Factor H-Dependent Alternative Pathway Inhibition Mediated by Porin B Contributes to Virulence of Neisseria meningitidis

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Factor H-Dependent Alternative Pathway Inhibition Mediated by Porin B Contributes to Virulence of Neisseria meningitidis

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ABSTRACT The identification of “factor H binding protein (fHbp)-null” invasive meningococcal isolates and the realization that widespread use of fHbp-based vaccines could herald selection of such strains prompted us to characterize novel mechanisms of alternative pathway (AP) inhibition on meningococci. Of seven strains engineered to lack four known AP-inhibiting molecules, capsular polysaccharide, lipooligosaccharide sialic acid, fHbp, and neisserial surface protein A (quadruple mutants), four strains inhibited human AP-mediated C3 deposition. All four expressed the porin B2 (PorB2) molecule, and three strains belonged to the hypervirulent ST-11 lineage. Consistent with reduced C3 deposition, the rate of C3a generation by a PorB2 isolate was lower than that by a PorB3 strain. Allelic replacement of PorB3 with PorB2, in both encapsulated and unencapsulated strains, confirmed the role of PorB2 in AP inhibition. Expression of PorB2 increased resistance to complement-dependent killing relative to that seen in an isogenic PorB3-expressing strain. Adult rabbit and mouse APs were unimpeded on all mutants, and human fH inhibited nonhuman C3 deposition on PorB2-expressing strains, which provided functional evidence for human fH-dependent AP regulation by PorB2. Low-affinity binding of full-length human fH to quadruple mutants expressing PorB2 was demonstrated. fH-like protein 1 (FHL-1; contains fH domains 1 through 7) and fH domains 6 and 7 fused to IgG Fc bound to one PorB2-expressing quadruple mutant, which suggested that fH domains 6 and 7 may interact with PorB2. These results associate PorB2 expression with serum resistance and presage the appearance of fHbp-null and hypervirulent ST-11 isolates that may evade killing by fHbp-based vaccines.

IMPORTANT The widespread use of antimeningococcal vaccines based on factor H (fH) binding protein (fHbp) is imminent. Meningococci that lack fHbp were recently isolated from persons with invasive disease, and these fHbp-null strains could spawn vaccine failure. Our report provides a molecular basis for an explanation of how fHbp-null strains may evade the host immune system. Meningococci possess several mechanisms to subvert killing by the alternative pathway (AP) of complement, including production of the fHbp and NspA fH binding proteins. Here we show that a meningococcal protein called porin B2 (PorB2) contributes to inhibition of the AP on the bacterial surface. A majority of the “fHbp-null” isolates identified, as well as all members of a “hypervirulent” lineage (called ST-11), express PorB2. Our findings highlight the potential for the emergence of fHbp-negative strains that are able to regulate the AP and may be associated with fHbp vaccine failure.

Neisseria meningitidis is an important cause of bacterial meningitis and sepsis worldwide. The complement system is an important component of innate immune defenses against this pathogen. Individuals deficient in terminal complement components or in components of the alternative pathway (AP) are at an increased risk of meningococcal disease (1). A key feature of the AP of complement is a positive-feedback loop that amplifies C3b deposition on microbial surfaces (2). The AP also plays an important role in maximizing the killing activity elicited by select antimeningococcal antibodies (Abs) (3). Under physiological conditions, factor H (fH) plays a major role in limiting unwanted activation of the AP (4). fH acts as a cofactor for the factor I-mediated cleavage of C3b to inactive C3b (iC3b) and also serves to limit C3 activation by irreversibly dissociating the AP C3 convertase (C3b, Bb). Microorganisms can hijack host fH and use this molecule to protect themselves from immune attack by downregulating deposition of C3b and inactivating C3b that is deposited.

The meningococcus possesses several distinct and often redundant mechanisms to limit AP activation. Meningococci directly bind human fH through two surface molecules, fH binding protein (fHbp) (5) and neisserial surface protein A (NspA) (6); both serve to limit C3 deposition and enhance resistance of the organism to complement-dependent killing. In addition, sialylation of meningococcal lacto-N-neotetraose (LNT) lipooligosaccharide (LOS) regulates the AP by enhancing the interactions of the
C-terminal domains of fH with surface-bound C3 fragments (7) and the group B and group C capsular polysaccharides limit AP-mediated deposition of C3 (8, 9).

Based on observations that binding of fH to meningococci was human specific (10), we developed a human fH transgenic (Tg) rat which enhanced the ability of H44/76 to cause bacteremia (bacteremia was observed after challenge with \(5 \times 10^5\) CFU) (11). Rather unexpectedly, deleting both known fH ligands, fHbp, and NspA, did not diminish the ability of H44/76 to cause bacteremia in human fH Tg infant rats (11); this result suggested the existence of an additional human fH-dependent mechanism(s) for complement evasion. LOS sialylation was identified as one such mechanism (7, 11).

Recent work by Lucidarme et al. (12) identified meningococcal isolates that lacked fHbp expression that had been collected from patients with invasive disease, which demonstrated that in select strains, fHbp was dispensable for virulence in humans. These findings imply that certain meningococcal strains possess novel means of inhibiting complement activation in the absence of fHbp. This observation is particularly important given that fHbp-based vaccines are on the verge of clinical use.

The aim of the present study was to identify novel mechanisms meningococci use to regulate the AP that could contribute to the virulence of this pathogen in the absence of fHbp. These mechanisms could contribute to the ability of fHbp-negative meningococci to maintain virulence and escape killing by fHbp-containing vaccines.

RESULTS
Meningococcal PorB2 strains regulate the human AP. In order to study AP regulation in the absence of factors known to modulate C3 deposition on meningococci, we created mutants that lacked expression of capsular polysaccharide, LOS sialic acid, fHbp, and NspA. Strains harboring these four mutations simultaneously are referred to here as “quadruple mutants.” Quadruple mutants, derived from 7 diverse strains of \(N. meningitidis\) (see Table S1 in the supplemental material), were screened for deposition of human C3 following incubation with normal human serum-MgCl\(_2\)-EGTA (NHS-Mg/EGTA) (20% [vol/vol]); Mg/EGTA blocked classical and lectin pathway activation and allowed assessment of AP activation only (Fig. 1). Significantly lower C3 deposition was seen on quadruple mutants generated from four strains that expressed the PorB2 molecule than on mutants generated from strains that expressed PorB3 (Fig. 1A and B). Similar results were seen with C2-depleted serum (only AP intact; classical and lectin pathways blocked) and with sera from two additional donors (data not shown).

To ensure that serum factors other than AP components did not contribute significantly to the differential C3 deposition on the meningococcal mutants, we examined C3 deposition on the strains using purified C3, factor B, factor D, and fH (Fig. 1C). Again, each of the four PorB2-expressing quadruple mutants showed significantly (\(P < 0.01\)) lower C3 deposition than each of the three PorB3 isolates. These data indicate that despite loss of capsule, LOS sialic acid, fHbp, and NspA, select PorB2 strains effectively regulated the human AP.

Decreased C3 deposition in PorB2 strains is associated with slower C3 activation. Decreased C3 deposition could result from reduced targets for covalent binding of C3 (i.e., fewer electron-donating –OH groups) or decreased activation of C3 on the bacterial surface (i.e., because of increased fH binding) or a combination of these events. The quantitative differences seen by flow cytometry (Fig. 1) were confirmed by Western blotting (see Fig. S1 in the supplemental material), which revealed both LOS and Opa as major targets for iC3b across all isolates, as has been previously described for \(Neisseria\) (13). LOS structure can modulate complement activation on neisseriae (7, 8, 14). All strains expressed an ~4.5-kDa LOS species; NZ98/254 showed an additional 3.6-kDa LOS species (data not shown). Thus, there was no obvious correlation between the LOS migration pattern and the amount of C3 deposited.

Activation of C3 is accompanied by release of the C3a fragment, and measurement of the rate of C3a generation reflects C3 activation kinetics. Based on previously published data that measured the rates of complement activation (9), we focused on C3a production during the first 15 min of incubation. Quadruple mutants of 4243 (PorB2) and H44/76 (PorB3) were chosen as examples of low- and high-level C3 binders, respectively. As shown in Fig. 1D, the reaction mixture containing the 4243 quadruple mutant generated C3a at a rate lower than that seen with H44/76. The total amounts of C3a in the reaction mixture reached similar levels across strains at time points at or beyond 20 min, likely because C3 convertases in the fluid phase contribute to overall complement activation (9). These data suggest that lower C3 deposition on strain 4243 occurs at least in part because of a lower rate of C3 activation.

Human fH regulates the AP of nonhuman complement on PorB2-expressing quadruple mutants. Binding of fH to the known meningococcal ligands for fH, fHbp, and NspA is human specific (6, 10). To determine if regulation of the AP by PorB2-bearing strains was also specific to humans, we measured deposition of rabbit and mouse C3 using adult sera. PorB2 did not regulate the AP of these adult animals, and similar amounts of C3 were deposited on all quadruple mutants (Fig. 2, black bars). The addition of purified human fH to these nonhuman sera resulted in significantly greater downregulation of the AP on PorB2 strains relative to that seen with PorB3 isolates (Fig. 2, gray bars). Fluorescence-activated cell sorter (FACS) tracings of a representative experiment are shown in Fig. S2 in the supplemental material. These data provide strong evidence for human fH-dependent regulation of the AP by PorB2-bearing strains compared to PorB3-bearing strains.

Confirmation of the role of PorB2 in regulating the human AP. PorB is a major outer membrane protein that contributes to the fitness of a strain (15), and its deletion could have pleiotropic effects with respect to bacterial growth in addition to any interactions with the complement system. To avoid these concerns, we created allelic exchanges of the entire porB2 and porB3 sequences in a homologous background and compared C3 depostions in these isogenic strains that differed only in PorB types. Two sets of mutants were created; the first set lacked fHbp, NspA, capsule, and LOS (H44/76 “quadruple”-PorB3 and H44/76 “quadruple”-PorB2), while the second set lacked fHbp and NspA but elaborated capsule and sialylated LOS (H44/76 fHbp nspA-PorB3 and H44/76 fHbp nspA-PorB2).

H44/76 that expressed PorB2 showed greater downregulation of C3 deposition than H44/76 that expressed its own PorB3 molecule (Fig. 3). The effect was most pronounced in the absence of capsule (Fig. 3A; “Cap−”) but was also observed in the encapsulated (Fig. 3C; “Cap+”) background (quantitative comparisons
are shown in Fig. 3B and D, respectively). These data provide strong evidence for a role for PorB2 in regulating the human AP on meningococci.

C3 deposition on PorB2-expressing strains displays a bimodal distribution (Fig. 1B and 3A; see also Fig. S3 in the supplemental material). These data suggest that there are two populations that differentially regulate the AP. PorB2 expression was normally distributed (unimodal) and did not correlate with variation in C3 deposition. Regions of low, intermediate, and high PorB2 expression from within the normally distributed PorB2-positive population all revealed bimodal C3 deposition (Fig. S3). Thus, molecules other than PorB2 could contribute to the observed heterogeneity in C3 deposition.

Binding of fH, fH/Fc fusion proteins, and fH-like protein 1 (FHL-1) to meningococci lacking fHbp and NspA. Having provided evidence for human fH-dependent downregulation of the AP by PorB2-bearing strains, we compared the abilities of H44/76 “quadruple”-PorB3 and H44/76 “quadruple”-PorB2 to bind to human fH. Using standard FACS analysis (Fig. 4A [see also Fig. S4A in the supplemental material]; labeled “no cross-linker”) and Western blot (data not shown) methods, we did not detect binding of full-length human fH (100 μg/ml) to either strain. Similar negative results were obtained with the 7 quadruple mutants that express either PorB2 (C2120, W171, 4243, and 2996) or PorB3 (A2594, H44/76, and NZ98/254) (Fig. 4B [see also Fig. S4B]; labeled “no cross-linker”). To enhance detection of low-affinity interactions between fH and PorB, we added paraformaldehyde to cross-link bound fH to the bacterial surface prior to washing and then detected bound fH by FACS analysis (the terms “low” and “high” affinity are henceforth used to refer to low-level fH binding that can be detected only with a cross-linker and high-level fH binding that can be detected without a cross-linker). In the presence of the cross-linker, the quadruple mutant of H44/76 expressing PorB2 bound greater amounts of fH than the isogenic H44/76 bearing PorB3 (Fig. 4A [see also Fig. S4A]; labeled “paraformaldehyde cross-linker”). In other experiments, quadruple
Expression of PorB2 enhances serum resistance. To determine the relative roles of PorB2 and PorB3 in serum resistance, we compared the abilities of H44/76 fHbp nspA-PorB3 and H44/76 fHbp nspA-PorB2 to resist killing by human serum. Strains lacking fHbp and NspA were used to avoid the confounding effects of fH binding to these ligands. Serum was absorbed against a mixture of fHbp and NspA to remove strain-specific antibodies. Absorbed serum (20%) alone did not kill either strain (Fig. 5). Expression of PorB2 by H44/76 fHbp nspA resulted in greater resistance to complement-dependent killing mediated by anti-group B capsule IgG antibody SEAM 12 (19).
than expression of PorB3 (Fig. 5). Absorbed serum treated with Mg/EGTA to selectively activate the AP did not initiate killing of either strain in the presence or absence of SEAM 12 (100% survival in both instances). As previously reported for other antime-ningococcal Abs (3), the AP was required for maximal killing; blocking the AP with an anti-factor Bb monoclonal antibody (MAb) decreased killing of H44/76 fHbp nspA-PorB3 and H44/76 fHbp nspA-PorB2; “Cap+”). Bacteria were incubated with NHS-Mg/EGTA (20% [vol/vol]), and C3 deposited on the bacterial surface was measured by FACS analysis. (A and C) Representative histogram tracings of C3 deposition on the unencapsulated (Cap-) and encapsulated (Cap+) strains. Bacteria incubated with heat-inactivated serum served as negative controls (dashed lines). (B and D) Quantitative representation (percentage of positive events on the y axis; mean [SEM] of the results of three independent experiments) of C3 deposition on the Cap- and Cap+ isogenic PorB mutants, respectively. The percentage of positive events relative to control organisms incubated with heat-inactivated serum (gated to yield 5% of positive events in the negative-control sample as shown in Fig. 1B [+ve]) is represented on the y axis. Wild-type parent strains H44/76 and 4243 are shown as comparators. Note the different y-axis scales in panels B and D. ***, P < 0.001; **, P < 0.01.

**FIG 3** PorB2 mediates regulation of the human AP. Isogenic strains that differed only in PorB expression (either PorB3H44/76 or PorB24243) were created in H44/76. All mutants lacked fHbp and NspA and were either unencapsulated with no LOS sialic acid (“quadruple”; “Cap-”) or possessed a capsule and LOS acid (H44/76 fHbp nspA-PorB3 and H44/76 fHbp nspA-PorB2; “Cap+”). Bacteria were incubated with NHS-Mg/EGTA (20% [vol/vol]), and C3 deposited on the bacterial surface was measured by FACS analysis. (A and C) Representative histogram tracings of C3 deposition on the unencapsulated (Cap-) and encapsulated (Cap+) strains. Bacteria incubated with heat-inactivated serum served as negative controls (dashed lines). (B and D) Quantitative representation (percentage of positive events on the y axis; mean [SEM] of the results of three independent experiments) of C3 deposition on the Cap- and Cap+ isogenic PorB mutants, respectively. The percentage of positive events relative to control organisms incubated with heat-inactivated serum (gated to yield 5% of positive events in the negative-control sample as shown in Fig. 1B [+ve]) is represented on the y axis. Wild-type parent strains H44/76 and 4243 are shown as comparators. Note the different y-axis scales in panels B and D. ***, P < 0.001; **, P < 0.01.

**DISCUSSION**

PorB, one of the most abundant proteins in the neisserial outer membrane, is an integral membrane protein organized as a β barrel with 16 transmembrane domains and 8 predicted surface loops (20). PorB functions as a porin, allowing the passage of small molecules across the outer membrane, and has also been implicated in virulence. Gonococcal PorB has been shown to play a key role in regulation of both the AP and classical pathways of complement by virtue of its ability to bind to fH and C4BP, respectively (21, 22). Meningococcal PorB is divided into two classes, PorB2 and PorB3, which are mutually exclusive and are expressed...
from alternate alleles (PorB2 and PorB3) at the porB locus. PorB3 of N. meningitidis serves as a ligand for Toll-like receptor 2 (TLR2) and activates cells through a TLR2/TLR1-dependent pathway (23). The goal of our study was to examine AP regulation in strains that did not rely on fHbp or NspA expression. We have identified a novel role for meningococcal PorB2 in regulation of the AP of complement. Concurrent with our studies, Lucidarme and colleagues described 29 invasive meningococcal isolates that lacked fHbp expression (12). Of these strains, nine belonged to the “hypervirulent” ST-11 lineage. It is noteworthy that strains that belong to the hypervirulent ST-11 clonal complex express PorB2. In an ongoing analysis of select strains reported by Lucidarme et al., three of four non-ST-11 isolates also expressed PorB2 (24). Although anti-capsule MAb SEAM 12 could overcome AP regulation and initiate in vitro killing of meningococci, maximal killing required the AP. Further, our studies demonstrated that expression of PorB2, relative to PorB3, enhanced the ability of H44/76 fHbp nspA to evade complement-mediated killing. Similarly, expression of PorB2 may render strains more resistant to complement-dependent killing in humans. The ability to regulate complement and cause invasive disease in the absence of fHbp expression may offer these strains a distinct advantage when faced with immune pressure by anti-fHbp antibodies, which is a likely occurrence with clinical use of fHbp-containing vaccines (25, 26).

Natural disruption of PorA (27, 28) and NadA (IS1301) (29), which are both components of the proposed vaccine, has also been reported in the ET-15 subset of the ST-11 complex.

FIG 4 PorB2 binds full-length fH, fH domains 6 and 7, and FHL-1. (A and B) Binding of human fH to quadruple mutants of H44/76 that express either PorB344/76 (H44/76-PorB3) or PorB24243 (H44/76-PorB2) (A) and quadruple mutants of N. meningitidis strains that express either PorB3 (A2594, H44/76, and NZ98/254) or PorB2 (C2120, W171, 4243, and 2996) (B). Quadruple mutants were incubated with purified fH (100 μg/ml), and bound fH was measured by FACS analysis (graphs labeled “no cross-linker”). In some experiments, bound fH was cross-linked to the surface with paraformaldehyde (final concentration, 1% [vol/vol]) prior to detection by FACS analysis (graphs labeled “paraformaldehyde cross-linker”). The negative control (control) was a reaction mixture that lacked fH (neg. control). Binding of fH to wild-type H44/76 that expresses fHbp, NspA, capsule, and LOS sialic acid is shown as a positive control in panel B (pos. control). The amount of bound fH is represented on the y axis as fold change in median fluorescence relative to the H44/76 quadruple mutant that expresses PorB3 (in panel A) or the H44/76 quadruple mutant (in panel B) with paraformaldehyde cross-linker. Each bar represents the SEM of the results of 2 to 3 independent experiments (note that some error bars are small and are not easily seen on the figure). **, P < 0.01; *, P < 0.05 (1-way ANOVA with Tukey’s posttest for pairwise comparisons). (C) PorB24243 binds to H67/Fc. Binding of human fH67/Fc to quadruple mutants of H44/76 (blue line), 4243 (green line), and H44/76 that express either PorB3 (H44/76-PorB3; red line) or PorB2 (H44/76-PorB2; gray shading) was measured by FACS analysis. The negative control (broken line) was a reaction mixture that lacked fH67/Fc. x axis, fluorescence on a log10 scale; y axis, counts. Data from an experiment representative of at least two reproducible repeats are shown. (D) A quadruple mutant of strain 4243 binds FHL-1. Binding of FHL-1 (10 μg/ml) to the 4243 quadruple mutant (gray-shaded histogram) was measured by FACS analysis using polyclonal goat anti-human fH. FHL-1 was omitted in the negative control (broken line). The positive control shows binding of FHL-1 to wild-type H44/76 (solid black line). Numbers alongside histograms represent the median fluorescence, and the fill/outline corresponds to that of the histograms.
We have provided considerable functional evidence for human fH-dependent inhibition of the AP by PorB2. A novel finding of this study was the low affinity of the interaction between full-length fH and PorB2 on intact bacteria. fH is present in serum at concentrations of 200 to 500 μg/ml (~1.3 to 3.3 μM), and, given these high ligand concentrations, it is plausible that low-affinity interactions between PorB2 and fH suffice for complement inhibition. Higher-affinity fH–ligand interactions may be important in niches where the concentration of fH is lower, such as at mucosal surfaces. Several studies have highlighted a lack of correlation between the amount of fH binding to meningococci as revealed by flow cytometry and complement inhibition on the bacterial surface. Seib et al. (30) and Dunphy et al. (31) have shown that neither differences in fHbp–fH affinity (over 2 orders of magnitude) nor differences in the amount of fH bound to bacteria by FACS predicted resistance to complement–dependent killing. Another example is the NspA–fH interaction; despite the fact that capsule and full-length LOS expression decreased fH binding, the NspA-fH interaction was demonstrated under these conditions (9). This “double” fHbp deletion mutant was also sensitive to infant rat complement-mediated lysis (such as, downregulation of C3 deposition).

We were able to demonstrate high-affinity binding of FHL-1 and fH67/Fc to strains expressing PorB2H44. These findings were confirmed in strain FAM18, which expresses a PorB2 with an amino acid sequence identical to that of 4243. High-affinity binding of these “fragments” of fH, but not the full-length fH, to strains expressing PorB2H44 may result from structural constraints in intact fH (32, 33) that reduce its affinity for intact bacteria. A more detailed understanding of the regions of PorB that are required for functional interactions with fH may aid in the prediction of specific PorB molecules that may be involved in complement evasion.

This report sheds light on an important immune evasion function mediated by meningococcal PorB. In light of the functional role of PorB in complement evasion, it is conceivable that implementation of fHbp-containing vaccines (25, 26) may drive (or select for) loss of fHbp expression in combination with positive selection for specific PorB molecules that are optimized for AP regulation and functional interaction with fH.

MATERIALS AND METHODS

Ethics statement. This study was approved by the Committee for the Protection of Human Subjects in Research at the University of Massachusetts Medical School. All subjects who donated blood for this study provided written informed consent.

Bacterial strains and culture conditions. Characteristics of the wild-type strains used to derive the mutants used in this study are listed in Table S1 in the supplemental material. N. meningitidis bacteria were routinely grown on chocolate agar plates supplemented with IsoVitalex equivalent at 37°C in an atmosphere with 5% CO2. GC plates supplemented with IsoVitalex equivalent were used for bacterial selection. Escherichia coli (Invitrogen, Carlsbad, CA) were cultured in Luria-Bertani broth or agar with antibiotics as needed. Antibiotics were used at the concentrations indicated in Table S2 in the supplemental material.

Strains were rendered unencapsulated by interruption of mynB (group A) or siaD (group B, C, or W135) (6, 34). Insertional inactivation of ics (Ist::kan) abrogated LOS sialylation of group B, C, and W135 isolates as previously described (6, 34). Group A strains do not sialylate their LOS unless cytidine monophospho-N-acetylneuraminic acid (CMP-NANA) is added to growth media. fHbp and NspA expression was abrogated (fHbp::erm [5] and nspA::spc [6], respectively) as previously described.

PorB deletion mutants of H44/76 fHbp nspa expressing either PorB2H44 or PorB2 open bar) or PorB3H44/76 (PorB3; gray bar) in 20% absorbed serum that contained anti-group B capsule antibody, SEAM 12, at the concentrations indicated. Heat-inactivated (HI) sera and NHS without added SEAM 12 (0 μg/ml) served as controls. Percent survival relative to time 0 is plotted on the y axis. Each bar represents the SEM of the results of three independent observations.

**,** P < 0.01; ****, P < 0.0001 (2-way ANOVA).

% survival

SEAM12 (μg/ml)

PorB2

PorB3

FIG 5 PorB2 expression enhances serum resistance. Data represent percent survival of isogenic H44/76 fHbp nspa expressing either PorB2H44 (PorB2; open bar) or PorB3H44/76 (PorB3; gray bar) in 20% absorbed serum that contained anti-group B capsule antibody, SEAM 12, at the concentrations indicated. Heat-inactivated (HI) sera and NHS without added SEAM 12 (0 μg/ml) served as controls. Percent survival relative to time 0 is plotted on the y axis. Each bar represents the SEM of the results of three independent observations.

**,** P < 0.01; ****, P < 0.0001 (2-way ANOVA).
pathways and isolate the AP as the only active pathway, MgCl₂ and EGTA (10 mM each) were added to NHS (NH₄-Mg/EGTA).

Absorbed sera, in which antibodies specific to H44/76 fHbp nspA PorB3 and H44/76 fHbp nspA PorB2 were removed, were prepared as follows. EDTA (10 mM) was added to fresh NHS, and the sera were dialyzed against phosphate-buffered saline (PBS)–1 mM EDTA for 4 h at 4°C using a 10-kDa molecular mass cutoff. Meningococcal strains were subjected to passage on overnight plates and added onto fresh chocolate agar plates and grown for 5 h. Bacteria were harvested and washed once with PBS–1 mM EDTA. Dialyzed serum was absorbed twice with bacteria (1-ml serum with a mixture of 100 ml of packed cells from each of the 2 mutant strains) processed end over end for 30 min at 4°C. The serum was filtered (0.22-μm pore-size filter), divided into aliquots, and stored at −80°C. Ca²⁺ and Mg²⁺ (2 mM each) were added prior to use. Similar amounts of fH were detected (MAb 90X) in NHS and absorbed sera spotted on polyvinylidene difluoride (PVDF), which suggested that the fH was not depleted by the absorption. The hemolytic activity of serum was confirmed using a Total hemolytic complement kit.

Mouse complement was obtained by cardiac puncture of adult BALB/c mice. Blood was allowed to clot at 22°C for 20 min, followed by incubation at 4°C for 20 min, and then spun, and the supernatant was divided into aliquots and stored at −80°C. Adult rabbit complement was purchased from Cedarlane Laboratories.

Bacterial killing by NHS. Susceptibility of neisserial strains to complement-mediated killing was determined using a serum bactericidal assay as described previously (6). Anti-group B capsule IgG antibody SEAM 12 (19) was added to absorbed sera at the concentrations indicated. Survival was calculated as the number of viable colonies at time 30 min relative to time 0.

Purified complement components. Factors B and D were from Complement Technology, Inc. (Tyler, TX). C3 was isolated from human plasma by polyethylene glycol (PEG) precipitation and chromatography over DEAE Sephacel using a modification of previously established methods (35). NHS immunodepleted of C3 (Complement Technology, Inc.) regained full hemolytic activity upon addition of the purified C3 to a concentration of 500 μg/ml, using a Total hemolytic complement kit (The Binding Site, Birmingham, United Kingdom).

C3 deposition on bacteria using purified complement proteins. To deposit C3 on bacteria using purified AP components, −10³ bacteria in Hanks balanced salt solution (HBSS)–1 mM MgCl₂–0.1% bovine serum albumin (BSA) were incubated with C3 (500 μg/ml), factor B (100 μg/ml), and fH (100 μg/ml) followed by factor D (2 μg/ml) in a final reaction volume of 50 μl for 30 min at 37°C. Deposited C3 was measured by FACS analysis.

Antibodies. Human, rabbit, and mouse C3 deposition on bacteria was measured using fluorescein isothiocyanate (FITC)-conjugated anti-human C3 (BioDesign/Meridian Life Sciences), anti-guinea pig C3, and anti-human IgG subclass, alternative complement pathway activation, and epitope density on bactericidal activity of antibodies to meningococcal factor H binding protein. Infect. Immun.

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