons remained morphologically intact. This time course supports previous findings (19), but allows for the unambiguous scoring of breakdown of the integrity of even single axons.

**WD in Wing Sensory Neurons Requires dsarm.** To determine whether axon degeneration in the L1 vein was dSarm-dependent, we generated dsarm mutant MARCM clones in the wing. Whereas in wild-type all severed axons underwent fragmentation and were cleared from the vein by 7 dpa, we found that severed dsarm axons remained intact and were morphologically indistinguishable from adjacent uninjured dsarm axons or control axons (Fig. 4 A and B). Thus, axon death in the L1 vein requires dsarm function, similarly to sensory neurons in antennae, and multiple neuronal subtypes in mammals (12).

The wing allowed us also to carefully quantify the persistence of severed but intact dsarm axons by counting the number of cell bodies (i.e., axons that are severed) in the cut off wing: we found that 100% of all severed dsarm axons remained intact at 7 dpa, compared with 0% in wild-type (Fig. 4C). Thus, severed axons remain fully protected in dsarm mutants, at least for 7 dpa.

**Rapid Identification of Dominant and Recessive highwire Alleles Through Forward Genetic Screens in the Wing.** To test the efficiency of the wing as a screening tool, we ethyl methane sulphonate (EMS)-mutagenized ~1,500 X chromosomes, and screened those for mutants defective in axon degeneration. We identified several mutants exhibiting potent protection of severed axons at 7 dpa (Fig. 5A). For example, all severed x052 mutant axons remained morphologically intact and undistinguishable from uninjured control axons at 7 dpa (Fig. 5 B and C). To determine whether x052 mutants could also protect other Drosophila axons, we axotomized ORNs (19). Unilateral antennal ablation resulted in axon degeneration and debris removal of wild-type ORNs within 10 d (Fig. 5D, arrow); however, intact severed x052 clones were observed at 10, 30, and even 50 dpa. To identify the molecular nature in x052 responsible for the axon protective phenotype, we used the Drosophila GEM.app whole-genome sequencing analysis approach followed by complementation tests (28). Through this approach, we identified a premature stop in codon 2791 (CAG → TAG) of the highwire locus, which has recently been shown to be required for axon degeneration (13). We tested for allelism by performing complementation tests using a large N-terminal deletion allele of hiw termed hiwΔN (29). At 1 dpa, severed wild-type and heterozygous

**Fig. 4. dsarm is required for axon death in the wing.** (A) Examples of wild-type and dsarm mutant axon clones, uninjured and 7 dpa, respectively. (Scale bar, 5 μm.) (B) Quantification of attached cell bodies and axons (gray and white, respectively) in uninjured control and 7-dpa wings. Data are shown as average ± SEM (n = 24). (C) Potent protection of severed axons in dsarm mutant clones at 7 dpa. First, the number of cut off cell bodies was determined, and then remaining cell bodies/axons scored at 7 dpa. Shown is the percentage of severed axons of wild-type and dsarm mutants that remain morphologically intact at 7 dpa. Genotypes are described in SI Materials and Methods.
mutant axons underwent catastrophic fragmentation; however, hiw^{ΔN}/x052 axons remained morphologically intact, even at 7 dpa (Fig. 5E). Furthermore, severed x052 clones coexpressing the cDNA of either hiw or gfp::hiw fragmented and their debris was cleared within 7 dpa (Fig. 5F and G). Taken together, these data suggest that x052 is an allele of highwire.

Surprisingly, we identified a total of 10 hiw alleles in the same screen (by similar procedures as described above): hiw^{157}, hiw^{052}, hiw^{687}, hiw^{352}, hiw^{0354}, hiw^{2754}, and hiw^{022}, where the molecular nature was identified, and three alleles, hiw^{771}, hiw^{649}, and hiw^{704}, where the molecular lesion was not determined (Fig. 5H). Highwire belongs to the family of RING domain E3 ubiquitin ligases, where the E3 RING domain is involved in the direct transfer of ubiquitin (Ub) from E2–Ub complex to the target substrate (30). Unexpectedly, we found that two alleles, hiw^{0354} and hiw^{2754}, dominantly suppress axon degeneration (Fig. 5I). Interestingly, genome sequencing revealed that both hiw^{0354} and hiw^{2754} contain mutations in or close to the Ring domain (Fig. 5H). These data argue that specific loss of the Hiw Ring domain—which would lead to a protein predicted to bind its substrate but not transfer ubiquitin—can dominantly suppress axon degeneration. Consistent with this notion, Gal4/UAS-mediated overexpression of Highwire containing a nonfunctional RING domain (HiwΔRING) in wild-type animal phenocopies loss of hiw function in both NMJ structural analysis, based on sectioning issues with a very thin

Discussion
Our work helps to further establish Drosophila as a powerful in vivo system in which to explore molecular pathways mediating axon degeneration. Our collection of tools allows for labeling, manipulation, and visualization of single axons in vivo. Individual axons can be severed and the resulting onset and progression of axon degeneration can be observed alongside uninjured neighboring axons. This process allows for counting actual axon numbers in vivo for very precise determination of levels of protection in different mutant backgrounds, rather than simply visualizing a large bundle of axons and scoring remaining GFP intensity (15). When large bundles of GFP^+ axons are visualized, one cannot conclusively demonstrate that individual axons have remained morphologically intact.

The newly generated FLP sources driven by the proneural promoters are convenient tools to easily generate MARCM clones early in neural lineages: our collection thus simplifies for screening, the wing vein is not ideal for some applications

In summary, the above data show that forward genetic MARCM screens in the Drosophila wing, coupled with whole-genome resequencing, is an extremely approach to isolate axon death signaling genes and, in the case of EMS mutagenesis, generate an allelic series that is informative regarding protein function.
nerve embedded within a thick cuticle. Nevertheless, the produc-
tion of additional markers for visualizing live axons, organ-
elles, or even proteins certainly allows for detailed visualization of
axon biology in live or fixed preparations (31).

We also note that care must be taken when interpreting
phenotypes after L1 vein axotomy: transection of the marginal
vein not only severed axons, but also local glial cells. In some
cases, axonal debris persisted for over a week close to the injury
site, even in controls animals. We suspect this represents an in-
jury-induced impairment of glial engulfment function because of
degeneration, apoptosis, or necrosis of glia adjacent to the injury
site after axotomy. Indeed, we have observed cell bodies of
sensory neurons proximal to the injury site degenerating after
axonotomy, probably because of local glial loss and, in turn, de-
generation of now naked, unsupported neuron cell bodies.

However, such problems can be avoided by cutting the wing
midway along the L1 nerve and then image axonal degeneration
in the most proximal region of the wing (box in Fig. 1 L).

Finally, by performing a small-scale EMS pilot screen, fol-
lowed by whole-genome resequencing using the web-based
GEM.app tool (28), as well as rescue and complementation tests,
the wing allowed us to readily isolate several alleles of highwire,
a gene recently identified to be required for axon degeneration
both in Drosophila and in mammals (13, 14). Among these hiw
alleles, we isolated an early stop at coding position 1312 (hiw157
K1312stop), as well as two dominant alleles that affect the RING
domain (hiw27sd and hiw30sd). Surprisingly, endogenous expres-
sion levels of one copy of hiw predicted to contain a mutation in
the RING domain was sufficient to provide potent protection to
severed axons. Thus, the Hiw RING domain is a key functional
domain required to drive axonal degeneration. It remains to be
determined how these dominant hiw alleles block axon de-
generation (13, 29).

Loss-of-function screens in the wing vein represent a powerful
approach to dissect the molecular basis of axon degeneration.
Axonal and synaptic degeneration is a hallmark of every neu-
rodegenerative disease and occurs in peripheral neuropathies
(1, 2), but mechanisms promoting axonal self-destruction remain
elusive. The identification of new genes that regulate axonal
degeneration will advance our fundamental knowledge of a basic
biological axon death process, and provide exciting new potential
drug targets for intervention in neurodegenerative disease.

Materials and Methods

See SI Materials and Methods for more information. SI Materials and
Methods contains detailed descriptions of the wing axotomy assay and
subsequent wing microscopy, the wing engulfment assay, and quantification
of axon degeneration. It also includes information about the ORN unilateral
ablation assay, brain dissection, EMS mutagenesis, and genome-wide se-
quence analyses.

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5. Ferri A, Sanes JR, Coleman MP, Cunningham JM, Kato AC (2003) Inhibiting axon de-
genration and synapse loss attenuates apoptosis and disease progression in a mouse
7. Verghese PB, et al. (2011) Nicotinamide mononucleotide adenylyltransferase 1 pro-
tects against acute neurodegeneration in developing CNS by inhibiting excitotoxic-
8. Howell GR, et al. (2007) Axons of retinal ganglion cells are insulted in the optic nerve
11. Ellis HM, Horvitz HR (1986) Genetic control of programmed cell death in the nema-
nerve injury reveals an essential role of Nmnat in maintaining axonal integrity. Curr
transporter: A marker protein for motoneurons and glutamatergic centers in the brain.
17. Palka J, Schubiger M, Ellison RL (1983) The polarity of axon growth in the wings of
18. Mack TG, et al. (2001) Wallerian degeneration of injured axons and synapses is de-
enzymatic activity and N16-VCP interactions to suppress Wallerian degeneration.
Drosophila glial cells in the late embryonic and early larval ventral nerve cord. Roux’s
platform allow for rapid gene identification in D. melanogaster EMS screens. Biology
ophila neuromuscular junction: spatial, structural, and temporal requirements. J
Neurosci 25(42):9557–9566.
chem 78:399–434.
31. Fang Y, Soares L, Bonini NM (2013) Design and implementation of in vivo im-
810–819.