# RNA Interference and mRNA Silencing, 2004: How Far Will They Reach?

# Thoru Pederson

Department of Biochemistry and Molecular Pharmacology and Program in Cell Dynamics, University of Massachusetts Medical School, Worcester, Massachusetts 01605

Submitted October 9, 2003; Revised October 30, 2003; Accepted November 3, 2003

Monitoring Editor: Thomas Pollard

The discoveries of RNA interference and RNA-mediated posttranscriptional gene silencing have opened an unanticipated new window on the regulation of gene expression as well as a facile and highly effective tool for knocking down gene expression in many organisms and cells. In addition, RNA interference and RNA silencing may conceivably be exploited for human therapeutics sometime in the future, possibly bringing greater clinical impact than have the so far disappointing antisense endeavors. This essay summarizes recent developments and offers some personalized perspectives, with emphasis on what we do not yet know.

#### INTRODUCTION

The discovery of RNA interference (Guo and Kemphues, 1995; Fire *et al.*, 1998) was unusual in the postmodern era of molecular biology in that, like immunoglobulin gene rearrangement, for example, it was almost entirely unanticipated. Beyond the action of RNA interference and RNA silencing at the translational level of gene expression, RNA-mediated phenomena are now known to direct the transcription-level silencing of some genes, and even the "editing down" of an organism's genome by selective excision events in certain cases (Selker, 2003; Yao *et al.*, 2003).

In addition to possessing the element of surprise, RNA interference and RNA silencing also explained certain previous findings that had been puzzling, and thus the discoveries genuinely warranted the term breakthrough. In retrospect, were there any clues that were missed?

### Hints of RNAs That Base Pair with mRNA

In the early 1970s, intramolecular double-stranded regions of large nuclear RNA molecules were described in both mammalian cells and sea urchin embryos (Jelinek and Darnell, 1972; Kronenberg and Humphreys, 1972; Ryskov et al., 1972). These studies used extracted, deproteinized nuclear RNA, but the in vivo authenticity of these double-stranded RNA (dsRNA) regions of nuclear RNA was subsequently verified in intact, living cells (Calvet and Pederson, 1979). An intriguing finding was that although cytoplasmic mRNA did not contain these intramolecular dsRNA regions, the dsRNA elements of nuclear RNA could nevertheless hybridize with cytoplasmic mRNA (Stampfer et al., 1972; Naora and Whitelam, 1975; Ryskov et al., 1976; Jelinek et al., 1978). The implication was that a portion of a given nuclear dsRNA region is conserved in mRNA with the remainder of the dsRNA region being eliminated, i.e., the nuclear dsRNAcontaining molecules are mRNA precursors. But alterna-

Article published online ahead of print. Mol. Biol. Cell 10.1091/mbc.E03–10–0726. Article and publication date are available at www.molbiolcell.org/cgi/doi/10.1091/mbc.E03–10–0726.

tively, if a dsRNA-containing nuclear RNA molecule produced a small RNA that ended up hydrogen-bonded to a complementary stretch in a (separately encoded and generated) cytoplasmic mRNA, as in the RNA silencing of mRNA that has now been discovered, the observations would have been the same as the ones made.

The idea that intermolecular RNA–RNA interactions were involved in gene regulation was also advanced on the basis of other studies (Britten and Davidson, 1969), and later experiments revealed that duplex regions form between HeLa cell heterogeneous nuclear RNA molecules when annealed under certain conditions (Fedoroff *et al.*, 1977). During the same period, there were numerous reports of small, translation-suppressing RNAs associated with inactive messenger ribonucleoproteins, called translation control RNAs (tcRNAs). In some experiments, these small tcRNAs behaved as if hydrogen bonded to the inactive mRNA (Heywood and Kennedy, 1976). The tcRNA work never really caught on at the time, but of course now seems provocative in retrospect.

# Pre-microRNAs in the Genome

The endogenous pathway of mRNA silencing involves small RNAs, called microRNAs (abbreviated miRNAs) that are encoded in the organism's genome and expressed in the appropriate developmental schedule (Moss, 2002; Pasquinelli, 2002). miRNAs are derived from larger precursor molecules in the nucleus, the most proximal of which are ~70-nt molecules consisting of imperfectly paired hairpins (Lee et al., 2002, 2003; Seitz et al., 2003) Very little is known about how these "pre-miRNAs" are themselves produced but the available data are compatible with the derivation of the ~70-nt pre-miRNAs from considerably longer nuclear transcripts that lack translation open reading frames. It is interesting to recall in this respect that early studies on mammalian nuclear RNA revealed that a substantial portion of large, poly(A)-terminated molecules do not contain any sequences homologous to mRNA (Herman et al., 1976; Salditt-Georgieff et al., 1981). The functional significance of this puzzling population of nuclear RNA has remained elusive for a quarter of a century. Plausibly, it might harbor precursors of the pre-miRNAs, i.e., the "pre-pre-miRNAs."

<sup>\*</sup> Corresponding author. E-mail address: thoru.pederson@ umassmed.

The initial studies of the intramolecular RNA duplex fraction of heterogeneous nuclear RNA (hnRNA) in HeLa cells operatively defined these regions by their resistance to RNases A and T1 (Jelinek and Darnell, 1972; Kronenberg and Humphreys, 1972; Ryskov et al., 1972; Calvet and Pederson, 1977; 1978) or RNases A and T2 (Roberston et al., 1977; Jelinek, 1977). One of these investigations scrutinized these double-stranded regions in particular detail, based on their relative resistance to single-strand-specific versus doublestrand-specific conditions of RNase attack, and identified two distinct classes of dsRNA within hnRNA, one more perfectly base paired than the other (Calvet and Pederson, 1977). The pre-micro RNAs identified so far are ~70-nt RNAs with some base-pairing mismatches (Lagos-Quintana et al., 2001; Lau et al., 2001; Lee and Ambros, 2001; Mourelatos et al., 2002). It is not possible at present to relate these identified nuclear pre-miRNAs to the previously characterized dsRNA regions of hnRNA, but this is a high priority for understanding the biosynthesis of the miRNAs. In addition, there are other sequence families in mammalian genomes (Alu sequences in human) that produce, mostly via transcription by RNA polymerase III, small RNAs that contain stem-loop domains very similar to ones in the presently defined pre-miRNAs. Unfortunately, nothing else is presently known about the primary transcripts that give rise to pre-miRNAs, including the particular RNA polymerase that makes them (presumably pol II). A recent study demonstrated that a human pre-miRNA contains a 5'-phosphate, a 3'-OH, and a 1-4 nt 3' overhang, consistent with an RNase III-mediated derivation (Basyuk et al., 2003).

## Is RNA Interference Restricted to mRNA Targets?

What is the target "reach" of RNA interference and mRNA silencing? Until recently, all the demonstrated RNA interference targets were messenger RNAs. For example, a recent study has demonstrated that certain endogenous small interfering RNA (siRNA)-like RNAs in the protozoan Trypanosoma brucei are complexed with polyribosomes in a manner that is dependent on active translation (Djikeng et al., 2003). The extant reports using siRNAs to knock down RNA viral replication, a situation in which there are RNA species other than translating mRNA as potential targets, are almost certainly due to action on viral mRNAs (Dector et al., 2002; Jacque et al., 2002; Ge et al., 2003; McCaffrey et al., 2003; Novina et al., 2003). Reports that translationally repressed, maternal mRNAs become susceptible to RNA interference only upon their developmental translational activation (Svoboda et al., 2000; Kennerdell et al., 2002) have fueled the idea of a direct connection between RNAi or mRNA silencing and the target's active translational status. So, is there any basis for considering RNA interference or mRNA silencing more broadly than only targeting translating mRNAs?

At first, the most plausible initial interpretation of the discovery of RNA interference was the one deriving from an evolutionary perspective, namely, that of an ancient defense against RNA genome-based infectious or parasitic organisms, a defense retained in extant organisms, but one that has also been tinkered with and tooled over ~2.5 billion years into a developmental gene regulation pathway in the metazoan Eukarya (Zamore, 2002). Such a mechanism would have likely first operated on an mRNA encoded (or brought in as genomic RNA) by a RNA virus. This idea sounds very plausible on evolutionary grounds. As for mRNA silencing by small RNAs, there is ample precedent in prokaryotes so perhaps this was an early feature of gene regulation that evolved before, or at least independently of, double-stranded RNA-triggered mechanisms.

#### RNAi Hits non-mRNA

The idea that the action of RNAi is restricted to translating mRNAs was perhaps not ever assumed by the field's leaders, but it is certainly the prevailing operative view of most cell biologists using siRNAs to knockdown target mRNAs. By way of an example, in a recent collaborative study of mine in this journal (Wang *et al.*, 2003), the use of siRNA to knock down a relevant mRNA in a mammalian cell line worked beautifully, as is very typical, but the more unorthodox possibility of using siRNA to knock down the small, RNA polymerase III transcripts that were of even greater interest in the study was not pursued.

The notion that siRNA targets must be mRNAs has recently been dramatically overturned (Liang et al., 2003). Ironically, one of the (three) organisms in which this study was done was the very same one in which the aforementioned link of RNAi action to translating mRNA was reported, namely, T. brucei, one of the first organisms in which RNA interference was described (Ngo et al., 1998). These investigators decided to target a group of small nucleolar RNAs (snoRNAs) that play a role in the biosynthesis of rRNA in the nucleolus (Bachellerie et al., 2002). They expressed siRNAs complementary to these snoRNAs and observed their destruction (Liang et al., 2003). This study is the first to demonstrate that siRNA can attack a nonpolysomal, non-mRNA target and in a cellular compartment other than the cytoplasm, the nucleus. Moreover, the target RNAs in this study have 5' termini different than mRNA and also lack 3' poly(A) tails, ruling out these ends of RNA as being required for siRNA action.

Can knocking down (or out) non-mRNA "housekeeping" RNAs, such as snoRNAs, be anything beyond just a good learning curve for understanding the molecular biology of cells? Consider a protein called Ro. It is a human autoantigen that is complexed with a group of small, cytoplasmic non-mRNAs in higher animals (Tan, 1982). When the gene for the Ro protein was knocked out of mice (by transgenic, heritable means, not transiently by RNA interference), the animals displayed phenotypes (Xue et al., 2003) that have the potential to contribute, with further work, to advances in our understanding of two of the most challenging and important current problems in all of medicine: innate immunity and immunological tolerance. Knockdown of the mRNA for Ro protein, or of the Ro RNAs themselves, by siRNA in mammalian cells or animals might offer additional experimental dimensions, now empowered by the transgenic knockout results. There are many other non-mRNAs of unknown function in eukaryotic cells, and it is possible that they too can be approached by RNA interference. Some of these may themselves turn out to be elements of RNAmediated gene regulation. For example, can siRNAs knock down miRNAs?

The scale of RNA interference-mediated analysis of gene expression has now reached breathtaking levels in ideal organisms. In the first two large-scale applications of this approach, the mRNAs from ~5000 genes in *Caenorhabditis elegans* were systematically deleted, and more recently several studies of nearly comparable scale have been carried out in *Drosophila* cells (Pollard, 2003). Microarray-based approaches to the discovery of novel microRNAs are also now coming onto the scene (Krichevsky *et al.*, 2003), so our understanding of the endogenous pathways of mRNA silencing and related miRNA functions, i.e., establishment of heterochromatin or genome reorganization, can be anticipated to rapidly expand during 2004.

#### Envisioning the Patient

Major questions remain about the utility of these discoveries for medicine, including the pharmacokinetics of administered siRNAs, the challenges of gene therapy-based approaches to implanting an intracellular supply of a miRNA in patients, and, most sobering of all, the implausibility of mRNA knockdown approaches for conditions based on loss of function mutations. Also to be borne in mind is the possibility that initially seductive mRNA targets revealed by microarray analysis may turn out not to be at the true center of the pathogenic process in the case of noninfectious disease (as opposed to the more immediately promising viral disease opportunities for siRNAs.) There is also the question of whether knockdown of a desired target, perhaps to the good, might bring with it an undesired shutdown of certain other, essential mRNAs in response, due to the operation of feedback controls that sense the targeted mRNA's presence or activity. These questions surrounding the utility of these discoveries for medicine (or agriculture) do not, of course, diminish the deserved excitement about the discoveries of RNA interference and RNA silencing as science. This essay has endeavored to add at least a small degree of perspective to this explosively emerging field. It seems likely that there is much more biology to be uncovered, and it is possible that major applications in medicine and agriculture may ensue, driven by the reliable engine of basic research.

#### **ACKNOWLEDGMENTS**

I thank my colleague Phillip Zamore for offering enabling comments on an early draft. My work is supported by grants GM-21595 and GM-60551 from the National Institutes of Health and a grant from the Human Frontier Scientific Program Organization. By the rules of the National Institutes of Health, I am required to affirm that the views expressed in this article do not represent the official position of the U.S. government. (If they did, it is amusing to ponder what America might be up to.). This article is dedicated to the memory of Charles A. Janeway, whose intellectual passion and outgoing generosity deeply touched me on several occasions over the years.

## **REFERENCES**

Bachellerie, J.-P., Cavaille, J., and Huttenhofer, A. (2002). The expanding snoRNA world. Biochimie 84, 775–790.

Basyuk, E., Suavet, F., Doglio, A., Bordonné, R., and Bertraud, E. (2003). Human let-7 stem-loop precursors harbor features of RNase II cleavage products. Nucleic Acids Res. 31, 6593–6597.

Britten, R.J., and Davidson, E.H. (1969). Gene regulation for higher cells: a theory. Science  $165,\,349-357.$ 

Calvet, J.P., and Pederson, T. (1977). Secondary structure of heterogeneous nuclear RNA: two classes of double-stranded RNA in native ribonucleoprotein. Proc. Natl. Acad. Sci. USA 74, 3705–3709.

Calvet, J.P., and Pederson, T. (1978). Nucleoprotein organization of inverted repeat DNA transcripts in heterogeneous nuclear RNA-ribonucleoprotein particles from HeLa cells. J. Mol. Biol. 122, 361–378.

Calvet, J.P., and Pederson, T. (1979). Heterogeneous nuclear RNA double-stranded regions probed in living HeLa cells by crosslinking with the psoralen derivative aminomethyltrioxsalen. Proc. Natl. Acad. Sci. USA 76, 755–750

Dector, M.A., Romero, P., Lopez, S., and Arias, C.F. (2002). Rotavirus gene silencing by small interfering RNAs. EMBO Rep. 3, 1175–1180.

Djikeng, A., Shi, H., Tschudi, C., Shen, S., and Ullu, E. (2003). An siRNA ribonucleoprotein is found associated with polyribosomes in *Trypanosoma brucei*. RNA 9, 802–808.

Fedoroff, N., Wellauer, P. K. and Wall, R. (1977). Intermolecular duplexes in heterogeneous nuclear RNA from HeLa cells. Cell 10, 597-610.

Fire, A., Xu, S., Montgomery, M.K., Kostas, S.A., Driver, S.E., and Mello, C.C. (1998). Potent and specific genetic interference by double-stranded RNA in *Caenorhabditis elegans*. Nature 391, 806–811.

Ge, Q., McManus, M.T., Nguyen, T., Shen, C-H., Sharp, P.A., Eisen, H.N., and Chen, J. (2003). RNA interference of influenza virus production by directly

targeting mRNA for degradation and indirectly inhibiting all viral RNA transcription. Proc. Natl. Acad. Sci. USA 100, 2718–2723.

Guo, S., and Kemphues, K.J. (1995). par-1, a gene required for establishing polarity in C. elegans embryos, encodes a putative Ser/Thr kinase that is asymmetrically distributed. Cell 81, 611–620.

Herman, R.C., Williams, J.G., and Penman, S. (1976). Message and non-message sequences adjacent to poly(A) in steady state heterogeneous nuclear RNA of HeLa cells. Cell 7, 429–437.

Heywood, S.M., and Kennedy, D.S. (1976). Purification of myosin translational control RNA and its interaction with myosin messenger RNA. Biochemistry 15, 3314–3319.

Jacque, J.M., Triques, K., and Stevenson, M. (2002). Modulation of HIV-1 replication by RNA interference. Nature 418, 435–438.

Jelinek, W., and Darnell, J.E. (1972). Double-stranded regions in heterogeneous nuclear RNA from HeLa cells. Proc. Natl. Acad. Sci. USA 69, 2537–2541.

Jelinek, W.R. (1977). Specific nucleotide sequences in HeLa cell inverted repeated DNA: enrichment for sequences found in double-stranded regions of heterogeneous nuclear RNA. J. Mol. Biol. 115, 591–601.

Jelinek, W.R., Evans, R., Wilson, M., Salditt-Georgieff, M., and Darnell, J.E. (1978). Oligonucleotides in heterogeneous nuclear RNA: similarity of inverted repeats and RNA from repetitious DNA sites. Biochemistry 17, 2776–2783.

Kennerdell, J.R., Yamaguchi, S., and Carthew, R.W. (2002). RNAi is activated during Drosophila oocyte maturation in a manner dependent on aubergine and spindle-E. Genes Dev. 16, 1884–1889.

Krichevsky, A.M., King, K.S., Donahue, C.P., Khrapko, K., and Kosik, K.S. (2003). A microRNA array reveals extensive regulation of microRNAs during brain development. RNA 9, 1274–1281.

Kronenberg, L.H., and Humphreys, T. (1972). Double-stranded ribonucleic acid in sea urchin embryos. Biochemistry 11, 2020–2026.

Lagos-Quintana, M., Rauhut, R., Lendeckel, W., and Tuschl, T. (2001). Identification of novel genes coding for small expressed RNAs. Science 294, 853–858

Lau, N.C., Lim, L.P., Weinstein, E.G., and Bartel, D.P. (2001). An abundant class of tiny RNAs with probable regulatory roles in *Caenorhabditis elegans*. Science 294, 858–862.

Lee, R.C., and Ambros, V. (2001). An abundant class of tiny RNAs with probable regulatory roles in *Caenorhabditis elegans*. Science 294, 862–864.

Lee, Y. et al. (2003). The nuclear RNase III Drosha initiates microRNA processing. Nature 425, 415–419.

Lee, Y., Jeon, K., Lee, J.-T., Kim, S., and Kim, V.M. (2002). MicroRNA maturation: stepwise processing and subcellular localization. EMBO J. 21, 4663–4670

Liang, X.-H., Liu, Q., and Michaeli, S. (2003). Small nucleolar RNA interference induced by antisense or double-stranded RNA in trypanosomatids. Proc. Natl. Acad. Sci. USA 100, 7521–7526.

McCaffrey, A.P., Nakai, H., Pandey, K., Huang, Z., Salazar, F.H., Xu, H., Wieland, S.F., Marion, P.L., and Kay, M.A. (2003). Inhibition of hepatitis B virus in mice by RNA interference. Nat. Biotech. 21, 639–644.

Moss, E.G. (2002). MicroRNAs: hidden in the genome. Curr. Biol. 12, R138-R140.

Mourelatos, Z., Dostie, J., Paushkin, S., Sharma, A., Charroux, B., Abel, L., Rappsilber, J., Mann, M., and Dreyfuss, G. (2002). miRNPs: a novel class of ribonucleoproteins containing numerous microRNAs. Genes Dev. 16, 720–729.

Naora, H., and Whitelam, J.M. (1975). Presence of sequences hybridisable to dsRNA in cytoplasmic mRNA molecules. Nature 256, 756–759.

Ngo, H., Tschudi, C., Gull, K., and Ullu, E. (1998). Double-stranded RNA induces mRNA degradation in *Trypanosoma brucei*. Proc. Natl. Acad. Sci. USA 95, 14687–14692.

Novina, C.D., Murray, M.F., Dykxhoorn, D.M., Beresford, P.J., Riess, J., Lee, S.-K., Collman, R.G., Lieberman, J., Shankar, P., and Sharp, P.A. (2002). siRNA-directed inhibition of HIV-1 infection. Nat. Med. *8*, 681–686.

Pasquinelli, A.E. (2002). MicroRNAs: deviants no longer. Trends Genet. 18, 171–173

Pollard, T. D. (2003). Functional genomics of cell morphology using RNA interference: pick your style, broad or deep. J. Biol. 2, 25 online (http://jbiol.com/content/2/4/25).

Robertson, H.D., Dickson, E., and Jelinek, W. (1977). Determination of nucleotide sequences from double-stranded regions of HeLa cell nuclear RNA. J. Mol. Biol. 115, 571–589.

Vol. 15, February 2004 409

Ryskov, A.P., Farashyan, V.R., and Georgiev, G.P. (1972). Ribonuclease-stable base sequences specific exclusively for giant dRNA. Biochim. Biophys. Acta 262, 568–572.

Ryskov, A.P., Kramerov, D.A., and Georgiev, G.P. (1976). The structural organization of nuclear messenger RNA precursor. I. Reassociation and hybridization properties of double-stranded hairpin-like loops in messenger RNA precursor. Biochim. Biophys. Acta 447, 214–229.

Salditt-Georgieff, M., Harpold, M.M., Wilson, M.C., and Darnell, J.E. (1981). Large heterogeneous nuclear ribonucleic acid has three times as many 5' caps as polyadenylic acid segments, and most caps do not enter polyribosomes. Mol. Cell. Biol. 1, 179–187.

Seitz, H., Youngson, N., Lin, S.-P., Dalbert, S., Paulsen, M., Bachellerie, J.-P., Ferguson-Smith, A.C., and Cavaille, J. (2003). Imprinted microRNA genes transcribed antisense to a reciprocally imprinted retrotransposon-like gene. Nat. Genet. 34, 261–262.

Selker, E.U. (2003). Molecular biology. A self-help guide for a trim genome. Science 300, 1517-1518.

Stampfer, M., Rosbash, M., Huang, A.S., and Baltimore, D. (1972). Complementarity between messenger RNA and nuclear RNA from HeLa cells. Biochem. Biophys. Res. Commun. 49, 217–224.

Svoboda, P., Stein, P., Hayashi, H., and Schultz, R.M. (2000). Selective reduction of dormant maternal mRNAs in mouse oocytes by RNA interference. Development 127, 4147–4156.

Tan, E.M. (1982). Autoantibodies to nuclear antigens (ANA): their immunobiology and medicine. Adv. Immunol. 33, 167–240.

Wang, C., Politz, J.C., Pederson, T., and Huang, S. (2003). RNA polymerase III transcripts and the PTB protein are essential for the integrity of the perinucleolar compartment. Mol. Biol. Cell 14, 2425–2435.

Xue, D. et al. (2003). A lupus-like syndrome develops in mice lacking the Ro 60-kDa protein, a major lupus autoantigen. Proc. Natl. Acad. Sci. USA *100*, 7503–7508.

Yao, M.-C., Fuller, P., and Xi, X. (2003). Programmed DNA deletion as an RNA-guided system of genome defense. Science 300, 1581–1584.

Zamore, P.D. (2002). Ancient pathways programmed by small RNAs. Science 296, 1265–1269.