IC97 Is a Novel Intermediate Chain of I1 Dynein That Interacts with Tubulin and Regulates Interdoublet Sliding

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Our goal is to understand the assembly and regulation of flagellar dyneins, particularly the Chlamydomonas inner arm dynein called I1 dynein. Here, we focus on the uncharacterized I1-dynein IC IC97. The IC97 gene encodes a novel IC without notable structural domains. IC97 shares homology with the murine lung adenoma susceptibility 1 (Las1) protein—a candidate tumor suppressor gene implicated in lung tumorigenesis. Multiple, independent biochemical assays determined that IC97 interacts with both α- and β-tubulin subunits within the axoneme. I1-dynein assembly mutants suggest that IC97 interacts with both the IC138 and IC40 subunits within the I1-dynein motor complex and that IC97 is part of a regulatory complex that contains IC138. Microtubule sliding assays, using axonemes containing I1 dynein but devoid of IC97, show reduced microtubule sliding velocities that are not rescued by kinase inhibitors, revealing a critical role for IC97 in I1-dynein function and control of dynein-driven motility.

INTRODUCTION

Dyneins are minus-end–directed microtubule motors important for a variety of cellular functions, including membrane-bound organelle transport, assembly and orientation of the mitotic spindle, nuclear migration, assembly of the Golgi apparatus, and ciliary and flagellar motility. In the ciliary/flagellar axoneme, the outer and inner dynein arms convert the energy derived from ATP hydrolysis into microtubule sliding, which in turn drives flagellar beating and bending. Analysis using the model genetic organism Chlamydomonas reinhardtii has revealed that the inner arm dynein system is responsible for generation of the flagellar waveform—the size and shape of the flagellar bend (Brokaw and Kamiya, 1987; Kamiya, 2002; King and Kamiya, 2009). The inner dynein arms, of which there are at least seven isoforms (King and Kamiya, 2009), are heterogeneous in composition and structural arrangement on the axoneme, binding to the axoneme in precise locations to form part of a 96-nm repeating module along each doublet microtubule (Goodenough and Heuser, 1985; Piperno et al., 1990; Burgess et al., 1991; Mastronarde et al., 1992; Porter et al., 1992; Nicastro et al., 2006; Bui et al., 2008). The two-headed I1 dynein isoform, also called dynein f, is distributed uniformly along the length of the axoneme as a triad structure located proximal to each radial spoke S1 (Nicastro et al., 2006; Porter and Sale, 2000; Wirschell et al., 2007).

I1 dynein is among the best characterized inner arm isoforms, containing two heavy chains (α-HC and β-HC), three intermediate chains (IC138, IC140, and IC97—also termed TcTex1 and TcTex2b) (Table 1; (Piperno et al., 1990; Myster et al., 1997; Harrision et al., 1998; Pazour et al., 1998; Perrone et al., 1998b, 2000; Yang and Sale, 1998; DiBella et al., 2004b; Hendrickson et al., 2004). Recently, a new I1-dynein–associated protein, flagellar-associated protein (FAP)120, has been identified that interacts with IC138, LC7b, and IC97 (Ikeda et al., 2008; Bower et al., 2009). Multiple lines of evidence demonstrate that I1 dynein is an unusual dynein motor that plays a critical regulatory role in the axoneme (Smith and Sale, 1991; Kotani et al., 2007; Wirschell et al., 2007). The I1-dynein complex has been implicated as a target of the regulatory signals that control flagellar motility (Porter et al., 1992; Porter and Sale, 2000; Smith and Yang, 2004; Wirschell et al., 2007). Flagella that are lacking I1 dynein exhibit defective flagellar waveform and phototaxis, indicating that I1 dynein plays a role in these processes (Brokaw and Kamiya, 1987; Brokaw, 1994; King and Dutcher, 1997; Okita et al., 2005). In Chlamydomonas, four independent loci, when defective, result in a specific failure to assemble the I1-dynein complex in the axoneme (DiBella and King, 2001). Three of these loci encode I1-dynein subunits; ida1, ida2, and ida7 mutants are defective in the α-HC, β-HC, and IC140, respectively (Myster et al., 1997; Perrone et al., 1998, 2000). Of particular use are novel
IC97 interacts with both IC140 and IC138 at the base of the dynein complex. IC97 forms part of a regulatory subcomplex of I1-dynein that control flagellar motility (reviewed in Porter and Sale, 2009; Bower et al., 2008) that is now referred to as the IC138 subcomplex. Mutations in I1-dynein subunits that permit assembly of partial I1-dynein complexes reveal that both IC97 and IC138, although not necessary for I1-dynein assembly, are required for regulation of I1-dynein activity. One such mutant, a new allele at the LC8 locus, fla14-3, assembles I1-dynein complexes that are devoid of IC97. fla14-3 mutant axonemes show reduced microtubule sliding velocities that are not rescued by kinase inhibitors even though IC138 seems to become dephosphorylated, demonstrating that IC97 plays a critical role in regulation of I1-dynein activity.

### MATERIALS AND METHODS

#### Strains and Culture Conditions

Chlamydomonas strains used in this study are summarized in Table 2. Cells were grown in Tris-acetate-phosphate medium or L-medium, with aerated 14:10-h light:dark cycle (Harris, 1989; Harris, 2009).

#### Cloning and Sequencing of IC97 Sequences

The cloning of IC97 was based on tandem mass spectrometry (MS/MS) identification of peptides derived from band-purified IC97 protein. Gene model C_850038 in the JGI Chlamydomonas genome database version 2.0 (http://shake.jgi-psf.org/chlre2/chlre2.home.html) encodes the IC97 gene (Supplemental Figure S1) and was used to design primers (Integrated DNA Technologies, Coralville, IA) for polymerase chain reaction (PCR) amplification of IC97 cDNA sequences (cDNA library 7 provided by Greg Pazour (University of Massachusetts, Amherst, MA). PCR products were cloned into pGEM-T-Easy (Promega, Madison, WI) or pcRI2.1/GW8/TOPO cloning vectors (Invitrogen, Carlsbad, CA) and sequenced to verify intron–exon boundaries (DNA Sequencing Facility, Iowa State University, Ames, IA). The full-length cDNA is 2,973 kb.

An IC97 cDNA probe containing base pairs 1343–2685 (Supplemental Figure S1B) was used to screen a BAC library (Clemson University Genomics Institute, Clemson, SC) and a 9.791-kb genomic EcoRI-HindIII fragment containing the IC97 gene was identified and subcloned into pBluescript SK-vector (pplasmid IC97-H). IC97 is found in the flagellar proteome as FAP94 (Pazour et al., 2005) and is published under accession FJ156240 and FJ156241 for the gene and cDNA sequences, respectively.

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### Table 1. I1-dynein components and mutants

<table>
<thead>
<tr>
<th>Subunit</th>
<th>Mol. wt.</th>
<th>Gene/mutant</th>
<th>Properties</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>1aHC</td>
<td>523</td>
<td>ida1 (pf9, pf30)</td>
<td>Lacks inner arm II, slow swimming</td>
<td>Kamiya et al. (1991)</td>
</tr>
<tr>
<td>1pHC</td>
<td>511</td>
<td>ida2</td>
<td>Lacks inner arm II, slow swimming</td>
<td>Kamiya et al. (1991)</td>
</tr>
<tr>
<td>IC140</td>
<td>140</td>
<td>ida7</td>
<td>Lacks inner arm II, slow swimming</td>
<td>Porter et al. (1992)</td>
</tr>
<tr>
<td>IC138</td>
<td>138</td>
<td>bop5</td>
<td>Partial I1 assembly, regulatory phosphoprotein part of IC138 subcomplex</td>
<td>Yang et al. (1998)</td>
</tr>
<tr>
<td>IC97 (IC110)</td>
<td>90 (110)</td>
<td></td>
<td>Non-WD repeat protein, homology to Las1/Casc1 proteins, part of IC138 subcomplex</td>
<td>This study</td>
</tr>
<tr>
<td>LC8</td>
<td>10</td>
<td>fla14</td>
<td>Required for flagellar assembly, part of I1 dynein, radial spokes and outer dynein arm</td>
<td>Yang et al. (2001)</td>
</tr>
<tr>
<td>LC7a</td>
<td>14</td>
<td>oda15</td>
<td>Required for outer arm assembly, slow swimming, associates with I1 dynein and may interact with LC7b</td>
<td>Pazour and Witman (2000)</td>
</tr>
<tr>
<td>LC7b</td>
<td>11</td>
<td></td>
<td>LC7/Robl family member, interacts with IC138, interacts with LC3 and DC2 of ODA</td>
<td>DiBella et al. (2004a)</td>
</tr>
<tr>
<td>Tctex1</td>
<td>13</td>
<td></td>
<td>Dimeric protein, potential cargo binding activity, also found in cytoplasmic dynein</td>
<td>Hendrickson et al. (2004)</td>
</tr>
<tr>
<td>Tctex2b</td>
<td>13.7</td>
<td>pf16-D2</td>
<td>Not required for I1 assembly, stabilizes I1 dynein</td>
<td>DiBella et al. (2004a)</td>
</tr>
<tr>
<td>FAP120</td>
<td>42</td>
<td></td>
<td>Not required for I1 assembly, ankryn repeat protein, part of IC138 subcomplex</td>
<td>Ikeda et al. (2009)</td>
</tr>
</tbody>
</table>
Table 2. Chlamydomonas strains used in this study

<table>
<thead>
<tr>
<th>Strain</th>
<th>Defect</th>
<th>Motility</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>CC124</td>
<td>WT</td>
<td>WT</td>
<td>Yang (1989, 2009)</td>
</tr>
<tr>
<td>CC1331 ( fla14-3)\ab</td>
<td>Radial spokes, I1 dynein, retrograde IFT</td>
<td>Paralyzed</td>
<td>Perrone et al. (2006)</td>
</tr>
<tr>
<td>CC-4077 (ida7-1:IC140 5A)\c</td>
<td>Truncated IC140; lacks outer dynein arm</td>
<td>Truncated IC140; lacks outer dynein arm</td>
<td>This study</td>
</tr>
<tr>
<td>CC-4080 ( bop5-1)</td>
<td>Partial I1 dynein assembly; IC138 point mutant</td>
<td>Slow-smooth swimming</td>
<td>Bower et al. (2000)</td>
</tr>
<tr>
<td>CC-2664 (ida1)</td>
<td>Lacks I1 dynein; 1oHC mutant</td>
<td>Slow-smooth swimming</td>
<td>Kamiya et al. (1990)</td>
</tr>
<tr>
<td>CC-2666 (ida2)</td>
<td>Lacks I1 dynein; 1BHC mutant</td>
<td>Slow-smooth swimming</td>
<td>Kamiya et al. (1990)</td>
</tr>
<tr>
<td>CC-2668 (ida3)</td>
<td>Lacks I1 dynein; gene product unknown</td>
<td>Slow-smooth swimming</td>
<td>Kamiya et al. (1990)</td>
</tr>
<tr>
<td>CC-2670 (ida4)</td>
<td>Lacks inner arm isoforms a, c, and d</td>
<td>Slow-smooth swimming</td>
<td>Kamiya et al. (1990)</td>
</tr>
<tr>
<td>CC-3921 (ida7)</td>
<td>Lacks I1 dynein; IC140 mutant</td>
<td>Slow-smooth swimming</td>
<td>Kamiya et al. (1990)</td>
</tr>
<tr>
<td>CC-262 (pf17 mt-)</td>
<td>Lacks radial spoke head; RSP1 mutant</td>
<td>Paralyzed</td>
<td>McVittie (1972)</td>
</tr>
<tr>
<td>5c12B</td>
<td>HA-tagged α-tubulin</td>
<td>Paralyzed</td>
<td>Kozminski et al. (1993)</td>
</tr>
<tr>
<td>pf28 pf30 ssh1\d</td>
<td>Lacks outer dynein arm; Lacks I1 dynein; suppressor of short mutation</td>
<td>Paralyzed</td>
<td>Ledizet and Piperno (1986)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Freshour et al. (2007)</td>
</tr>
</tbody>
</table>

\a Yang, Yang, Wirschell, and Davis (unpublished) determined that the pf5 mutation in strain CC1331 is an allele at the FLA14 locus (fla14-3) that encodes the LC8 protein. Other strains denoted as pf5 in the Chlamydomonas Stock Center contain a mutation that is tightly linked to the FLA14 locus and thus are not fla14-3/pf5 alleles.

\b Assembly defects in fla14-3 include an effect on the retrograde IFT motor as evidenced by assembly of half to full-length flagellae; no defects in assembly of the outer dynein arm are observed; II-dynein assembly defects are restricted to loss of IC97 and FAP120 specifically; and the radial spokes are reduced resulting in the paralysed flagellar phenotype.

\c The ida7-1::IC140 5A strain was first described in Perrone et al. (1998).

\d Strain pf28 pf30 ssh1 is a triple mutant lacking both the outer dynein arm and I1 dynein; the ssh1 mutation allows for wild-type length flagella in the double dynein mutant background (LeDizet and Piperno, 1995; Freshour et al., 2006).

**Computational Analyses**

Sequence alignments were performed using LaserGene SeqMan (DNASTar, Madison, WI). The LaserGene EditSeq module was used for translations and to estimate the predicted mass of the IC97 protein. Primers were designed using Primer3 (www.genome.wi.mit.edu/cgi-bin/primer/primer3) (Rozen and Skaletsky, 2000). The BLAST program (Altschul et al., 1990) was used to search for homologous sequences. The COILS program (www.ch.embnet.org/software/COILS_form.html) was used to predict coiled-coil regions in the protein sequence.

**Isolation of Axonemes, Dynein Purification, and Biochemical Analyses**

Flagella were isolated as described previously (Witman, 1986) and demembranated using NP-40 (Calbiochem, San Diego, CA). Axonemes were resuspended in HMDKEP (30 mM HEPES, pH 7.4, 5 mM MgSO₄, 1 mM dithiothreitol [DTT], 0.5 mM EDTA, 25 mM KCl, and 1 mM phenylmethylsulfonyl fluoride) plus the protease inhibitors apotinin and leupeptin (Sigma-Aldrich, St. Louis MO). Dyneins were extracted in HMDKEP plus 0.6 M KCl and fractionated by fast-performance liquid chromatography (FPLC) on a Mono Q column (GE Healthcare, Piscataway, NJ) as described previously (Kagami and Kamiya, 1995).

Chemical cross-linking was carried out using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) by treating microtubule-bound I1-dynein fractions or treating isolated axonemes with EDC for 30–60 min at room temperature, respectively (Benashski and King, 2000; Wirschell et al., 2008). All cross-linking reactions were performed in HMEK buffer (30 mM HEPES, pH 7.4, 5 mM MgO₄, 0.5 mM EDTA, and 25 mM KCl) supplemented with protease inhibitors. Reactions were neutralized with a 20-fold molar excess of β-mercaptoethanol. Cross-linked axonemes were collected by centrifugation and resuspended to 2 mg/ml with sample buffer. SDS-PAGE was performed by standard techniques using Precision Plus dual-color protein standards to estimate relative mobility (Mⱼ) (Bio-Rad Laboratories, Hercules, CA).

IC138 hyperphosphorylation was determined by comparing untreated and phosphate-treated axonemes (calf intestinal phosphatase [CIP]) by SDS-PAGE. Briefly, axonemes were treated with buffer or CIP for 30 min at room temperature and then directly fixed for SDS-PAGE analysis.

IC138 phosphorylation status was determined by in vitro phosphorylation of isolated axonemes using radioactive [γ-32P]ATP (PerkinElmer Life and Analytical Sciences, Boston, MA) (Yang and Sale, 2000). p17 and fla14-3 axonemes were resuspended to 5 mg/ml in CK reaction buffer (25 mM Tris, pH 8.0, 0.05 mM EDTA, 0.1% β-mercaptoethanol, 3.5 mM magnesium acetate, 0.01% Brij 35, 10 mM NaCl, and 30 μM sodium orthovanadate) and incubated at 30°C for 30 min. [γ-32P]ATP was added to a final concentration of 40 μM, and axonemes were incubated at 30°C for another 5 min. Samples were immediately fixed with sample buffer for analysis on 5% SDS-PAGE, transferred to nitrocellulose, and subsequently detected by autoradiography. The migration of IC138 was confirmed by Western blot of unlabeled axonemes run on the same gel.

MS/MS analyses of band-purified IC97 and the IC97 cross-linked product were performed at University of Massachusetts Proteomic Mass Spectrometry Laboratory by Dr. John Leszyk. MS/MS spectra were used to search the JGI Chlamydomonas Genome database, version 2.0.

**Reconstitution of II-Dynein Complexes onto Isolated Axonemes**

FPLC-purified II dynein and IC97 fractions derived from the double mutant ida7-1::IC140 5A ssh1 were dialyzed and then combined with isolated pf28 pf30 ssh1 axonemes. The reactions were incubated for 15 min at room temperature in HMDKEP buffer. Axonemes and supernatant fractions were collected by centrifugation and reconstitution of II-dynein components analyzed by Western blot.

**Antibody Production and Western Blot Analyses**

A restriction fragment containing nucleotides 1737-2492 of the IC97 cDNA (Supplemental Figure S1B) was digested from clone pMW173.1 with BamHI and EcoRI and subcloned into the pet28b expression vector (Novagen, San Diego, CA) to create plasmid pMW185.1 (Supplemental Figure S1B).
Cloning IC97 Sequences
To verify the exact sequence of the protein encoded by gene model C_850038, we designed primers based on predicted exons and the peptides identified by MS/MS. Sequences from a wild-type Chlamydomonas cDNA library were amplified by PCR and sequenced to confirm the complete open reading frame (ORF) plus 412-base pairs of 5' and 282-base pairs of 3' UTRs. The ORF predicts a 759-amino acid protein with a mass of 81.5 kDa (Supplemental Figure S1A). Using the antisense strand of the cDNA as a probe, we performed a Northern blot on RNA derived from cells either before or during flagellar regeneration. An ~2.8-kb transcript is detected that is up-regulated in response to deflagellation and flagellar regrowth (data not shown), a hallmark of mRNAs encoding flagellar proteins (Silflow et al., 1982).

IC97 Is a Non-WD Repeat IC That Has Orthologues in Ciliated Organisms
The predicted protein was analyzed by BLAST and found to have homology to several proteins, including a Ciona intestinalis axonal protein (BAB88834.1) and a murine protein, Lsl1 (Zhang et al., 2003; Manenti et al., 2004), that is part of the multigenePasI locus associated with an inherited predisposition to pulmonary carcinoma in mice. Alignment of theChlamydomonasand murine proteins shows the homology spans the entire sequence (Figure 1). Other than a small region in the N terminus that is predicted to form a coiled-coil, the IC97 protein does not have any notable motifs including WD-repeat domains that are found in other dynein intermediate chains.

IC97 Antiserum Detects the IC97 Component of I1 Dynein
To determine whether the gene we identified by MS/MS does indeed encode the IC97 subunit of I1 dynein, we made a specific polyclonal immune sera to an epitope-tagged fusion protein containing amino acids 443-693 of the predicted IC97 protein (Supplemental Figure S1B). In Western blots of isolated axonemes, the antibody detects a band having a $M_r$ of ~90,000 on SDS-PAGE (Figure 2A), well in agreement with the predicted mass of 81.5 kDa ($M_r$ of 90,000 is estimated from our SDS-PAGE gels. The mass of IC97 has been previously estimated as 97 and 110 kDa; Porter et al., 1992; Habermacher and Sale, 1997. This discrepancy is most likely due to differences in the molecular weight standards used). This band is specifically missing in I1-dynein–deficient axonemes (ida1, ida3, and ida7), indicating that the antisera detects an I1-dynein component. To further verify this, we purified I1 dynein by FPLC fractionation. The antisera detects the same size band in purified I1-dynein (dynein-f peak) fractions (Figure 2B). These results demonstrate that our antibody specifically detects the IC97 component of I1 dynein.

IC97 Interacts with $\alpha$- and $\beta$-Tubulin Components
The IC97 protein sequence does not predict any notable domains or motifs that would lend insight into the function of IC97. Thus, to better understand the role of IC97 in I1 dynein, we took advantage of EDC chemical cross-linking to identify IC97-binding proteins. EDC is a proven reagent for identifying axonemal interactions, particularly for the axonemal dyneins (King et al., 1991; Yang and Sale, 1998; Benashski and King, 2000; DiBella et al., 2004a; Hendrickson et al., 2004). Western blots of EDC-treated axonemes were probed with the IC97 antibody, which detected an EDC cross-linked product, with an $M_r$ of ~140,000, suggesting
IC97 (M, of ~90,000) is in direct contact with an axonemal protein with an estimated mass of 50 kDa (Figure 3A).

To determine the identity of the IC97-interacting protein, we used the IC97 antiserum to immunoprecipitate the IC97 cross-linked product in sufficient yield for identification (Figure 3B). MS/MS analysis of the IC97 cross-linked product identified peptides corresponding to both α- and β-tubulin in addition to IC97. The identification of IC97 peptides demonstrates that the band isolated for MS/MS is the IC97 cross-linked product. The Mr of the cross-linked product is ~140,000 on SDS-PAGE, suggesting that IC97 does not cross-link to a tubulin dimer, but rather IC97 independently cross-links to α- and β-tubulin. Hence, the cross-linked product is a mixture of IC97-α-tubulin and IC97-β-tubulin species. In support of this model, the ratio of IC97:α-tubulin:β-tubulin peptides recovered is 1.5:1:1, indicating that both α- and β-tubulin are present in equivalent amounts compared with a nearly twofold amount of IC97.

To further validate the results obtained by MS/MS, we used several approaches. First, we immunoprecipitated the IC97 cross-linked product (Figure 3C, IC97) from SDS-solubilized EDC-treated axonemes containing an HA-tagged α-tubulin (Kozminski et al., 1993). The IC97 cross-linked product is detected using an antibody to the HA tag, indicating that it contains the epitope-tagged tubulin (Figure 3C, HA). Second, we bound purified I1 dynein to Taxol-stabilized microtubules and then performed EDC cross-linking. The IC97 cross-linked product is formed indicating that IC97 interacts with tubulin components in vitro (Figure 3D). Third, we took advantage of a blot overlay assay that uses biotinylated tubulin as a probe to identify tubulin-interacting proteins (Yanagisawa and Kamiya, 2004). Blot overlays of purified dynein fractions detect atubulin-interacting band at the expected size for IC97 in purified-I1 dynein (dynein-f peak) fractions (asterisk in Figure 3E).

IC97 Interacts with IC138 and IC140 within I1 Dynein

To further characterize how IC97 assembles within I1-dynein, we took advantage of unique I1-dynein mutants that assemble partial I1-dynein complexes. Figure 4A shows Western blots of isolated axonemes from three mutant IC97 (M, of ~90,000) is in direct contact with an axonemal protein with an estimated mass of 50 kDa (Figure 3A).

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To further characterize how IC97 assembles within I1-dynein, we took advantage of unique I1-dynein mutants that assemble partial I1-dynein complexes. Figure 4A shows Western blots of isolated axonemes from three mutant IC97 (M, of ~90,000) is in direct contact with an axonemal protein with an estimated mass of 50 kDa (Figure 3A).

To determine the identity of the IC97-interacting protein, we used the IC97 antiserum to immunoprecipitate the IC97 cross-linked product in sufficient yield for identification (Figure 3B). MS/MS analysis of the IC97 cross-linked product identified peptides corresponding to both α- and β-tubulin in addition to IC97. The identification of IC97 peptides demonstrates that the band isolated for MS/MS is the IC97 cross-linked product. The Mr of the cross-linked product is ~140,000 on SDS-PAGE, suggesting that IC97 does not cross-link to a tubulin dimer, but rather IC97 independently cross-links to α- and β-tubulin. Hence, the cross-linked product is a mixture of IC97-α-tubulin and IC97-β-tubulin species. In support of this model, the ratio of IC97:α-tubulin:β-tubulin peptides recovered is 1.5:1:1, indicating that both α- and β-tubulin are present in equivalent amounts compared with a nearly twofold amount of IC97.

To further validate the results obtained by MS/MS, we used several approaches. First, we immunoprecipitated the IC97 cross-linked product (Figure 3C, IC97) from SDS-solubilized EDC-treated axonemes containing an HA-tagged α1-tubulin (Kozminski et al., 1993). The IC97 cross-linked product is detected using an antibody to the HA tag, indicating that it contains the epitope-tagged tubulin (Figure 3C, HA). Second, we bound purified I1 dynein to Taxol-stabilized microtubules and then performed EDC cross-linking. The IC97 cross-linked product is formed indicating that IC97 interacts with tubulin components in vitro (Figure 3D). Third, we took advantage of a blot overlay assay that uses biotinylated tubulin as a probe to identify tubulin-interacting proteins (Yanagisawa and Kamiya, 2004). Blot overlays of purified dynein fractions detect a tubulin-interacting band at the expected size for IC97 in purified-I1 dynein (dynein-f peak) fractions (asterisk in Figure 3E).
strains: bop5-1—expressing and assembling a truncated IC138 but missing the LC7b and FAP120 subunits of the IC138 subcomplex (Hendrickson et al., 2004; Ikeda et al., 2008), ida7-1::IC140 5A—expressing and assembling a truncated IC140 lacking the N terminus (see table 1 in Perrone et al., 1998), and bop5-2—an IC138-null allele that assembles I1 dynein lacking IC138, LC7b, and FAP120 (Ikeda et al., 2009; Bower et al., 2009) (Figure 4A). These observations are consistent with other reports, indicating that IC97 is a component of the IC138 subcomplex and dependent upon IC138 for assembly into I1 dynein (Ikeda et al., 2008; Bower et al., 2009). This interaction is not mediated by LC7b or FAP120 because IC97 assembles in bop5-1, where LC7b and FAP120 are missing (Hendrickson et al., 2004; Ikeda et al., 2008). Thus, we conclude that IC97 is interacting with IC138 directly. Furthermore, in the bop5-1 mutant, IC97 copurifies with I1-dynein by FPLC fractionation, indicating that it is stably associated with I1 dynein (Hendrickson et al., 2004; our

Figure 2. Antibodies to C,850038 recognize the IC97 subunit of I1 dynein. (A) The IC97 antibody was used to probe Western blots of axonemes derived from wild-type and several dynein mutants. The antibody recognizes a band, with an $M_r$ of ~90,000 that is present in wild type (WT) and dynein mutants that are defective in the outer dynein arm (idd2), or inner arm subtypes a, c, and d (idd4). This band is specifically missing in I1-dynein mutants (idd1, idd3, and idd7) that fail to assemble I1 dynein in the axoneme. The lower panel contains the Coomassie-stained gel showing protein loads. (B) Western blots of FPLC fractions from idd2 dynein extracts were probed with the IC97 antibody (top). The band recognized by the antibody cova- tionates with the I1-dynein complex in the dynein-peak (detected with antibodies to the IC138 subunit; bottom) confirming that the antibody specifically recognizes the IC97 I1-dynein subunit (Porter et al., 1992; Pazour et al., 1999; Yang and Sale, 1998; Kamiya et al., 1991; King and Dutcher, 1997; Myster et al., 1997; Harrison et al., 1998; Bowman et al., 1999; Pazour and Witman, 2000; Perrone et al., 1998, 2000; Dibella et al., 2004b,c; Hendrickson et al., 2004).

Figure 3. IC97 interacts with both $\alpha$- and $\beta$-tubulin. (A) Western blots of wild-type axonemes that were treated with EDC were probed with the IC97 antibody. A prominent cross-linked product of ~140-kDa is formed (arrow; uncross-linked IC97 is marked by the arrowhead). (B) The IC97 antibody was used to immu- noprecipitate the cross-linked product (arrow; uncross-linked IC97 is marked by the arrowhead). The image is a representative silver-stained gel of the immune complexes showing that the IC97 antibody pulls down both uncross-linked IC97 (arrowhead) and the EDC-generated cross-linked product (arrow). The band corre- sponding to the cross-linked product was excised from an identical SYPRO Ruby-stained gel and components identified by tandem mass spectrometry. The cross-linked product represents a mixture of IC97 cross-linked to $\alpha$-tubulin and IC97 cross-linked to $\beta$-tubulin. (C) Western blots of the IC97-immune complexes derived from EDC-treated axonemes containing HA-tagged $\alpha$-tubulin (Kozinski et al., 1993) were probed with the IC97 and HA antibodies. The IC97 antibody detects both uncross-linked (arrowhead) and cross-linked IC97 (arrow). The HA antibody detects only the cross-linked product indicating that it contains IC97 and HA-tagged $\alpha$-tubulin. (D) FPLC-purified I1 dy- nein was bound to Taxol-stabilized microtubules (as- sembled from purified tubulin) and then EDC cross- linked. The 140-kDa IC97-cross-linked product is formed indicating that IC97 cross-links to tubulin in vitro. (E) Blot overlays using biotinylated tubulin (left) reveal a tubulin-interacting band at the expected size for IC97 in FPLC-purified I1 dynein fractions (asterisk). Also evident is tubulin binding to IC140/IC138 (diamond) of I1 dynein and IC1 (circle) of the outer dynein arm (King et al., 1991). The IC2 component of the outer dynein arm is observed in the Coomassie-stained gel (right), but it is not detected in the blot overlay. The left lanes ($\alpha\beta + e$) are purified outer dynein arm fractions; the right lanes (f) are purified I1-dynein fractions.

IC97 is a component of the IC138 subcomplex and dependent upon IC138 for assembly into I1 dynein (Ikeda et al., 2008; Bower et al., 2009). This interaction is not mediated by LC7b or FAP120 because IC97 assembles in bop5-1, where LC7b and FAP120 are missing (Hendrickson et al., 2004; Ikeda et al., 2008). Thus, we conclude that IC97 is interacting with IC138 directly. Furthermore, in the bop5-1 mutant, IC97 copurifies with I1-dynein by FPLC fractionation, indicating that it is stably associated with I1 dynein (Hendrickson et al., 2004; our...
IC97 is not required for I1-dynein assembly or anchoring in the axoneme. (A) Western blots of partial I1-dynein assembly mutants bop5-1, ida7-1::IC140 5A, and bop5-2. IC97 assembles into axonemes of both ida7-1::IC140 5A and bop5-1 but does not localize to the axoneme when IC138, FAP120, and LC7b are missing (bop5-2). Bottom, Coomassie-stained gel of the same samples. (B) FPLC fractions of dynein extracts from a double mutant ida7-1::IC140 5A ada9 demonstrate that IC97, while assembled in the axoneme, does not cofractionate with I1 dynein, but rather is detected in earlier fractions. (C) Western blots of axonemes from a new LC8 mutant, fla14-3 that expresses a larger LC8 protein (top, left; Yang, Yang, Wirschell, and Davis, unpublished data), demonstrates that IC97 and FAP120 fail to assemble in the axoneme (bottom). The rest of the I1-dynein complex seems to assemble in the absence of IC97/FAP120 (represented by IC140; top, right).
IC97 Interacts with I1-Dynein Subunits

In Figure 6A, we propose a model for how IC97 interacts with the axoneme and I1-dynein components. Shown in red are interactions between IC97-tubulin, IC97-IC140, and the IC97-IC138 subcomplex, including FAP120.

Based on the analysis of the bop5-2 mutant (Bower et al., 2009), we conclude that IC97 assembles into I1 dynein in an IC138-dependent manner, suggesting that these two I1-dynein subunits interact. This interaction is not mediated by the C terminus of IC138, LC7b, or FAP120 because IC97 is present in bop5-1, where these proteins are missing (Figure 6C) (Bower et al., 2009). This result suggests that IC97 and IC138 interact directly.

The IC97-IC140 interaction is supported by analysis of the ida7-1::IC140 5A strain (Perrone et al., 1998). Although an intact I1 dynein assembles in this mutant, IC97 is specifically lost upon extraction of I1 dynein from the axoneme (Figure 6B), indicating that IC97 requires the N terminus of IC140 for a stable association with I1 dynein in vitro. No other I1-dynein subunit seems to dissociate in the ida7-1::IC140 5A mutant, suggesting that the IC97-IC140 interaction is also direct.

The model also depicts a possible interaction between IC97 and LC8. Analysis of the fla14-3 mutant indicates that assembly of IC97 and FAP120 are specifically disrupted. However, we predict that the loss of IC97/FAP120 from I1 dynein in fla14-3 is indirect (see below).

IC97 Is Required for I1-Dynein–mediated Control of Microtubule Sliding In Vitro

I1 dynein is an essential part of a regulatory pathway that controls microtubule sliding and regulates axonemal bend-
ing. Figure 6B is founded in part on analysis of microtubule sliding in axonemes isolated from paralyzed flagellar mutants defective in assembly of the radial spokes or central pair structures (reviewed in Porter and Sale 2000; Smith and Yang 2004; Wirschell et al., 2007).

Here, we provide evidence that assembly of IC97 is necessary for regulation of I1 dynein and control of microtubule sliding. fla14-3 axonemes assemble an intact I1-dynein that appears to lack only the IC97 and FAP120 subunits (Figure 4C). This mutant allele of LC8 manifests assembly defects in I1 as well as the radial spokes (Huang et al., 1981; Yang, Yang, Wirschell, and Davis, unpublished data). The defect in radial spoke assembly intrinsic to this mutant allele allowed us to use the microtubule sliding assay to directly test the effects of the loss of IC97 on the regulatory pathway that controls I1 dynein. Similar to radial spoke mutants like pf17; IC138 is hyperphosphorylated in fla14-3 (Figure 5A). Furthermore, despite inhibition of IC138 phosphorylation, fla14-3 axonemes do not increase microtubule sliding rates upon treatment with kinase inhibitors, indicating that IC97 is required, along with IC138, for control of I1 dynein. Evidently, dephosphorylation of IC138 is not sufficient for control of microtubule sliding; the assembly of IC97 also is required (Figure 6B).

We cannot rule out the possibility that the microtubule sliding defect in fla14-3 is a result of the mutation in LC8 and a novel defect in radial spoke assembly that allows fla14-3 to behave differently than other radial spoke mutants. However, we propose that the failure to rescue microtubule sliding velocities in fla14-3 is due to the specific loss of IC97 for the following reasons. 1) Recent models propose that the LC8 dimer functions in a structural capacity; establishing and driving the formation of large macromolecular complexes like the dynein motors (Williams et al., 2007; Barbar, 2008; King, 2008). For example, the IC of cytoplasmic dynein is partially disordered and gains structure upon binding to tubulin (Nyarko et al., 2004). Thus, loss of IC97/FAP120 from fla14-3 I1-dynein may be the result of a defective I1-dynein structure that is not competent to recruit these subunits. Such a model would indicate that IC97/FAP120 and LC8 do not interact directly. 2) More importantly, IC97 requires IC138 for assembly into I1 dynein and together they are localized within a subcomplex at the base of I1-dynein (Bower et al., 2009). IC97 can assemble in the absence of LC7b and FAP120
IC97 Is Related to Proteins Implicated in Pulmonary Carcinoma

IC97 has orthologues in many species that contain motile cilia and flagella. Of interest is the observed homology between IC97 and Las—a protein implicated as a lung tumour susceptibility gene (Zhang et al., 2003). Las1 is highly expressed in lung and testis—tissues that contain motile cilia and in the kidney—a tissue containing immotile, primary cilia (Zhang et al., 2003). Moreover, in dividing cells, Las1 is expressed in G2 and its degradation during mitosis is required for cell cycle progression (Liu et al., 2007). The finding that Las1 is a microtubule-binding protein (Liu et al., 2007) is consistent with the IC97–tubulin interaction described here, suggesting that an interaction with tubulin may be a conserved feature of IC97/Las1 function. The significance of the homology between Chlamydomonas IC97 and the Las1 protein involved in lung tumorigenesis is unclear, but raises the questions of whether Las1 localizes to motile cilia, as well as primary cilia, and whether Las1 plays functional roles both in cilia and outside of the axoneme.

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